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(54) Title: EPITOPE SEQUENCES

(57) Abstract: Disclosed herein are polypeptides, including epitopes, clusters, and antigens. Also disclosed are compositions that include said polypeptides and methods for their use.

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EPITOPE SEQUENCES

Background of the Invention

Field of the Invention

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The present invention generally relates to peptides, and nucleic acids encoding peptides, that are useful epitopes of target-associated antigens. More specifically, the invention relates to epitopes that have a high affinity for MHC class I and that are produced by target-specific proteasomes.

Description of the Related Art

Neoplasia and the Immune System

The neoplastic disease state commonly known as cancer is thought to result generally from a single cell growing out of control. The uncontrolled growth state typically results from a multistep process in which a series of cellular systems fail, resulting in the genesis of a neoplastic cell. The resulting neoplastic cell rapidly reproduces itself, forms one or more tumors, and eventually may cause the death of the host.

Because the progenitor of the neoplastic cell shares the host's genetic material, neoplastic cells are largely unassailed by the host's immune system. During immune surveillance, the process in which the host's immune system surveys and localizes foreign materials, a neoplastic cell will appear to the host's immune surveillance machinery as a "self" cell.

Viruses and the Immune System

In contrast to cancer cells, virus infection involves the expression of clearly non-self antigens. As a result, many virus infections are successfully dealt with by the immune system with minimal clinical sequela. Moreover, it has been possible to develop effective vaccines for many of those infections that do cause serious disease. A variety of vaccine approaches have been used successfully to combat various diseases. These approaches include subunit vaccines consisting of individual proteins produced through recombinant DNA technology. Notwithstanding these advances, the selection and effective administration of minimal epitopes for use as viral vaccines has remained problematic.

In addition to the difficulties involved in epitope selection stands the problem of viruses that have evolved the capability of evading a host's immune system. Many viruses, especially viruses that establish persistent infections, such as members of the herpes and pox virus families, produce immunomodulatory molecules that permit the virus to evade the host's immune system. The effects of these immunomodulatory molecules on antigen presentation may be overcome by the targeting of select epitopes for administration as immunogenic compositions. To better understand the interaction of neoplastic cells and virally infected cells with the host's immune system, a discussion of the system's components follows below.



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The immune system functions to discriminate molecules endogenous to an organism ("self' molecules) from material exogenous or foreign to the organism ("non-self' molecules). The immune system has two types of adaptive responses to foreign bodies based on the components that mediate the response: a humoral response and a cell-mediated response. The humoral response is mediated by antibodies, while the cell-mediated response involves cells classified as lymphocytes. Recent anticancer and antiviral strategies have focused on mobilizing the host immune system as a means of anticancer or antiviral treatment or therapy.

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The immune system functions in three phases to protect the host from foreign bodies: the cognitive phase, the activation phase, and the effector phase. In the cognitive phase, the immune system recognizes and signals the presence of a foreign antigen or invader in the body. The foreign antigen can be, for example, a cell surface marker from a neoplastic cell or a viral protein. Once the system is aware of an invading body, antigen specific cells of the immune system proliferate and differentiate in response to the invader-triggered signals. The last stage is the effector stage in which the effector cells of the immune system respond to and neutralize the detected invader.

An array of effector cells implements an immune response to an invader. One type of effector cell, the B cell, generates antibodies targeted against foreign antigens encountered by the host. In combination with the complement system, antibodies direct the destruction of cells or organisms bearing the targeted antigen. Another type of effector cell is the natural killer cell (NK cell), a type of lymphocyte having the capacity to spontaneously recognize and destroy a variety of virus infected cells as well as malignant cell types. The method used by NK cells to recognize target cells is poorly understood.

Another type of effector cell, the T cell, has members classified into three subcategories, each playing a different role in the immune response. Helper T cells secrete cytokines which stimulate the proliferation of other cells necessary for mounting an effective immune response, while suppressor T cells down-regulate the immune response. A third category of T cell, the cytotoxic T cell (CTL), is capable of directly lysing a targeted cell presenting a foreign antigen on its surface.

The Major Histocompatibility Complex and T Cell Target Recognition

T cells are antigen-specific immune cells that function in response to specific antigen signals. B lymphocytes and the antibodies they produce are also antigen-specific entities. However, unlike B lymphocytes, T cells do not respond to antigens in a free or soluble form. For a T cell to respond to an antigen, it requires the antigen to be processed to peptides which are then bound to a presenting structure encoded in the major histocompatibility complex (MHC). This requirement is called "MHC restriction" and it is the mechanism by which T cells differentiate "self" from "non-self" cells. If an antigen is not displayed by a recognizable MHC molecule, the T cell will not recognize and act on the antigen signal. T cells specific for a peptide bound to a



recognizable MHC molecule bind to these MHC-peptide complexes and proceed to the next stages of the immune response.

There are two types of MHC, class I MHC and class II MHC. T Helper cells (CD4⁺) predominately interact with class II MHC proteins while cytolytic T cells (CD8⁺) predominately interact with class I MHC proteins. Both classes of MHC protein are transmembrane proteins with a majority of their structure on the external surface of the cell. Additionally, both classes of MHC proteins have a peptide binding cleft on their external portions. It is in this cleft that small fragments of proteins, endogenous or foreign, are bound and presented to the extracellular environment.

Cells called "professional antigen presenting cells" (pAPCs) display antigens to T cells using the MHC proteins but additionally express various co-stimulatory molecules depending on the particular state of differentiation/activation of the pAPC. When T cells, specific for the peptide bound to a recognizable MHC protein, bind to these MHC-peptide complexes on pAPCs, the specific co-stimulatory molecules that act upon the T cell direct the path of differentiation/activation taken by the T cell. That is, the co-stimulation molecules affect how the T cell will act on antigenic signals in future encounters as it proceeds to the next stages of the immune response.

As discussed above, neoplastic cells are largely ignored by the immune system. A great deal of effort is now being expended in an attempt to harness a host's immune system to aid in combating the presence of neoplastic cells in a host. One such area of research involves the formulation of anticancer vaccines.

Anticancer Vaccines

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Among the various weapons available to an oncologist in the battle against cancer is the immune system of the patient. Work has been done in various attempts to cause the immune system to combat cancer or neoplastic diseases. Unfortunately, the results to date have been largely disappointing. One area of particular interest involves the generation and use of anticancer vaccines.

To generate a vaccine or other immunogenic composition, it is necessary to introduce to a subject an antigen or epitope against which an immune response may be mounted. Although neoplastic cells are derived from and therefore are substantially identical to normal cells on a genetic level, many neoplastic cells are known to present tumor-associated antigens (TuAAs). In theory, these antigens could be used by a subject's immune system to recognize these antigens and attack the neoplastic cells. In reality, however, neoplastic cells generally appear to be ignored by the host's immune system.

A number of different strategies have been developed in an attempt to generate vaccines with activity against neoplastic cells. These strategies include the use of tumor-associated antigens



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as immunogens. For example, U.S. Patent No. 5,993,828, describes a method for producing an immune response against a particular subunit of the Urinary Tumor Associated Antigen by administering to a subject an effective dose of a composition comprising inactivated tumor cells having the Urinary Tumor Associated Antigen on the cell surface and at least one tumor associated antigen selected from the group consisting of GM-2, GD-2, Fetal Antigen and Melanoma Associated Antigen. Accordingly, this patent describes using whole, inactivated tumor cells as the immunogen in an anticancer vaccine.

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Another strategy used with anticancer vaccines involves administering a composition containing isolated tumor antigens. In one approach, MAGE-A1 antigenic peptides were used as an immunogen. (See Chaux, P., et al., "Identification of Five MAGE-A1 Epitopes Recognized by Cytolytic T Lymphocytes Obtained by *In Vitro* Stimulation with Dendritic Cells Transduced with MAGE-A1," J. Immunol., 163(5):2928-2936 (1999)). There have been several therapeutic trials using MAGE-A1 peptides for vaccination, although the effectiveness of the vaccination regimes was limited. The results of some of these trials are discussed in Vose, J.M., "Tumor Antigens Recognized by T Lymphocytes," 10th European Cancer Conference, Day 2, Sept. 14, 1999.

In another example of tumor associated antigens used as vaccines, Scheinberg, et al. treated 12 chronic myelogenous leukemia (CML) patients already receiving interferon (IFN) or hydroxyurea with 5 injections of class I-associated bcr-abl peptides with a helper peptide plus the adjuvant QS-21. Scheinberg, D.A., et al., "BCR-ABL Breakpoint Derived Oncogene Fusion Peptide Vaccines Generate Specific Immune Responses in Patients with Chronic Myelogenous Leukemia (CML) [Abstract 1665], American Society of Clinical Oncology 35th Annual Meeting, Atlanta (1999). Proliferative and delayed type hypersensitivity (DTH) T cell responses indicative of T-helper activity were elicited, but no cytolytic killer T cell activity was observed within the fresh blood samples.

Additional examples of attempts to identify TuAAs for use as vaccines are seen in the recent work of Cebon, et al. and Scheibenbogen, et al. Cebon, et al. immunized patients with metastatic melanoma using intradermally administered MART-1₂₆₋₃₅ peptide with IL-12 in increasing doses given either subcutaneously or intravenously. Of the first 15 patients, 1 complete remission, 1 partial remission, and 1 mixed response were noted. Immune assays for T cell generation included DTH, which was seen in patients with or without IL-12. Positive CTL assays were seen in patients with evidence of clinical benefit, but not in patients without tumor regression. Cebon, et al., "Phase I Studies of Immunization with Melan-A and IL-12 in HLA A2+ Positive Patients with Stage III and IV Malignant Melanoma," [Abstract 1671], American Society of Clinical Oncology 35th Annual Meeting, Atlanta (1999).

Scheibenbogen, et al. immunized 18 patients with 4 HLA class I restricted tyrosinase peptides, 16 with metastatic melanoma and 2 adjuvant patients. Scheibenbogen, et al.,

"Vaccination with Tyrosinase peptides and GM-CSF in Metastatic Melanoma: a Phase II Trial," [Abstract 1680], American Society of Clinical Oncology 35th Annual Meeting, Atlanta (1999). Increased CTL activity was observed in 4/15 patients, 2 adjuvant patients, and 2 patients with evidence of tumor regression. As in the trial by Cebon, *et al.*, patients with progressive disease did not show boosted immunity. In spite of the various efforts expended to date to generate efficacious anticancer vaccines, no such composition has yet been developed.

Antiviral Vaccines

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Vaccine strategies to protect against viral diseases have had many successes. Perhaps the most notable of these is the progress that has been made against the disease small pox, which has been driven to extinction. The success of the polio vaccine is of a similar magnitude.

Viral vaccines can be grouped into three classifications: live attenuated virus vaccines, such as vaccinia for small pox, the Sabin poliovirus vaccine, and measles mumps and rubella; whole killed or inactivated virus vaccines, such as the Salk poliovirus vaccine, hepatitis A virus vaccine and the typical influenza virus vaccines; and subunit vaccines, such as hepatitis B. Due to their lack of a complete viral genome, subunit vaccines offer a greater degree of safety than those based on whole viruses.

The paradigm of a successful subunit vaccine is the recombinant hepatitis B vaccine based on the viruses envelope protein. Despite much academic interest in pushing the reductionist subunit concept beyond single proteins to individual epitopes, the efforts have yet to bear much fruit. Viral vaccine research has also concentrated on the induction of an antibody response although cellular responses also occur. However, many of the subunit formulations are particularly poor at generating a CTL response.

Summary of the Invention

Previous methods of priming professional antigen presenting cells (pAPCs) to display target cell epitopes have relied simply on causing the pAPCs to express target-associated antigens (TAAs), or epitopes of those antigens which are thought to have a high affinity for MHC I molecules. However, the proteasomal processing of such antigens results in presentation of epitopes on the pAPC that do not correspond to the epitopes present on the target cells.

Using the knowledge that an effective cellular immune response requires that pAPCs present the same epitope that is presented by the target cells, the present invention provides epitopes that have a high affinity for MHC I, and that correspond to the processing specificity of the housekeeping proteasome, which is active in peripheral cells. These epitopes thus correspond to those presented on target cells. The use of such epitopes in compositions, such as vaccines and other immunogenic compositions (including pharmaceutical and immunotherapeutic compositions) can activate the cellular immune response to recognize the correctly processed TAA and can result in removal of target cells that present such epitopes. In some embodiments, the housekeeping

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epitopes provided herein can be used in combination with immune epitopes, generating a cellular immune response that is competent to attack target cells both before and after interferon induction. In other embodiments the epitopes are useful in the diagnosis and monitoring of the target-associated disease and in the generation of immunological reagents for such purposes.

Embodiments of the invention relate to isolated epitopes, antigens and/or polypeptides. The isolated antigens and/or polypeptides can include the epitopes. Preferred embodiments include an epitope or antigen having the sequence as disclosed in Tables 1A or 1B. Other embodiments can include an epitope cluster comprising a polypeptide from Tables 1A or 1B. Further, embodiments include a polypeptide having substantial similarity to the already mentioned epitopes, polypeptides, antigens, or clusters. Other preferred embodiments include a polypeptide having functional similarity to any of the above. Still further embodiments relate to a nucleic acid encoding the polypeptide of any of the epitopes, clusters, antigens, and polypeptides from Tables 1A or 1B and mentioned herein.

For purposes of the following summary and discussion of other embodiments of the invention, reference to "the epitope," "the epitopes," or "epitope from Tables 1A or 1B" may include without limitation to all of the foregoing forms of the epitope including an epitope with the sequence set forth in the Tables or elsewhere herein, a cluster comprising such an epitope or epitopes, a polypeptide having substantial or functional similarity to those epitopes or clusters, and the like.

The polypeptide or epitope can be immunologically active. The polypeptide comprising the epitope can be less than about 30 amino acids in length, more preferably, the polypeptide is 8 to 10 amino acids in length, for example. Substantial or functional similarity can include addition of at least one amino acid, for example, and the at least one additional amino acid can be at an N-terminus of the polypeptide. The substantial or functional similarity can include a substitution of at least one amino acid.

The epitope, cluster, or polypeptide comprising the same can have affinity to an HLA-A2 molecule. The affinity can be determined by an assay of binding, by an assay of restriction of epitope recognition, by a prediction algorithm, and the like. The epitope, cluster, or polypeptide comprising the same can have affinity to an HLA-B7, HLA-B51 molecule, and the like.

In preferred embodiments the polypeptide can be a housekeeping epitope. The epitope or polypeptide can correspond to an epitope displayed on a tumor cell, to an epitope displayed on a neovasculature cell, and the like. The epitope or polypeptide can be an immune epitope. The epitope, cluster and/or polypeptide can be a nucleic acid. The epitope, cluster and/or polypeptide can be encoded by a nucleic acid.

Other embodiments relate to compositions, including pharmaceutical or immunogenic compositions comprising the polypeptides, including an epitope from Tables 1A or 1B, a cluster, or

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a polypeptide comprising the same, and a pharmaceutically acceptable adjuvant, carrier, diluent, excipient, and the like. The adjuvant can be a polynucleotide. The polynucleotide can include a dinucleotide, which can be CpG, for example. The adjuvant can be encoded by a polynucleotide. The adjuvant can be a cytokine and the cytokine can be, for example, GM-CSF.

The compositions can further include a professional antigen-presenting cell (pAPC). The pAPC can be a dendritic cell, for example. The composition can further include a second epitope. The second epitope can be a polypeptide, a nucleic acid, a housekeeping epitope, an immune epitope, and the like.

Still further embodiments relate to compositions, including pharmaceutical and immunogenic compositions that include any of the nucleic acids discussed herein, including those that encode polypeptides that comprise epitopes or antigens from Tables 1A or 1B. Such compositions can include a pharmaceutically acceptable adjuvant, carrier, diluent, excipient, and the like.

Other embodiments relate to recombinant constructs that include such a nucleic acid as described herein, including those that encode polypeptides that comprise epitopes or antigens from Tables 1A or 1B. The constructs can further include a plasmid, a viral vector, an artificial chromosome, and the like. The construct can further include a sequence encoding at least one feature, such as for example, a second epitope, an IRES, an ISS, an NIS, a ubiquitin, and the like.

Further embodiments relate to purified antibodies that specifically bind to at least one of the epitopes in Tables 1A or 1B. Other embodiments relate to purified antibodies that specifically bind to a peptide-MHC protein complex comprising an epitope disclosed in Tables 1A or 1B or any other suitable epitope. The antibody from any embodiment can be a monoclonal antibody or a polyclonal antibody.

Still other embodiments relate to multimeric MHC-peptide complexes that include an epitope, such as, for example, an epitope disclosed in Tables 1A or 1B. Also, contemplated are antibodies specific for the complexes.

Embodiments relate to isolated T cells expressing a T cell receptor specific for an MHC-peptide complex. The complex can include an epitope, such as, for example, an epitope disclosed in Tables 1A or 1B. The T cell can be produced by an *in vitro* immunization and can be isolated from an immunized animal. Embodiments relate to T cell clones, including cloned T cells, such as those discussed above. Embodiments also relate to polyclonal population of T cells. Such populations can include a T cell, as described above, for example.

Still further embodiments relate to compositions, including pharmaceutical and immunogenic compositions that include a T cell, such as those described above, for example, and a pharmaceutically acceptable adjuvant, carrier, diluent, excipient, and the like.

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Embodiments of the invention relate to isolated protein molecules comprising the binding domain of a T cell receptor specific for an MHC-peptide complex. The complex can include an epitope as disclosed in Tables 1A or 1B. The protein can be multivalent. Other embodiments relate to isolated nucleic acids encoding such proteins. Still further embodiments relate to recombinant constructs that include such nucleic acids.

Other embodiments of the invention relate to host cells expressing a recombinant construct as described above and elsewhere herein. The host cells can include constructs encoding an epitope, a cluster or a polypeptide comprising said epitope or said cluster. The epitope or epitope cluster can be one or more of those disclosed in Tables 1A or 1B, for example, and as otherwise defined. The host cell can be a dendritic cell, macrophage, tumor cell, tumor-derived cell, a bacterium, fungus, protozoan, and the like. Embodiments also relate to compositions, including pharmaceutical and immunogenic compositions that include a host cell, such as those discussed herein, and a pharmaceutically acceptable adjuvant, carrier, diluent, excipient, and the like.

Still other embodiments relate to compositions including immunogenic compositions, such as for example, vaccines or immunotherapeutic compositions. The compositions can include at least one component, such as, for example, an epitope disclosed in Tables 1A or 1B or otherwise described herein; a cluster that includes such an epitope, an antigen or polypeptide that includes such an epitope; a composition as described above and herein; a construct as described above and herein, a T cell, a construct comprising a nucleic acid encoding a T cell receptor binding domain specific for an MHC-peptide complex and compositions including the same, a host cell as described above and herein, and compositions comprising the same.

Further embodiments relate to methods of treating an animal. The methods can include administering to an animal a composition, including a pharmaceutical or an immunogenic composition, such as, a vaccine or immunotherapeutic composition, including those disclosed above and herein. The administering step can include a mode of delivery, such as, for example, transdermal, intranodal, perinodal, oral, intravenous, intradermal, intramuscular, intraperitoneal, mucosal, aerosol inhalation, instillation, and the like. The method can further include a step of assaying to determine a characteristic indicative of a state of a target cell or target cells. The method can include a first assaying step and a second assaying step, wherein the first assaying step precedes the administering step, and wherein the second assaying step follows the administering step. The method can further include a step of comparing the characteristic determined in the first assaying step with the characteristic determined in the second assaying step to obtain a result. The result can be for example, evidence of an immune response, a diminution in number of target cells, a loss of mass or size of a tumor comprising target cells, a decrease in number or concentration of an intracellular parasite infecting target cells, and the like.

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Embodiments relate to methods of evaluating immunogenicity of a composition, including a vaccine or an immunotherapeutic composition. The methods can include administering to an animal a vaccine or immunotherapeutic, such as those described above and elsewhere herein, and evaluating immunogenicity based on a characteristic of the animal. The animal can be MHC-transgenic.

Other embodiments relate to methods of evaluating immunogenicity that include *in vitro* stimulation of a T cell with the vaccine or immunotherapeutic composition, such as those described above and elsewhere herein, and evaluating immunogenicity based on a characteristic of the T cell. The stimulation can be a primary stimulation.

Still further embodiments relate to methods of making a passive/adoptive immunotherapeutic. The methods can include combining a T cell or a host cell, such as those described above and elsewhere herein, with a pharmaceutically acceptable adjuvant, carrier, diluent, excipient, and the like.

Other embodiments relate to methods of determining specific T cell frequency, and can include the step of contacting T cells with a MHC-peptide complex comprising an epitope disclosed in Tables 1A or 1B, or a complex comprising a cluster or antigen comprising such an epitope. The contacting step can include at least one feature, such as, for example, immunization, restimulation, detection, enumeration, and the like. The method can further include ELISPOT analysis, limiting dilution analysis, flow cytometry, in situ hybridization, the polymerase chain reaction, any combination thereof, and the like.

Embodiments relate to methods of evaluating immunologic response. The methods can include the above-described methods of determining specific T cell frequency carried out prior to and subsequent to an immunization step.

Other embodiments relate to methods of evaluating immunologic response. The methods can include determining frequency, cytokine production, or cytolytic activity of T cells, prior to and subsequent to a step of stimulation with MHC-peptide complexes comprising an epitope, such as, for example an epitope from Tables 1A or 1B, a cluster or a polypeptide comprising such an epitope.

Further embodiments relate to methods of diagnosing a disease. The methods can include contacting a subject tissue with at least one component, including, for example, a T cell, a host cell, an antibody, a protein, including those described above and elsewhere herein; and diagnosing the disease based on a characteristic of the tissue or of the component. The contacting step can take place *in vivo* or *in vitro*, for example.

Still other embodiments relate to methods of making a composition, including for example, a vaccine. The methods can include combining at least one component. For example, the component can be an epitope, a composition, a construct, a T cell, a host cell; including any of

those described above and elsewhere herein, and the like, with a pharmaceutically acceptable adjuvant, carrier, diluent, excipient, and the like.

Embodiments relate to computer readable media having recorded thereon the sequence of any one of SEQ ID NOS: 108-610, in a machine having a hardware or software that calculates the physical, biochemical, immunologic, molecular genetic properties of a molecule embodying said sequence, and the like.

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Still other embodiments relate to methods of treating an animal. The methods can include combining the method of treating an animal that includes administering to the animal a vaccine or immunotherapeutic composition, such as described above and elsewhere herein, combined with at least one mode of treatment, including, for example, radiation therapy, chemotherapy, biochemotherapy, surgery, and the like.

Further embodiments relate to isolated polypeptides that include an epitope cluster. In preferred embodiments the cluster can be from a target-associated antigen having the sequence as disclosed in any one of Tables 68-73, wherein the amino acid sequence includes not more than about 80% of the amino acid sequence of the antigen.

Other embodiments relate to immunogenic compositions, including vaccines or immunotherapeutic products that include an isolated peptide as described above and elsewhere herein. Still other embodiments relate to isolated polynucleotides encoding a polypeptide as described above and elsewhere herein. Other embodiments relate vaccines or immunotherapeutic products that include these polynucleotides. The polynucleotide can be DNA, RNA, and the like.

Still further embodiments relate to kits comprising a delivery device and any of the embodiments mentioned above and elsewhere herein. The delivery device can be a catheter, a syringe, an internal or external pump, a reservoir, an inhaler, microinjector, a patch, and any other like device suitable for any route of delivery. As mentioned, the kit, in addition to the delivery device also includes any of the embodiments disclosed herein. For example, without limitations, the kit can include an isolated epitope, a polypeptide, a cluster, a nucleic acid, an antigen, a pharmaceutical composition that includes any of the foregoing, an antibody, a T cell, a T cell receptor, an epitope-MHC complex, a vaccine, an immunotherapeutic, and the like. The kit can also include items such as detailed instructions for use and any other like item.

Brief Description of the Drawings

Figure 1A-1C is a sequence alignment of NY-ESO-1 and several similar protein sequences. Figure 2 graphically represents a plasmid vaccine backbone useful for delivering nucleic acid-encoded epitopes.

Figures 3A and 3B are FACS profiles showing results of HLA-A2 binding assays for tyrosinase₂₀₇₋₂₁₅ and tyrosinase₂₀₈₋₂₁₆.

Figure 3C shows cytolytic activity against a tyrosinase epitope by human CTL induced by *in vitro* immunization.

Figure 4 is a T=120 min. time point mass spectrum of the fragments produced by proteasomal cleavage of SSX-2₃₁₋₆₈.

Figure 5 shows a binding curve for HLA-A2:SSX-241-49 with controls.

Figure 6 shows specific lysis of SSX-2₄₁₋₄₉-pulsed targets by CTL from SSX-2₄₁₋₄₉-immunized HLA-A2 transgenic mice.

Figure 7A, B, and C show results of N-terminal pool sequencing of a T=60 min. time point aliquot of the PSMA₁₆₃₋₁₉₂ proteasomal digest.

Figure 8 shows binding curves for HLA-A2:PSMA₁₆₈₋₁₇₇ and HLA-A2:PSMA₂₈₈₋₂₉₇ with controls.

Figure 9 shows results of N-terminal pool sequencing of a T=60 min. time point aliquot of the PSMA₂₈₁₋₃₁₀ proteasomal digest.

Figure 10 shows binding curves for HLA-A2:PSMA₄₆₁₋₄₆₉, HLA-A2:PSMA₄₆₀₋₄₆₉, and HLA-A2:PSMA₆₆₃₋₆₇₁, with controls.

Figure 11 shows the results of a γ (gamma)-IFN-based ELISPOT assay detecting PSMA₄₆₃₋₄₇₁-reactive HLA-A1⁺ CD8⁺ T cells.

Figure 12 shows blocking of reactivity of the T cells used in figure 10 by anti-HLA-A1 mAb, demonstrating HLA-A1-restricted recognition.

Figure 13 shows a binding curve for HLA-A2:PSMA₆₆₃₋₆₇₁, with controls.

Figure 14 shows a binding curve for HLA-A2:PSMA₆₆₂₋₆₇₁, with controls.

Figure 15. Comparison of anti-peptide CTL responses following immunization with various doses of DNA by different routes of injection.

Figure 16. Growth of transplanted gp33 expressing tumor in mice immunized by i.ln. injection of gp33 epitope-expressing, or control, plasmid.

Figure 17. Amount of plasmid DNA detected by real-time PCR in injected or draining lymph nodes at various times after i.ln. of i.m. injection, respectively.

Figures 18-70 are proteasomal digestion maps depicting the mapping of mass spectrum peaks from the digest onto the sequence of the indicated substrate.

Detailed Description of the Preferred Embodiment

Definitions

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Unless otherwise clear from the context of the use of a term herein, the following listed terms shall generally have the indicated meanings for purposes of this description.

PROFESSIONAL ANTIGEN-PRESENTING CELL (pAPC) – a cell that possesses T cell costimulatory molecules and is able to induce a T cell response. Well characterized pAPCs include dendritic cells, B cells, and macrophages.

PERIPHERAL CELL – a cell that is not a pAPC.

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HOUSEKEEPING PROTEASOME – a proteasome normally active in peripheral cells, and generally not present or not strongly active in pAPCs.

IMMUNE PROTEASOME – a proteasome normally active in pAPCs; the immune proteasome is also active in some peripheral cells in infected tissues.

EPITOPE – a molecule or substance capable of stimulating an immune response. In preferred embodiments, epitopes according to this definition include but are not necessarily limited to a polypeptide and a nucleic acid encoding a polypeptide, wherein the polypeptide is capable of stimulating an immune response. In other preferred embodiments, epitopes according to this definition include but are not necessarily limited to peptides presented on the surface of cells, the peptides being non-covalently bound to the binding cleft of class I MHC, such that they can interact with T cell receptors (TCR). Epitopes presented by class I MHC may be in immature or mature form. "Mature" refers to an MHC epitope in distinction to any precursor ("immature") that may include or consist essentially of a housekeeping epitope, but also includes other sequences in a primary translation product that are removed by processing, including without limitation, alone or in any combination proteasomal digestion, N-terminal trimming, or the action of exogenous enzymatic activities. Thus, a mature epitope may be provided embedded in a somewhat longer polypeptide, the immunological potential of which is due, at least in part, to the embedded epitope; or in its ultimate form that can bind in the MHC binding cleft to be recognized by TCR, respectively.

MHC EPITOPE – a polypeptide having a known or predicted binding affinity for a mammalian class I or class II major histocompatibility complex (MHC) molecule.

HOUSEKEEPING EPITOPE – In a preferred embodiment, a housekeeping epitope is defined as a polypeptide fragment that is an MHC epitope, and that is displayed on a cell in which housekeeping proteasomes are predominantly active. In another preferred embodiment, a housekeeping epitope is defined as a polypeptide containing a housekeeping epitope according to the foregoing definition, that is flanked by one to several additional amino acids. In another preferred embodiment, a housekeeping epitope is defined as a nucleic acid that encodes a housekeeping epitope according to the foregoing definitions.

IMMUNE EPITOPE – In a preferred embodiment, an immune epitope is defined as a polypeptide fragment that is an MHC epitope, and that is displayed on a cell in which immune proteasomes are predominantly active. In another preferred embodiment, an immune epitope is defined as a polypeptide containing an immune epitope according to the foregoing definition, that is flanked by one to several additional amino acids. In another preferred embodiment, an immune epitope is defined as a polypeptide including an epitope cluster sequence, having at least two polypeptide sequences having a known or predicted affinity for a class I MHC. In yet another

preferred embodiment, an immune epitope is defined as a nucleic acid that encodes an immune epitope according to any of the foregoing definitions.

TARGET CELL – a cell to be targeted by the vaccines and methods of the invention. Examples of target cells according to this definition include but are not necessarily limited to: a neoplastic cell and a cell harboring an intracellular parasite, such as, for example, a virus, a bacterium, or a protozoan.

TARGET-ASSOCIATED ANTIGEN (TAA) – a protein or polypeptide present in a target cell.

TUMOR-ASSOCIATED ANTIGENS (TuAA) – a TAA, wherein the target cell is a neoplastic cell.

HLA EPITOPE – a polypeptide having a known or predicted binding affinity for a human class I or class II HLA complex molecule.

ANTIBODY – a natural immunoglobulin (Ig), poly- or monoclonal, or any molecule composed in whole or in part of an Ig binding domain, whether derived biochemically or by use of recombinant DNA. Examples include *inter alia*, F(ab), single chain Fv, and Ig variable region-phage coat protein fusions.

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ENCODE – an open-ended term such that a nucleic acid encoding a particular amino acid sequence can consist of codons specifying that (poly)peptide, but can also comprise additional sequences either translatable, or for the control of transcription, translation, or replication, or to facilitate manipulation of some host nucleic acid construct.

SUBSTANTIAL SIMILARITY – this term is used to refer to sequences that differ from a reference sequence in an inconsequential way as judged by examination of the sequence. Nucleic acid sequences encoding the same amino acid sequence are substantially similar despite differences in degenerate positions or modest differences in length or composition of any non-coding regions. Amino acid sequences differing only by conservative substitution or minor length variations are substantially similar. Additionally, amino acid sequences comprising housekeeping epitopes that differ in the number of N-terminal flanking residues, or immune epitopes and epitope clusters that differ in the number of flanking residues at either terminus, are substantially similar. Nucleic acids that encode substantially similar amino acid sequences are themselves also substantially similar.

FUNCTIONAL SIMILARITY – this term is used to refer to sequences that differ from a reference sequence in an inconsequential way as judged by examination of a biological or biochemical property, although the sequences may not be substantially similar. For example, two nucleic acids can be useful as hybridization probes for the same sequence but encode differing amino acid sequences. Two peptides that induce cross-reactive CTL responses are functionally similar even if they differ by non-conservative amino acid substitutions (and thus do not meet the substantial similarity definition). Pairs of antibodies, or TCRs, that recognize the same epitope can

be functionally similar to each other despite whatever structural differences exist. In testing for functional similarity of immunogenicity one would generally immunize with the "altered" antigen and test the ability of the elicited response (Ab, CTL, cytokine production, etc.) to recognize the target antigen. Accordingly, two sequences may be designed to differ in certain respects while retaining the same function. Such designed sequence variants are among the embodiments of the present invention.

VACCINE – this term is used to refer to those immunogenic compositions that are capable of eliciting prophylactic and/or therapeutic responses that prevent, cure, or ameliorate disease.

IMMUNOGENIC COMPOSITION - this term is used to refer to compositions capable of inducing an immune response, a reaction, an effect, and/or an event. In some embodiments, such responses, reactions, effects, and/or events can be induced *in vitro* or *in vivo*, for example. Included among these embodiments are the induction, activation, or expansion of cells involved in cell mediated immunity, for example. One example of such cells is cytotoxic T lymphocytes (CTLs). A vaccine is one type of immunogenic composition. Another example of such a composition is one that induces, activates, or expands CTLs *in vitro*. Further examples include pharmaceutical compositions and the like.

Table 1A. SEQ ID NOS.* including epitopes in Examples 1-7, 13, 14.

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SEQ ID NO	IDENTITY	SEQUENCE
1	Tyr 207-216	FLPWHRLFLL
2	Tyrosinase protein	Accession number**: P14679
3	SSX-2 protein	Accession number: NP 003138
4	PSMA protein	Accession number: NP 004467
5	Tyrosinase cDNA	Accession number: NM 000372
6	SSX-2 cDNA	Accession number: NM 003147
7	PSMA cDNA	Accession number: NM 004476
8	Tyr 207-215	FLPWHRLFL
9	Туг 208-216	LPWHRLFLL
10		YFSKEEWEKMKASEKIFYVYMKRKYEAMTKLGF
	SSX-2 31-68	KATLP
11	SSX-2 32-40	FSKEEWEKM
12	SSX-2 39-47	KMKASEKIF
13	SSX-2 40-48	MKASEKIFY
14	SSX-2 39-48	KMKASEKIFY
15	SSX-2 41-49	KASEKIFYV
16	SSX-2 40-49	MKASEKIFYV
17	SSX-2 41-50	KASEKIFYVY
18	SSX-2 42-49	ASEKIFYVY
19	SSX-2 53-61	RKYEAMTKL
20	SSX-2 52-61	KRKYEAMTKL
21	SSX-2 54-63	KYEAMTKLGF
22	SSX-2 55-63	YEAMTKLGF
23	SSX-2 56-63	EAMTKLGF

SEQ ID NO	IDENTITY	SEQUENCE
24	HBV18-27	FLPSDYFPSV
25	HLA-B44 binder	AEMGKYSFY
26	SSX-1 41-49	KYSEKISYV
27	SSX-3 41-49	KVSEKIVYV
28	SSX-4 41-49	KSSEKIVYV
29	SSX-5 41-49	KASEKIIYV
30	PSMA163-192	AFSPQGMPEGDLVYVNYARTEDFFKLERDM
31	PSMA 168-190	GMPEGDLVYVNYARTEDFFKLER
32	PSMA 169-177	MPEGDLVYV
33	PSMA 168-177	GMPEGDLVYV
34	PSMA 168-176	GMPEGDLVY
35	PSMA 167-176	QGMPEGDLVY
36	PSMA 169-176	MPEGDLVY
37	PSMA 171-179	EGDLVYVNY
38	PSMA 170-179	PEGDLVYVNY
39	PSMA 174-183	LVYVNYARTE
40	PSMA 177-185	VNYARTEDF
41	PSMA 176-185	YVNYARTEDF
42	PSMA 178-186	NYARTEDFF
43	PSMA 179-186	YARTEDFF
44	PSMA 181-189	RTEDFFKLE
45	PSMA 281-310	RGIAEAVGLPSIPVHPIGYYDAQKLLEKMG
46	PSMA 283-307	IAEAVGLPSIPVHPIGYYDAQKLLE
47	PSMA 289-297	LPSIPVHPI
48	PSMA 288-297	GLPSIPVHPI
49	PSMA 297-305	IGYYDAQKL
50	PSMA 296-305	PIGYYDAQKL
51	PSMA 291-299	SPVHPIGY
52	PSMA 290-299	PSIPVHPIGY
53	PSMA 292-299	IPVHPIGY
54	PSMA 299-307	YYDAQKLLE
55	PSMA454-481	SSIEGNYTLRVDCTPLMYSLVHLTKEL
56	PSMA 456-464	IEGNYTLRV
57	PSMA 455-464	SIEGNYTLRV
58	PSMA 457-464	EGNYTLRV
59	PSMA 461-469	TLRVDCTPL
60	PSMA 460-469	YTLRVDCTPL
61	PSMA 462-470	LRVDCTPLM
62	PSMA 463-471	RVDCTPLMY
63	PSMA 462-471	LRVDCTPLMY
64	PSMA653-687	FDKSNPIVLRMMNDQLMFLERAFIDPLGLPDRPFY
65	PSMA 660-681	VLRMMNDQLMFLERAFIDPLGL
66	PSMA 663-671	MMNDQLMFL
67	PSMA 662-671	RMMNDQLMFL
68	PSMA 662-670	RMMNDQLMF
69	Tyr 1-17	MLLAVLYCLLWSFQTSA
70	GP100 protein ²	Accession number: P40967
71	MAGE-1 protein	Accession number: P43355
72	MAGE-2 protein	Accession number: P43356

SEQ ID NO	IDENTITY	SEQUENCE
73	MAGE-3 protein	Accession number: P43357
74	NY-ESO-1 protein	Accession number: P78358
75	LAGE-1a protein	Accession number: CAA11116
76	LAGE-1b protein	Accession number: CAA11117
77	PRAME protein	Accession number: NP 006106
78	PSA protein	Accession number: P07288
79	PSCA protein	Accession number: O43653
80	GP100 cds	Accession number: U20093
81	MAGE-1 cds	Accession number: M77481
. 82	MAGE-2 cds	Accession number: L18920
83	MAGE-3 cds	Accession number: U03735
84	NY-ESO-1 cDNA	Accession number: U87459
85	PRAME cDNA	Accession number: NM 006115
86	PSA cDNA	Accession number: NM 001648
87	PSCA cDNA	Accession number: AF043498
88	CEA protein	Accession number: P06731
. 89	CEA cDNA	Accession number: NM 004363
90	Her2/Neu protein	Accession number: P04626
91	Her2/Neu cDNA	Accession number: M11730
92	SCP-1 protein	Accession number: Q15431
93	SCP-1 cDNA	Accession number: X95654
94	SSX-4 protein	Accession number: O60224
95	SSX-4 cDNA	Accession number: NM_005636
96	GAGE-1 protein	Accession number: Q13065
97	GAGE-1 cDNA	Accession number: U19142
98	Suvivin protein	Accession number: O15392
99	Survivin cDNA	Accession number: NM_001168
100	Melan-A protein	Accession number: Q16655
101	Melan-A cDNA	Accession number: U06452
102	BAGE protein	Accession number: Q13072
103	BAGE cDNA	Accession number: U19180
104	PSA 59-67	WVLTAAHCI
105	Glandular	Accession number: P06870
106	Kallikrein 1	
106	Elastase 2A	Accession number: P08217
107	Pancreatic elastase IIB	Accession number: NP_056933
	ш	

Table 1B. SEQ ID NOS.* including epitopes in Examples 15-67.

SEQ ID NO	IDENTITY	SEQUENCE
108	Tyr 171-179	NIYDLFVWM
109	Tyr 173-182	YDLFVWMHYY
110	Tyr 174-182	DLFVWMHYY
111	Tyr 186-194	DALLGGSEI
112	Tyr 191-200	GSEIWRDIDF
113	Tyr 192-200	SEIWRDIDF
114	Тут 193-201	EIWRDIDFA

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SEQ ID NO	IDENTITY	SEQUENCE
115	Tyr 407-416	LQEVYPEANA
116	Tyr 409-418	EVYPEANAPI
117	Tyr 410-418	VYPEANAPI
118	Tyr 411-418	YPEANAPI
119	Tyr 411-420	YPEANAPIGH
120	Tyr 416-425	APIGHNRESY
121	Tyr 417-425	PIGHNRESY
122	Tyr 417-426	PIGHNRESYM
123	Tyr 416-425	APIGHNRESY
124	Tyr 417-425	PIGHNRESY
125	Tyr 423-430	ESYMVPFI
126	Tyr 423-432	ESYMVPFIPL
127	Tyr 424-432	SYMVPFIPL
128	Tyr 424-433	SYMVPFIPLY
129	Tyr 425-433	YMVPFIPLY
130	Tyr 426-434	MVPFIPLYR
131	Tyr 426-435	MVPFIPLYRN
132	Тут 427-434	VPFPLYR
133	Tyr 430-437	IPLYRNGD
134	Tyr 430-439	IPLYRNGDFF
135	Tyr 431-439	PLYRNGDFF
136	Tyr 431-440	PLYRNGDFFI
137	Tyr 434-443	RNGDFFISSK
138	Tyr 435-443	NGDFFISSK
139	Тут 463-471	YIKSYLEQA
140	Tyr 466-474	SYLEQASRI
141	Tyr 469-478	EQASRIWSWL
142	Tyr 470-478	QASRIWSWL
143	Tyr 471-478	ASRIWSWL
144	Tyr 471-479	ASRIWSWLL
145	Tyr 473-481	RIWSWLLGA
146	CEA 92-100	GPAYSGREI
147	CEA 92-101	GPAYSGREII
148	CEA 93-100	PAYSGREI
149	CEA 93-101	PAYSGREII
150	CEA 93-102	PAYSGREIIY
151	CEA 94-102	AYSGREIIY
152	CEA 97-105	GREIIYPNA
153	CEA 98-107	REIIYPNASL
154	CEA 99-107	EIIYPNASL
155	CEA 99-108	EIIYPNASLL
156	CEA 100-107	IIYPNASL
157	CEA 100-108	IIYPNASLL
158	CEA 100-109	IIYPNASLLI
. 159	CEA 102-109	YPNASLLI
160	CEA 107-116	LLIQNIIQND
161	CEA 132-141	EEATGQFRVY
162	CEA 133-141	EATGQFRVY
163	CEA 141-149	YPELPKPSI

SEQ ID NO	IDENTITY	SEQUENCE.
164	CEA 142-149	PELPKPSI
165	CEA 225-233	RSDSVILNV
166	CEA 225-234	RSDSVILNVL
167	CEA 226-234	SDSVILNVL
168	CEA 226-235	SDSVILNVLY
169	CEA 227-235	DSVILNVLY
170	CEA 233-242	VLYGPDAPTI
171	CEA 234-242	LYGPDAPTI
172	CEA 235-242	YGPDAPTI
173	CEA 236-245	GPDAPTISPL
174	CEA 237-245	PDAPTISPL
175	CEA 238-245	DAPTISPL
176	CEA 239-247	APTISPLNT
177	CEA 240-249	PTISPLNTSY
178	CEA 241-249	TISPLNTSY
179	CEA 240-249	PTISPLNTSY
180	CEA 241-249	TISPLNTSY
181	CEA 246-255	NTSYRSGENL
182	CEA 247-255	TSYRSGENL
183	CEA 248-255	SYRSGENL
184	CEA 248-257	SYRSGENLNL
185	CEA 249-257	YRSGENLNL
186	CEA 251-259	SGENLNLSC
187	CEA 253-262	ENLNLSCHAA
188	CEA 254-262	NLNLSCHAA
189	CEA 260-269	HAASNPPAQY
190	CEA 261-269	AASNPPAQY
191	CEA 264-273	NPPAQYSWFV
192	CEA 265-273	PPAQYSWFV
193	CEA 266-273	PAQYSWFV
194	CEA 272-280	FVNGTFQQS
195	CEA 310-319	RTTVTTITVY
196	CEA 311-319	TTVTTITVY
197	CEA 319-327	YAEPPKPFI
198	CEA 319-328	YAEPPKPFIT
199	CEA 320-327	AEPPKPFI
200	CEA 321-328	EPPKPFIT
201	CEA 321-329	EPPKPFITS
202	CEA 322-329	PPKPFITS
203	CEA 382-391	SVTRNDVGPY
204	CEA 383-391	VTRNDVGPY
205	CEA 389-397	GPYECGIQN
206	CEA 391-399	YECGIQNEL
207	CEA 394-402	GIQNELSVD
208	CEA 403-411	HSDPVILNV
209	CEA 403-412	HSDPVILNVL
210	CEA 404-412	SDPVILNVL
211	CEA 404-413	SDPVILNVLY
212	CEA 405-412	DPVILNVL

SEQ ID NO	IDENTITY	SEQUENCE
213	CEA 405-413	DPVILNVLY
214	CEA 408-417	ILNVLYGPDD
215	CEA 411-420	VLYGPDDPTI
216	CEA 412-420	LYGPDDPTI
217	CEA 413-420	YGPDDPTI
218	CEA 417-425	DPTISPSYT
219	CEA 418-427	PTISPSYTYY
220	CEA 419-427	TISPSYTYY
221	CEA 418-427	PTISPSYTYY
222	CEA 419-427	TISPSYTYY
223	CEA 419-428	TISPSYTYYR
224	CEA 424-433	YTYYRPGVNL
225	CEA 425-433	TYYRPGVNL
226	CEA 426-433	YYRPGVNL
227	CEA 426-435	YYRPGVNLSL
228	CEA 427-435	YRPGVNLSL
229	CEA 428-435	RPGVNLSL
230	CEA 428-437	RPGVNLSLSC
231	CEA 430-438	GVNLSLSCH
232	CEA 431-440	VNLSLSCHAA
233	CEA 432-440	NLSLSCHAA
234	CEA 438-447	HAASNPPAQY
235	CEA 439-447	AASNPPAQY
236	CEA 442-451	NPPAQYSWLI
237	CEA 443-451	PPAQYSWLI
238	CEA 444-451	PAQYSWLI
239	CEA 449-458	WLIDGNIQQH
240	CEA 450-458	LIDGNIQQH
241	CEA 450-459	LIDGNIQQHT
242	CEA 581-590	RSDPVTLDVL
243	CEA 582-590	SDPVTLDVL
244	CEA 582-591	SDPVTLDVLY
245	CEA 583-590	DPVTLDVL
246	CEA 583-591	DPVTLDVLY
247	CEA 588-597	DVLYGPDTPI
248	CEA 589-597	VLYGPDTPI
249	CEA 596-605	PIISPPDSSY
250	CEA 597-605	IISPPDSSY
251	CEA 597-606	IISPPDSSYL
252	CEA 599-606	SPPDSSYL
253	CEA 600-608	PPDSSYLSG
254	CEA 600-609	PPDSSYLSGA
255	CEA 602-611	DSSYLSGANL
256	CEA 603-611	SSYLSGANL
257	CEA 604-613	SYLSGANLNL
258	CEA 605-613	YLSGANLNL
259	CEA 610-618	NLNLSCHSA
260	CEA 620-629	NPSPQYSWRI
261	CEA 622-629	SPQYSWRI

SEQ ID NO	IDENTITY	SEQUENCE
262	CEA 627-635	WRINGIPOO
263	CEA 628-636	RINGIPQQH
264	CEA 628-637	RINGIPQQHT
265	CEA 631-639	GIPQOHTOV
266	CEA 632-639	IPQQHTQV
267	CEA 644-653	KITPNNNGTY
268	CEA 645-653	ITPNNNGTY
269	CEA 647-656	PNNNGTYACF
270	CEA 648-656	NNNGTYACF
271	CEA 650-657	NGTYACFV
272	CEA 661-670	ATGRNNSIVK
273	CEA 662-670	TGRNNSIVK
274	CEA 664-672	RNNSIVKSI
275	CEA 666-674	NSIVKSITV
276	GAGE-1 7-16	STYRPRPRRY
277	GAGE-1 8-16	TYRPRPRRY
278	GAGE-1 10-18	RPRPRRYVE
279	GAGE-1 16-23	YVEPPEMI
280	GAGE-1 22-31	MIGPMRPEOF
281	GAGE-1 23-31	IGPMRPEOF
282	GAGE-1 24-31	GPMRPEQF
283	GAGE-1 105-114	KTPEEEMRSH
284	GAGE-1 106-115	TPEEEMRSHY
285	GAGE-1 107-115	PEEEMRSHY
286	GAGE-1 110-119	EMRSHYVAQT
287	GAGE-1 113-121	SHYVAQTGI
288	GAGE-1 115-124	YVAQTGILWL
289	GAGE-1 116-124	VAQTGILWL
290	GAGE-1 116-125	VAQTGILWLL
291	GAGE-1 117-125	AQTGILWLL
292	GAGE-1 118-126	QTGILWLLM
293	GAGE-1 118-127	QTGILWLLMN
294	GAGE-1 120-129	GILWLLMNNC
295	GAGE-1 121-129	ILWLLMNNC
296	GAGE-1 124-131	LLMNNCFL
297	GAGE-1 123-131	WLLMNNCFL
298	GAGE-1 122-130	LWLLMNNCF
299	GAGE-1 121-130	ILWLLMNNCF
300	GAGE-1 121-129	ILWLLMNNC
301	GAGE-1 120-129	GILWLLMNNC
302	GAGE-1 118-127	QTGILWLLMN
303	GAGE-1 118-126	QTGILWLLM
304	GAGE-1 117-125	AQTGILWLL
305	GAGE-1 116-125	VAQTGILWLL
306	GAGE-1 116-124	VAQTGILWL
307	GAGE-1 115-124	YVAQTGILWL
308	GAGE-1 113-121	SHYVAQTGI
309	MAGE-1 62-70	SAFPTTINF
310	MAGE-1 61-70	ASAFPTTINF

SEQ ID NO IDENTITY SEQUENCE 311 MAGE-1 60-68 GASAFPTTI 312 MAGE-1 57-66 SPQGASAFPT 313 MAGE-1 144-151 FGKASESL	
312 MAGE-1 57-66 SPQGASAFPT	
314 MAGE-1 143-151 IFGKASESL	
315 MAGE-1 142-151 EIFGKASESL	
316 MAGE-1 142-149 EIFGKASE	
317 MAGE-1 133-140 IKNYKHCF	
318 MAGE-1 132-140 VIKNYKHCF	
319 MAGE-1 131-140 SVIKNYKHCF	
320 MAGE-1 132-139 VIKNYKHC	
321 MAGE-1 131-139 SVIKNYKHC	
322 MAGE-1 128-136 MLESVIKNY	
323 MAGE-1 127-136 EMLESVIKNY	
324 MAGE-1 126-134 AEMLESVIK	
325 MAGE-2 274-283 GPRALIETSY	
326 MAGE-2 275-283 PRALIETSY	
327 MAGE-2 276-284 RALIETSYV	
328 MAGE-2 277-286 ALIETSYVKV	
329 MAGE-2 278-286 LIETSYVKV	
330 MAGE-2 278-287 LIETSYVKVL	
331 MAGE-2 279-287 IETSYVKVL	
332 MAGE-2 280-289 ETSYVKVLHH	
333 MAGE-2 282-291 SYVKVLHHTL	
334 MAGE-2 283-291 YVKVLHHTL	
335 MAGE-2 285-293 KVLHHTLKI	
336 MAGE-2 303-311 PLHERALRE	-
337 MAGE-2 302-309 PPLHERAL	
338 MAGE-2 301-309 YPPLHERAL	
339 MAGE-2 300-309 SYPPLHERAL	
340 MAGE-2 299-307 ISYPPLHER	
341 MAGE-2 298-307 HISYPPLHER	
342 MAGE-2 292-299 KIGGEPHI	
343 MAGE-2 291-299 LKIGGEPHI	
344 MAGE-2 290-299 TLKIGGEPHI	
345 MAGE-3 303-311 PLHEWVLRE	
346 MAGE-3 302-309 PPLHEWVL	
347 MAGE-3 301-309 YPPLHEWVL	
348 MAGE-3 301-308 YPPLHEWV	
349 MAGE-3 300-308 SYPPLHEWV	
350 MAGE-3 299-308 ISYPPLHEWV	
351 MAGE-3 298-307 HISYPPLHEW	
352 MAGE-3 293-301 ISGGPHISY	
353 MAGE-3 292-301 KISGGPHISY	
354 Melan-A 45-54 CWYCRRRNGY	
355 Melan-A 46-54 WYCRRRNGY	
356 Melan-A 47-55 YCRRRNGYR	
357 Melan-A 49-57 RRRNGYRAL	
358 Melan-A 51-60 RNGYRALMDK	
359 Melan-A 52-60 NGYRALMDK	

SEQ ID NO	IDENTITY	SEQUENCE
360	Melan-A 55-63	RALMDKSLH
361	Melan-A 56-63	ALMDKSLH
362	Melan-A 55-64	RALMDKSLHV
363	Melan-A 56-64	ALMDKSLHV
364	PRAME 275-284	YISPEKEEQY
365	PRAME 276-284	ISPEKEEQY
366	PRAME 277-285	SPEKEEQYI
367	PRAME 278-285	PEKEEQYI
368	PRAME 279-288	EKEEQYIAQF
369	PRAME 280-288	KEEQYIAQF
370	PRAME 283-292	QYIAQFTSQF
371	PRAME 284-292	YIAQFTSQF
372	PRAME 284-293	YIAQFTSQFL
373	PRAME 285-293	IAQFTSQFL
374	PRAME 286-295	AQFTSQFLSL
375	PRAME 287-295	QFTSQFLSL
376	PRAME 290-298	SQFLSLQCL
377	PRAME 439-448	VLYPVPLESY
378	PRAME 440-448	LYPVPLESY
379	PRAME 446-455	ESYEDIHGTL
380	PRAME 448-457	YEDIHGTLHL
381	PRAME 449-457	EDIHGTLHL
382	PRAME 451-460	IHGTLHLERL
383	PRAME 454-463	TLHLERLAYL
384	PRAME 455-463	LHLERLAYL
385	PRAME 456-463	HLERLAYL
386	PRAME 456-465	HLERLAYLHA.
387	PRAME 458-467	ERLAYLHARL
388	PRAME 459-467	RLAYLHARL
389	PRAME 459-468	RLAYLHARLR
390	PRAME 460-467	LAYLHARL
391 392	PRAME 460-468	LAYLHARLR
	PRAME 461-470	AYLHARLREL
393 394	PRAME 462-470	YLHARLREL
395	PRAME 462-471	YLHARLRELL
396	PRAME 463-471	LHARLRELL
397	PRAME 464-471	HARLRELL
398	PRAME 464-472 PRAME 469-478	HARLRELLC
399	PRAME 470-478	ELLCELGRPS
400	PSA 144-153	LLCELGRPS
401	PSA 145-153	QEPALGTTCY PPALGTTCY
402	PSA 162-171	EPALGTTCY DEEE! TDVVI
403	PSA 163-171	PEEFLTPKKL EEFLTPKKL
404	PSA 165-173	FLTPKKLQC
405	PSA 165-174	FLTPKKLQCV
406	PSA 166-174	LTPKKLQCV
407	PSA 167-174	TPKKLQCV
408	PSA 167-175	TPKKLQCVD
100	[15/X 10/-1/3	Ιτινντής Δη

CTO TO NO	I was the same of	
SEQ ID NO	IDENTITY	SEQUENCE
409	PSA 170-179	KLQCVDLHVI
410	PSA 171-179	LQCVDLHVI
411	PSCA 73-81	DSQDYYVGK
412	PSCA 74-82	SQDYYVGKK
413	PSCA 74-83	SQDYYVGKKN
414	PSCA 76-84	DYYVGKKNI
415	PSCA 77-84	YYVGKKNI
416	PSCA 78-86	YVGKKNITC
417	PSCA 78-87	YVGKKNITCC
418	PSMA 381-390	WVFGGIDPQS
419	PSMA 385-394	GIDPQSGAAV
420	PSMA 386-394	IDPQSGAAV
421	PSMA 387-394	DPQSGAAV
422	PSMA 387-395	DPQSGAAVV
423	PSMA 387-396	DPQSGAAVVH
424	PSMA 388-396	PQSGAAVVH
425	PSMA 389-398	QSGAAVVHEI
426	PSMA 390-398	SGAAVVHEI
427	PSMA 391-398	GAAVVHEI
428	PSMA 391-399	GAAVVHEIV
429	PSMA 392-399	AAVVHEIV
430	PSMA 597-605	CRDYAVVLR
431	PSMA 598-607	RDYAVVLRKY
432	PSMA 599-607	DYAVVLRKY
433	PSMA 600-607	YAVVLRKY
434	PSMA 602-611	VVLRKYADKI
435	PSMA 603-611	VLRKYADKI
436	PSMA 603-612	VLRKYADKIY
437	PSMA 604-611	LRKYADKI
438	PSMA 604-612	LRKYADKIY
.439	PSMA 605-614	RKYADKIYSI
440	PSMA 606-614	KYADKIYSI
441	PSMA 607-614	YADKIYSI
442	PSMA 616-625	MKHPQEMKTY
443	PSMA 617-625	KHPQEMKTY
444	PSMA 618-627	HPQEMKTYSV
445	SCP-1 62-71	DSDPALQKV
446	SCP-1 63-71	DSDPALQKV
447	SCP-1 67-76	ALQKVNFLPV
448	SCP-1 70-78	KVNFLPVLE
449	SCP-1 71-80	VNFLPVLEQV
450	SCP-1 72-80	NFLPVLEQV
451	SCP-1 75-84	PVLEQVGNSD
452	SCP-1 76-84	VLEQVGNSD
453	SCP-1 202-210	YEREETRQV
454	SCP-1 202-211	YEREETRQVY
455	SCP-1 203-211	EREETRQVY
456	SCP-1 203-212	EREETRQVYM
457	SCP-1 204-212	REETRQVYM

SEQ ID N		SEQUENCE
458	SCP-1 211-220	YMDLNSNIEK
459	SCP-1 213-221	DLNSNIEKM
460	SCP-1 216-226	SNIEKMITAF
461	SCP-1 217-225	NIEKMITAF
462	SCP-1 218-225	IEKMITAF
463	SCP-1 397-406	RLENYEDQLI
464	SCP-1 398-406	LENYEDQLI
465	SCP-1 398-407	LENYEDQLII
466	SCP-1 399-407	ENYEDQLII
467	SCP-1 399-408	ENYEDQLIIL
468	SCP-1 400-408	NYEDQLIIL
469	SCP-1 400-409	NYEDQLIILT
470	SCP-1 401-409	YEDQLIILT
471	SCP-1 401-410	YEDQLIILTM
472	SCP-1 402-410	EDQLILTM
473	SCP-1 406-415	
474	SCP-1 407-415	IILTMELQKT
475	SCP-1 424-432	ILTMELQKT
476	SCP-1 424-433	KLTNNKEVE
477	SCP-1 425-433	KLTNNKEVEL
478	SCP-1 429-438	LTNNKEVEL
479	SCP-1 430-438	KEVELEELKK
480	SCP-1 430-439	EVELEELKK
481	SCP-1 431-439	EVELEELKKV
482	SCP-1 530-539	VELEELKKV
483	SCP-1 531-539	ETSDMTLELK
484	SCP-1 548-556	TSDMTLELK
485	SCP-1 553-562	NKKQEERML
486	SCP-1 554-562	ERMLTQIENL
487	SCP-1 555-562	RMLTQIENL
488	SCP-1 555-564	MLTQIENL
489	SCP-1 560-569	MLTQIENLQE
490	SCP-1 561-569	ENLQETETQL
491	SCP-1 561-570	NLQETETQL
492	SCP-1 567-576	NLQETETQLR
493	SCP-1 568-576	TQLRNELEYV
494	SCP-1 571-580	QLRNELEYV
495	SCP-1 572-580	NELEYVREEL
496	SCP-1 573-580	ELEYVREEL
497	SCP-1 574-583	LEYVREEL
498	SCP-1 575-583	EYVREELKQK
499	SCP-1 675-684	YVREELKQK
500		LLEEVEKAKV
501	SCP-1 676-684	LEEVEKAKV
502	SCP-1 676-685	LEEVEKAKVI
503	SCP-1 677-685	EEVEKAKVI
504	SCP-1 681-690	KAKVIADEAV
505	SCP-1 683-692	KVIADEAVKL
506	SCP-1 684-692	VIADEAVKL
200	SCP-1 685-692	IADEAVKL

SEQ ID NO	IDENTITY	SEQUENCE
507	SCP-1 694-702	KEIDKRCQH
508	SCP-1 694-703	KEIDKRCOHK
509	SCP-1 695-703	EIDKRCOHK
510	SCP-1 695-704	EIDKRCQHKI
511	SCP-1 696-704	IDKRCOHKI
512	SCP-1 697-704	DKRCOHKI
513	SCP-1 698-706	KRCQHKIAE
514	SCP-1 698-707	KRCOHKIAEM
515	SCP-1 699-707	RCOHKIAEM
516	SCP-1 701-710	QHKIAEMVAL
517	SCP-1 702-710	HKIAEMVAL
518	SCP-1 703-710	KIAEMVAL
519	SCP-1 737-746	QEQSSLRASL
520	SCP-1 738-746	EQSSLRASL
521 .	SCP-1 739-746	QSSLRASL
522	SCP-1 741-750	SLRASLEIEL
523	SCP-1 742-750	LRASLEIEL
524	SCP-1 743-750	RASLEIEL
525	SCP-1 744-753	ASLEIELSNL
526	SCP-1 745-753	SLEIELSNL
527	SCP-1 745-754	SLEIELSNLK
528	SCP-1 746-754	LEIELSNLK
529	SCP-1 747-755	EIELSNLKA
530	SCP-1 749-758	ELSNLKAELL
531	SCP-1 750-758	LSNLKAELL
532	SCP-1 751-760	SNLKAELLSV
533	SCP-1 752-760	NLKAELLSV
534	SCP-1 752-761	NLKAELLSVK
535	SCP-1 753-761	LKAELLSVK
536	SCP-1 753-762	LKAELLSVKK
537	SCP-1 754-762	KAELLSVKK
538	SCP-1 755-763	AELLSVKKQ
539	SCP-1 787-796	EKKDKKTQTF
540	SCP-1 788-796	KKDKKTOTF
541	SCP-1 789-796	KDKKTQTF
542	SCP-1 797-806	LLETPDIYWK
543	SCP-1 798-806	LETPDIYWK
544	SCP-1 798-807	LETPDIYWKL
545	SCP-1 799-807	ETPDIYWKL
546	SCP-1 800-807	TPDIYWKL
547	SCP-1 809-817	SKAVPSQTV
548	SCP-1 810-817	KAVPSQTV
549	SCP-1 812-821	VPSQTVSRNF
550	SCP-1 815-824	QTVSRNFTSV
551	SCP-1 816-824	TVSRNFTSV
552	SCP-1 816-825	TVSRNFTSVD
553	SCP-1 823-832	SVDHGISKDK
554	SCP-1 829-838	SKDKRDYLWT
555	SCP-1 832-840	KRDYLWTSA

SEQ ID NO	IDENTITY	SEQUENCE
556	SCP-1 832-841	KRDYLWTSAK
557	SCP-1 833-841	RDYLWTSAK
558	SCP-1 835-843	YLWTSAKNT
559	SCP-1 835-844	YLWTSAKNTL
560	SCP-1 837-844	WTSAKNTL
561	SCP-1 841-850	KNTLSTPLPK
562	SCP-1 842-850	NTLSTPLPK
563	SCP-1 832-840	KRDYLWTSA
564	SCP-1 832-841	KRDYLWTSAK
565	SCP-1 833-841	RDYLWTSAK
566	SCP-1 835-843	YLWTSAKNT
567	SCP-1 839-846	SAKNTLST
568	SCP-1 841-850	KNTLSTPLPK
569	SCP-1 842-850	NTLSTPLPK
570	SCP-1 843-852	TLSTPLPKAY
571	SCP-1 844-852	LSTPLPKAY
572	SSX-2 5-12	DAFARRPT
573	SSX-2 7-15	FARRPTVGA
574	SSX-2 8-17	ARRPTVGAOI
575	SSX-2 9-17	RRPTVGAQI
576	SSX-2 10-17	RPTVGAQI
577	SSX-2 13-21	VGAQIPEKI
578	SSX-2 14-21	GAQIPEKI
579	SSX-2 15-24	AQIPEKIQKA
580	SSX-2 16-24	QIPEKIQKA
581	SSX-2 16-25	QIPEKIQKAF
582	SSX-2 17-24	IPEKIQKA
583	SSX-2 17-25	IPEKIQKAF
584	SSX-2 18-25	PEKIQKAF
· 585	Survivin 116-124	ETNNKKKEF
586	Survivin 117-124	TNNKKKEF
587	Survivin 122-131	KEFEETAKKV
588	Survivin 123-131	EFEETAKKV
589	Survivin 127-134	TAKKVRRA
590	Survivin 126-134	ETAKKVRRA
591	Survivin 128-136	AKKVRRAIE
592	Survivin 129-138	KKVRRAIEQL
593	Survivin 130-138	KVRRAIEQL
594	Survivin 130-139	KVRRAIEQLA
595	Survivin 131-138	VRRAIEQL
596	BAGE 24-31	SPVVSWRL
597	BAGE 21-29	KEESPVVSW
598	BAGE 19-27	LMKEESPVV
599	BAGE 18-27	RLMKEESPVV
600	BAGE 18-26	RLMKEESPV
601	BAGE 14-22	LLQARLMKE
602	BAGE 13-22	QLLQARLMKE
603	Survivin 13-28	FLKDHRISTFKNWPFL
604	Survivin 79-111	KHSSGCAFLSVKKQFEELTLGEFLKLDRERAKN

SEQ ID NO	DENTITY	SEQUENCE
605	Survivin 130-141	KVRRAIEQLAAM
606	GAGE-1 116-133	VAQTGILWLLMNNCFLNL
607	BAGE 7-17	FLALSAQLLQA
608	BAGE 18-27	RLMKEESPVV
609	BAGE 2-27	AARAVFLALSAQLLQARLMKEESPVV
610	BAGE 30-39	RLEPEDGTAL

*Any of SEQ ID NOS. 108-602 can be useful as epitopes in any of the various embodiments of the invention. Any of SEQ ID NOS. 603-610 can be useful as sequences containing epitopes or epitope clusters, as described in various embodiments of the invention.

**All accession numbers used here and throughout can be accessed through the NCBI databases, for example, through the Entrez seek and retrieval system on the world wide web.

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Note that the following discussion sets forth the inventors' understanding of the operation of the invention. However, it is not intended that this discussion limit the patent to any particular theory of operation not set forth in the claims.

In pursuing the development of epitope vaccines others have generated lists of predicted epitopes based on MHC binding motifs. Such peptides can be immunogenic, but may not correspond to any naturally produced antigenic fragment. Therefore, whole antigen will not elicit a similar response or sensitize a target cell to cytolysis by CTL. Therefore such lists do not differentiate between those sequences that can be useful as vaccines and those that cannot. Efforts to determine which of these predicted epitopes are in fact naturally produced have often relied on screening their reactivity with tumor infiltrating lymphocytes (TIL). However, TIL are strongly biased to recognize immune epitopes whereas tumors (and chronically infected cells) will generally present housekeeping epitopes. Thus, unless the epitope is produced by both the housekeeping and immuno- proteasomes, the target cell will generally not be recognized by CTL induced with TILidentified epitopes. The epitopes of the present invention, in contrast, are generated by the action of a specified proteasome, indicating that they can be naturally produced, and enabling their appropriate use. The importance of the distinction between housekeeping and immune epitopes to vaccine design is more fully set forth in PCT publication WO 01/82963A2. The teachings and embodiments disclosed in said PCT publication are contemplated as supporting principals and embodiments related to and useful in connection with the present invention.

The epitopes of the invention include or encode polypeptide fragments of TAAs that are precursors or products of proteasomal cleavage by a housekeeping or immune proteasome, and that contain or consist of a sequence having a known or predicted affinity for at least one allele of MHC I. In some embodiments, the epitopes include or encode a polypeptide of about 6 to 25 amino acids in length, preferably about 7 to 20 amino acids in length, more preferably about 8 to 15 amino acids in length, and still more preferably 9 or 10 amino acids in length. However, it is understood that the polypeptides can be larger as long as N-terminal trimming can produce the MHC epitope or that

they do not contain sequences that cause the polypeptides to be directed away from the proteasome or to be destroyed by the proteasome. For immune epitopes, if the larger peptides do not contain such sequences, they can be processed in the pAPC by the immune proteasome. Housekeeping epitopes may also be embedded in longer sequences provided that the sequence is adapted to facilitate liberation of the epitope's C-terminus by action of the immunoproteasome. The foregoing discussion has assumed that processing of longer epitopes proceeds through action of the immunoproteasome of the pAPC. However, processing can also be accomplished through the contrivance of some other mechanism, such as providing an exogenous protease activity and a sequence adapted so that action of the protease liberates the MHC epitope. The sequences of these epitopes can be subjected to computer analysis in order to calculate physical, biochemical, immunologic, or molecular genetic properties such as mass, isoelectric point, predicted mobility in electrophoresis, predicted binding to other MHC molecules, melting temperature of nucleic acid probes, reverse translations, similarity or homology to other sequences, and the like.

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In constructing the polynucleotides encoding the polypeptide epitopes of the invention, the gene sequence of the associated TAA can be used, or the polynucleotide can be assembled from any of the corresponding codons. For a 10 amino acid epitope this can constitute on the order of 10^6 different sequences, depending on the particular amino acid composition. While large, this is a distinct and readily definable set representing a miniscule fraction of the >10¹⁸ possible polynucleotides of this length, and thus in some embodiments, equivalents of a particular sequence disclosed herein encompass such distinct and readily definable variations on the listed sequence. In choosing a particular one of these sequences to use in a vaccine, considerations such as codon usage, self-complementarity, restriction sites, chemical stability, etc. can be used as will be apparent to one skilled in the art.

The invention contemplates producing peptide epitopes. Specifically these epitopes are derived from the sequence of a TAA, and have known or predicted affinity for at least one allele of MHC I. Such epitopes are typically identical to those produced on target cells or pAPCs.

<u>Compositions Containing Active Epitopes</u>

Embodiments of the present invention provide polypeptide compositions, including vaccines, therapeutics, diagnostics, pharmacological and pharmaceutical compositions. The various compositions include newly identified epitopes of TAAs, as well as variants of these epitopes. Other embodiments of the invention provide polynucleotides encoding the polypeptide epitopes of the invention. The invention further provides vectors for expression of the polypeptide epitopes for purification. In addition, the invention provides vectors for the expression of the polypeptide epitopes in an APC for use as an anti-tumor vaccine. Any of the epitopes or antigens, or nucleic acids encoding the same, from Table 1 can be used. Other embodiments relate to methods of making and using the various compositions.

A general architecture for a class I MHC-binding epitope can be described, and has been reviewed more extensively in Madden, D.R. Annu. Rev. Immunol. 13:587-622, 1995. Much of the binding energy arises from main chain contacts between conserved residues in the MHC molecule and the N- and C-termini of the peptide. Additional main chain contacts are made but vary among MHC alleles. Sequence specificity is conferred by side chain contacts of so-called anchor residues with pockets that, again, vary among MHC alleles. Anchor residues can be divided into primary and secondary. Primary anchor positions exhibit strong preferences for relatively well-defined sets of amino acid residues. Secondary positions show weaker and/or less well-defined preferences that can often be better described in terms of less favored, rather than more favored, residues. Additionally, residues in some secondary anchor positions are not always positioned to contact the pocket on the MHC molecule at all. Thus, a subset of peptides exists that bind to a particular MHC molecule and have a side chain-pocket contact at the position in question and another subset exists that show binding to the same MHC molecule that does not depend on the conformation the peptide assumes in the peptide-binding groove of the MHC molecule. The C-terminal residue (P Ω ; omega) is preferably a primary anchor residue. For many of the better studied HLA molecules (e.g. A2, A68, B27, B7, B35, and B53) the second position (P2) is also an anchor residue. However, central anchor residues have also been observed including P3 and P5 in HLA-B8, as well as P5 and PΩ(omega)-3 in the murine MHC molecules H-2Db and H-2Kb, respectively. Since more stable binding will generally improve immunogenicity, anchor residues are preferably conserved or optimized in the design of variants, regardless of their position.

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Because the anchor residues are generally located near the ends of the epitope, the peptide can buckle upward out of the peptide-binding groove allowing some variation in length. Epitopes ranging from 8-11 amino acids have been found for HLA-A68, and up to 13 amino acids for HLA-A2. In addition to length variation between the anchor positions, single residue truncations and extensions have been reported and the N- and C-termini, respectively. Of the non-anchor residues, some point up out of the groove, making no contact with the MHC molecule but being available to contact the TCR, very often P1, P4, and P Ω (omega)-1 for HLA-A2. Others of the non-anchor residues can become interposed between the upper edges of the peptide-binding groove and the TCR, contacting both. The exact positioning of these side chain residues, and thus their effects on binding, MHC fine conformation, and ultimately immunogenicity, are highly sequence dependent. For an epitope to be highly immunogenic it must not only promote stable enough TCR binding for activation to occur, but the TCR must also have a high enough off-rate that multiple TCR molecules can interact sequentially with the same peptide-MHC complex (Kalergis, A.M. et al., Nature Immunol. 2:229-234, 2001). Thus, without further information about the ternary complex, both conservative and non-conservative substitutions at these positions merit consideration when designing variants.

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The polypeptide epitope variants can be made, for example, using any of the techniques and guidelines for conservative and non-conservative mutations. Variants can be derived from substitution, deletion or insertion of one or more amino acids as compared with the native sequence. Amino acid substitutions can be the result of replacing one amino acid with another amino acid having similar structural and/or chemical properties, such as the replacement of a threonine with a serine, for example. Such replacements are referred to as conservative amino acid replacements, and all appropriate conservative amino acid replacements are considered to be embodiments of one invention. Insertions or deletions can optionally be in the range of about 1 to 4, preferably 1 to 2, amino acids. It is generally preferable to maintain the "anchor positions" of the peptide which are responsible for binding to the MHC molecule in question. Indeed, immunogenicity of peptides can be improved in many cases by substituting more preferred residues at the anchor positions (Franco, et al., Nature Immunology, 1(2):145-150, 2000). Immunogenicity of a peptide can also often be improved by substituting bulkier amino acids for small amino acids found in non-anchor positions while maintaining sufficient cross-reactivity with the original epitope to constitute a useful vaccine. The variation allowed can be determined by routine insertions, deletions or substitutions of amino acids in the sequence and testing the resulting variants for activity exhibited by the polypeptide epitope. Because the polypeptide epitope is often 9 amino acids, the substitutions preferably are made to the shortest active epitope, for example, an epitope of 9 amino acids.

Variants can also be made by adding any sequence onto the N-terminus of the polypeptide epitope variant. Such N-terminal additions can be from 1 amino acid up to at least 25 amino acids. Because peptide epitopes are often trimmed by N-terminal exopeptidases active in the pAPC, it is understood that variations in the added sequence can have no effect on the activity of the epitope. In preferred embodiments, the amino acid residues between the last upstream proteasomal cleavage site and the N-terminus of the MHC epitope do not include a proline residue. Serwold, T. at al., Nature Immunol. 2:644-651, 2001. Accordingly, effective epitopes can be generated from precursors larger than the preferred 9-mer class I motif.

Generally, peptides are useful to the extent that they correspond to epitopes actually displayed by MHC I on the surface of a target cell or a pACP. A single peptide can have varying affinities for different MHC molecules, binding some well, others adequately, and still others not appreciably (Table 2). MHC alleles have traditionally been grouped according to serologic reactivity which does not reflect the structure of the peptide-binding groove, which can differ among different alleles of the same type. Similarly, binding properties can be shared across types; groups based on shared binding properties have been termed supertypes. There are numerous alleles of MHC I in the human population; epitopes specific to certain alleles can be selected based on the genotype of the patient.

<u>Table 2.</u>

<u>Predicted Binding of Tyrosinase₂₀₇₋₂₁₆ (SEQ ID NO. 1) to Various MHC types</u>

MHC I type	*Half time of dissociation (min)
A1	0.05
A*0201	1311.
A*0205	50.4
A3	2.7
A*1101 (part of the A3 supertype)	0.012
A24	6.0
B7	4.0
B8	8.0
B14 (part of the B27 supertype)	60.0
B*2702	0.9
B*2705	30.0
B*3501 (part of the B7 supertype)	2.0
B*4403	0.1
B*5101 (part of the B7 supertype)	26.0
B*5102	55.0
B*5801	0.20
B60	0.40
B62	2.0

*HLA Peptide Binding Predictions (world wide web hypertext transfer protocol "access at bimas.dcrt.nih.gov/molbio/hla_bin").

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In further embodiments of the invention, the epitope, as peptide or encoding polynucleotide, can be administered as a pharmaceutical composition, such as, for example, a vaccine or an immunogenic composition, alone or in combination with various adjuvants, carriers, or excipients. It should be noted that although the term vaccine may be used throughout the discussion herein, the concepts can be applied and used with any other pharmaceutical composition, including those mentioned herein. Particularly advantageous adjuvants include various cytokines and oligonucleotides containing immunostimulatory sequences (as set forth in greater detail in the co-pending applications referenced herein). Additionally the polynucleotide encoded epitope can be contained in a virus (e.g. vaccinia or adenovirus) or in a microbial host cell (e.g. Salmonella or Listeria monocytogenes) which is then used as a vector for the polynucleotide (Dietrich, G. et al. Nat. Biotech. 16:181-185, 1998). Alternatively a pAPC can be transformed, ex vivo, to express the epitope, or pulsed with peptide epitope, to be itself administered as a vaccine. To increase efficiency of these processes, the encoded epitope can be carried by a viral or bacterial vector, or complexed with a ligand of a receptor found on pAPC. Similarly the peptide epitope can be complexed with or conjugated to a pAPC ligand. A vaccine can be composed of more than a single epitope.

Particularly advantageous strategies for incorporating epitopes and/or epitope clusters, into a vaccine or pharmaceutical composition are disclosed in PCT Publication WO 01/82963 and U.S.

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Patent Application No. 09/560,465 entitled "EPITOPE SYNCHRONIZATION IN ANTIGEN PRESENTING CELLS," filed on April 28, 2000. The teaching and embodiments disclosed in said PCT publication are contemplated as supporting principals and embodiments related to and useful in connection with the present invention. Epitope clusters for use in connection with this invention are disclosed in PCT Publication WO 01/82963 and U.S. Patent Application No. 09/561,571 entitled "EPITOPE CLUSTERS," filed on April 28, 2000. The teaching and embodiments disclosed in said PCT publication are contemplated as supporting principals and embodiments related to and useful in connection with the present invention.

Preferred embodiments of the present invention are directed to vaccines and methods for causing a pAPC or population of pAPCs to present housekeeping epitopes that correspond to the epitopes displayed on a particular target cell. Any of the epitopes or antigens in Table 1, can be used for example. In one embodiment, the housekeeping epitope is a TuAA epitope processed by the housekeeping proteasome of a particular tumor type. In another embodiment, the housekeeping epitope is a virus-associated epitope processed by the housekeeping proteasome of a cell infected with a virus. This facilitates a specific T cell response to the target cells. Concurrent expression by the pAPCs of multiple epitopes, corresponding to different induction states (pre- and post-attack), can drive a CTL response effective against target cells as they display either housekeeping epitopes or immune epitopes.

By having both housekeeping and immune epitopes present on the pAPC, this embodiment can optimize the cytotoxic T cell response to a target cell. With dual epitope expression, the pAPCs can continue to sustain a CTL response to the immune-type epitope when the tumor cell switches from the housekeeping proteasome to the immune proteasome with induction by IFN, which, for example, may be produced by tumor-infiltrating CTLs.

In a preferred embodiment, immunization of a patient is with a vaccine that includes a housekeeping epitope. Many preferred TAAs are associated exclusively with a target cell, particularly in the case of infected cells. In another embodiment, many preferred TAAs are the result of deregulated gene expression in transformed cells, but are found also in tissues of the testis, ovaries and fetus. In another embodiment, useful TAAs are expressed at higher levels in the target cell than in other cells. In still other embodiments, TAAs are not differentially expressed in the target cell compare to other cells, but are still useful since they are involved in a particular function of the cell and differentiate the target cell from most other peripheral cells; in such embodiments, healthy cells also displaying the TAA may be collaterally attacked by the induced T cell response, but such collateral damage is considered to be far preferable to the condition caused by the target cell.

The vaccine contains a housekeeping epitope in a concentration effective to cause a pAPC or populations of pAPCs to display housekeeping epitopes. Advantageously, the vaccine can

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include a plurality of housekeeping epitopes or one or more housekeeping epitopes optionally in combination with one or more immune epitopes. Formulations of the vaccine contain peptides and/or nucleic acids in a concentration sufficient to cause pAPCs to present the epitopes. The formulations preferably contain epitopes in a total concentration of about 1µg-1mg/100µl of vaccine preparation. Conventional dosages and dosing for peptide vaccines and/or nucleic acid vaccines can be used with the present invention, and such dosing regimens are well understood in the art. In one embodiment, a single dosage for an adult human may advantageously be from about 1 to about 5000 µl of such a composition, administered one time or multiple times, e.g., in 2, 3, 4 or more dosages separated by 1 week, 2 weeks, 1 month, or more. insulin pump delivers 1 ul per hour (lowest frequency) ref intranodal method patent.

The compositions and methods of the invention disclosed herein further contemplate incorporating adjuvants into the formulations in order to enhance the performance of the vaccines. Specifically, the addition of adjuvants to the formulations is designed to enhance the delivery or uptake of the epitopes by the pAPCs. The adjuvants contemplated by the present invention are known by those of skill in the art and include, for example, GMCSF, GCSF, IL-2, IL-12, BCG, tetanus toxoid, osteopontin, and ETA-1.

In some embodiments of the invention, the vaccines can include a recombinant organism, such as a virus, bacierium or parasite, genetically engineered to express an epitope in a host. For example, *Listeria monocytogenes*, a gram-positive, facultative intracellular bacterium, is a potent vector for targeting TuAAs to the immune system. In a preferred embodiment, this vector can be engineered to express a housekeeping epitope to induce therapeutic responses. The normal route of infection of this organism is through the gut and can be delivered orally. In another embodiment, an adenovirus (Ad) vector encoding a housekeeping epitope for a TuAA can be used to induce anti-virus or anti-tumor responses. Bone marrow-derived dendritic cells can be transduced with the virus construct and then injected, or the virus can be delivered directly via subcutaneous injection into an animal to induce potent T-cell responses. Another embodiment employs a recombinant vaccinia virus engineered to encode amino acid sequences corresponding to a housekeeping epitope for a TAA. Vaccinia viruses carrying constructs with the appropriate nucleotide substitutions in the form of a minigene construct can direct the expression of a housekeeping epitope, leading to a therapeutic T cell response against the epitope.

The immunization with DNA requires that APCs take up the DNA and express the encoded proteins or peptides. It is possible to encode a discrete class I peptide on the DNA. By immunizing with this construct, APCs can be caused to express a housekeeping epitope, which is then displayed on class I MHC on the surface of the cell for stimulating an appropriate CTL response. Constructs generally relying on termination of translation or non-proteasomal proteases for generation of proper termini of housekeeping epitopes have been described in PCT Publication

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WO 01/82963 and U.S. Patent application No. 09/561,572 entitled EXPRESSION VECTORS ENCODING EPITOPES OF TARGET-ASSOCIATED ANTIGENS, filed on April 28, 2000. The teaching and embodiments disclosed in said PCT publication are contemplated as supporting principals and embodiments related to and useful in connection with the present invention.

As mentioned, it can be desirable to express housekeeping peptides in the context of a larger protein. Processing can be detected even when a small number of amino acids are present beyond the terminus of an epitope. Small peptide hormones are usually proteolytically processed from longer translation products, often in the size range of approximately 60-120 amino acids. This fact has led some to assume that this is the minimum size that can be efficiently translated. In some embodiments, the housekeeping peptide can be embedded in a translation product of at least about 60 amino acids. In other embodiments the housekeeping peptide can be embedded in a translation product of at least about 50, 30, or 15 amino acids.

Due to differential proteasomal processing, the immune proteasome of the pAPC produces peptides that are different from those produced by the housekeeping proteasome in peripheral body cells. Thus, in expressing a housekeeping peptide in the context of a larger protein, it is preferably expressed in the APC in a context other than its full length native sequence, because, as a housekeeping epitope, it is generally only efficiently processed from the native protein by the housekeeping proteasome, which is not active in the APC. In order to encode the housekeeping epitope in a DNA sequence encoding a larger protein, it is useful to find flanking areas on either side of the sequence encoding the epitope that permit appropriate cleavage by the immune proteasome in order to liberate that housekeeping epitope. Altering flanking amino acid residues at the N-terminus and C-terminus of the desired housekeeping epitope can facilitate appropriate cleavage and generation of the housekeeping epitope in the APC. Sequences embedding housekeeping epitopes can be designed *de novo* and screened to determine which can be successfully processed by immune proteasomes to liberate housekeeping epitopes.

Alternatively, another strategy is very effective for identifying sequences allowing production of housekeeping epitopes in APC. A contiguous sequence of amino acids can be generated from head to tail arrangement of one or more housekeeping epitopes. A construct expressing this sequence is used to immunize an animal, and the resulting T cell response is evaluated to determine its specificity to one or more of the epitopes in the array. By definition, these immune responses indicate housekeeping epitopes that are processed in the pAPC effectively. The necessary flanking areas around this epitope are thereby defined. The use of flanking regions of about 4-6 amino acids on either side of the desired peptide can provide the necessary information to facilitate proteasome processing of the housekeeping epitope by the immune proteasome. Therefore, a sequence ensuring epitope synchronization of approximately 16-22 amino acids can be inserted into, or fused to, any protein sequence effectively to result in that

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housekeeping epitope being produced in an APC. In alternate embodiments the whole head-to-tail array of epitopes, or just the epitopes immediately adjacent to the correctly processed housekeeping epitope can be similarly transferred from a test construct to a vaccine vector.

In a preferred embodiment, the housekeeping epitopes can be embedded between known immune epitopes, or segments of such, thereby providing an appropriate context for processing. The abutment of housekeeping and immune epitopes can generate the necessary context to enable the immune proteasome to liberate the housekeeping epitope, or a larger fragment, preferably including a correct C-terminus. It can be useful to screen constructs to verify that the desired epitope is produced. The abutment of housekeeping epitopes can generate a site cleavable by the immune proteasome. Some embodiments of the invention employ known epitopes to flank housekeeping epitopes in test substrates; in others, screening as described below are used whether the flanking regions are arbitrary sequences or mutants of the natural flanking sequence, and whether or not knowledge of proteasomal cleavage preferences are used in designing the substrates.

Cleavage at the mature N-terminus of the epitope, while advantageous, is not required, since a variety of N-terminal trimming activities exist in the cell that can generate the mature N-terminus of the epitope subsequent to proteasomal processing. It is preferred that such N-terminal extension be less than about 25 amino acids in length and it is further preferred that the extension have few or no proline residues. Preferably, in screening, consideration is given not only to cleavage at the ends of the epitope (or at least at its C-terminus), but consideration also can be given to ensure limited cleavage within the epitope.

Shotgun approaches can be used in designing test substrates and can increase the efficiency of screening. In one embodiment multiple epitopes can be assembled one after the other, with individual epitopes possibly appearing more than once. The substrate can be screened to determine which epitopes can be produced. In the case where a particular epitope is of concern a substrate can be designed in which it appears in multiple different contexts. When a single epitope appearing in more than one context is liberated from the substrate additional secondary test substrates, in which individual instances of the epitope are removed, disabled, or are unique, can be used to determine which are being liberated and truly constitute sequences ensuring epitope synchronization.

Several readily practicable screens exist. A preferred *in vitro* screen utilizes proteasomal digestion analysis, using purified immune proteasomes, to determine if the desired housekeeping epitope can be liberated from a synthetic peptide embodying the sequence in question. The position of the cleavages obtained can be determined by techniques such as mass spectrometry, HPLC, and N-terminal pool sequencing; as described in greater detail in U. S. Patent Applications entitled METHOD OF EPITOPE DISCOVERY, EPITOPE SYNCHRONIZATION IN ANTIGEN

PRESENTING CELLS, PCT Publication, U.S. applications and Provisional U.S. Patent Applications entitled EPITOPE SEQUENCES.

Alternatively, in vivo screens such as immunization or target sensitization can be employed. For immunization a nucleic acid construct capable of expressing the sequence in question is used. Harvested CTL can be tested for their ability to recognize target cells presenting the housekeeping epitope in question. Such targets cells are most readily obtained by pulsing cells expressing the appropriate MHC molecule with synthetic peptide embodying the mature housekeeping epitope. Alternatively, cells known to express housekeeping proteasome and the antigen from which the housekeeping epitope is derived, either endogenously or through genetic engineering, can be used. To use target sensitization as a screen, CTL, or preferably a CTL clone, that recognizes the housekeeping epitope can be used. In this case it is the target cell that expresses the embedded housekeeping epitope (instead of the pAPC during immunization) and it must express immune proteasome. Generally, the target cell can be transformed with an appropriate nucleic acid construct to confer expression of the embedded housekeeping epitope. Loading with a synthetic peptide embodying the embedded epitope using peptide loaded liposomes or a protein transfer reagent such as BIOPORTERTM (Gene Therapy Systems, San Diego, CA) represents an alternative.

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Additional guidance on nucleic acid constructs useful as vaccines in accordance with the present invention are disclosed in WO 01/82963 and U.S. Patent Application No. 09/561,572 entitled "EXPRESSION VECTORS ENCODING EPITOPES OF TARGET-ASSOCIATED ANTIGENS," filed on April 28, 2000. Further, expression vectors and methods for their design, which are useful in accordance with the present invention are disclosed in PCT Publication WO 03/063770; U.S. Patent Application No. 10/292,413, filed on November 7, 2002; and U.S. Provisional Application No. 60/336,968 (attorney docket number CTLIMM.022PR) entitled "EXPRESSION VECTORS ENCODING EPITOPES OF TARGET-ASSOCIATED ANTIGENS AND METHODS FOR THEIR DESIGN," filed on 11/7/2001. The teaching and embodiments disclosed in said PCT publications are contemplated as supporting principals and embodiments related to and useful in connection with the present invention.

A preferred embodiment of the present invention includes a method of administering a vaccine including an epitope (or epitopes) to induce a therapeutic immune response. The vaccine is administered to a patient in a manner consistent with the standard vaccine delivery protocols that are known in the art. Methods of administering epitopes of TAAs including, without limitation, transdermal, intranodal, perinodal, oral, intravenous, intradermal, intramuscular, intraperitoneal, and mucosal administration, including delivery by injection, instillation or inhalation. A particularly useful method of vaccine delivery to elicit a CTL response is disclosed in Australian Patent No. 739189 issued January 17, 2002; PCT Publication No. WO 099/02183; U.S. Patent

Application No. 09/380,534, filed on September 1, 1999; a Continuation-in-Part thereof U.S. Patent Application No. 09/776,232 both entitled "A METHOD OF INDUCING A CTL RESPONSE," filed on February 2, 2001, published as 20020007173; and PCT Publication No. WO 02/062368. The teachings and embodiments disclosed in said publications and applications are contemplated as supporting principals and embodiments related to and useful in connection with the present invention.

Reagents Recognizing Epitopes

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In another aspect of the invention, proteins with binding specificity for the epitope and/or the epitope-MHC molecule complex are contemplated, as well as the isolated cells by which they can be expressed. In one set of embodiments these reagents take the form of immunoglobulins: polyclonal sera or monoclonal antibodies (mAb), methods for the generation of which are well know in the art. Generation of mAb with specificity for peptide-MHC molecule complexes is known in the art. See, for example, Aharoni et al. *Nature* 351:147-150, 1991; Andersen et al. *Proc. Natl. Acad. Sci. USA* 93:1820-1824, 1996; Dadaglio et al. *Immunity* 6:727-738, 1997; Duc et al. *Int. Immunol.* 5:427-431,1993; Eastman et al. *Eur. J. Immunol.* 26:385-393, 1996; Engberg et al. *Immunotechnology* 4:273-278, 1999; Porgdor et al. *Immunol.* 6:715-726, 1997; Puri et al. *J. Immunol.* 158:2471-2476, 1997; and Polakova, K., et al. *J. Immunol.* 165 342-348, 2000.

In other embodiments the compositions can be used to induce and generate, in vivo and in vitro, T-cells specific for the any of the epitopes and/or epitope-MHC complexes. In preferred embodiments the epitope can be any one or more of those listed in TABLE 1, for example. Thus, embodiments also relate to and include isolated T cells, T cell clones, T cell hybridomas, or a protein containing the T cell receptor (TCR) binding domain derived from the cloned gene, as well as a recombinant cell expressing such a protein. Such TCR derived proteins can be simply the extra-cellular domains of the TCR, or a fusion with portions of another protein to confer a desired property or function. One example of such a fusion is the attachment of TCR binding domains to the constant regions of an antibody molecule so as to create a divalent molecule. The construction and activity of molecules following this general pattern have been reported, for example, Plaksin, D. et al. J. Immunol. 158:2218-2227, 1997 and Lebowitz, M.S. et al. Cell Immunol. 192:175-184, 1999. The more general construction and use of such molecules is also treated in U.S. patent 5,830,755 entitled T CELL RECEPTORS AND THEIR USE IN THERAPEUTIC AND DIAGNOSTIC METHODS.

The generation of such T cells can be readily accomplished by standard immunization of laboratory animals, and reactivity to human target cells can be obtained by immunizing with human target cells or by immunizing HLA-transgenic animals with the antigen/epitope. For some therapeutic approaches T cells derived from the same species are desirable. While such a cell can be created by cloning, for example, a murine TCR into a human T cell as contemplated above, in

vitro immunization of human cells offers a potentially faster option. Techniques for in vitro immunization, even using naive donors, are know in the field, for example, Stauss et al., *Proc. Natl. Acad. Sci. USA* 89:7871-7875, 1992; Salgaller et al. *Cancer Res.* 55:4972-4979, 1995; Tsai et al., *J. Immunol.* 158:1796-1802, 1997; and Chung et al., *J. Immunother.* 22:279-287, 1999.

Any of these molecules can be conjugated to enzymes, radiochemicals, fluorescent tags, and toxins, so as to be used in the diagnosis (imaging or other detection), monitoring, and treatment of the pathogenic condition associated with the epitope. Thus a toxin conjugate can be administered to kill tumor cells, radiolabeling can facilitate imaging of epitope positive tumor, an enzyme conjugate can be used in an ELISA-like assay to diagnose cancer and confirm epitope expression in biopsied tissue. In a further embodiment, such T cells as set forth above, following expansion accomplished through stimulation with the epitope and/or cytokines, can be administered to a patient as an adoptive immunotherapy.

Reagents Comprising Epitopes

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A further aspect of the invention provides isolated epitope-MHC complexes. In a particularly advantageous embodiment of this aspect of the invention, the complexes can be soluble, multimeric proteins such as those described in U. S. Patent No. 5,635,363 (tetramers) or U. S. Patent No. 6,015,884 (Ig-dimers). Such reagents are useful in detecting and monitoring specific T cell responses, and in purifying such T cells.

Isolated MHC molecules complexed with epitopic peptides can also be incorporated into planar lipid bilayers or liposomes. Such compositions can be used to stimulate T cells *in vitro* or, in the case of liposomes, *in vivo*. Co-stimulatory molecules (e.g. B7, CD40, LFA-3) can be incorporated into the same compositions or, especially for *in vitro* work, co-stimulation can be provided by anti-co-receptor antibodies (e.g. anti-CD28, anti-CD154, anti-CD2) or cytokines (e.g. IL-2, IL-12). Such stimulation of T cells can constitute vaccination, drive expansion of T cells *in vitro* for subsequent infusion in an immuotherapy, or constitute a step in an assay of T cell function.

The epitope, or more directly its complex with an MHC molecule, can be an important constituent of functional assays of antigen-specific T cells at either an activation or readout step or both. Of the many assays of T cell function current in the art (detailed procedures can be found in standard immunological references such as Current Protocols in Immunology 1999 John Wiley & Sons Inc., N.Y.) two broad classes can be defined, those that measure the response of a pool of cells and those that measure the response of individual cells. Whereas the former conveys a global measure of the strength of a response, the latter allows determination of the relative frequency of responding cells. Examples of assays measuring global response are cytotoxicity assays, ELISA, and proliferation assays detecting cytokine secretion. Assays measuring the responses of individual cells (or small clones derived from them) include limiting dilution analysis (LDA),

ELISPOT, flow cytometric detection of unsecreted cytokine (described in U.S. Patent No. 5,445,939, entitled "METHOD FOR ASSESSMENT OF THE MONONUCLEAR LEUKOCYTE IMMUNE SYSTEM" and U.S. Patent Nos 5,656,446; and 5,843,689, both entitled "METHOD FOR THE ASSESSMENT OF THE MONONUCLEAR LEUKOCYTE IMMUNE SYSTEM," reagents for which are sold by Becton, Dickinson & Company under the tradename 'FASTIMMUNE') and detection of specific TCR with tetramers or Ig-dimers as stated and referenced above. The comparative virtues of these techniques have been reviewed in Yee, C. et al. Current Opinion in Immunology, 13:141–146, 2001. Additionally detection of a specific TCR rearrangement or expression can be accomplished through a variety of established nucleic acid based techniques, particularly in situ and single-cell PCR techniques, as will be apparent to one of skill in the art.

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These functional assays are used to assess endogenous levels of immunity, response to an immunologic stimulus (e.g. a vaccine), and to monitor immune status through the course of a disease and treatment. Except when measuring endogenous levels of immunity, any of these assays presume a preliminary step of immunization, whether *in vivo* or *in vitro* depending on the nature of the issue being addressed. Such immunization can be carried out with the various embodiments of the invention described above or with other forms of immunogen (e.g., pAPC-tumor cell fusions) that can provoke similar immunity. With the exception of PCR and tetramer/Ig-dimer type analyses which can detect expression of the cognate TCR, these assays generally benefit from a step of *in vitro* antigenic stimulation which can advantageously use various embodiments of the invention as described above in order to detect the particular functional activity (highly cytolytic responses can sometimes be detected directly). Finally, detection of cytolytic activity requires epitope-displaying target cells, which can be generated using various embodiments of the invention. The particular embodiment chosen for any particular step depends on the question to be addressed, ease of use, cost, and the like, but the advantages of one embodiment over another for any particular set of circumstances will be apparent to one of skill in the art.

The peptide MHC complexes described in this section have traditionally been understood to be non-covalent associations. However it is possible, and can be advantageous, to create a covalent linkages, for example by encoding the epitope and MHC heavy chain or the epitope, ß2-microglobulin, and MHC heavy chain as a single protein (Yu, Y.L.Y., et al., J. Immunol. 168:3145-3149, 2002; Mottez, E., et at., J. Exp. Med. 181:493,1995; Dela Cruz, C. S., et al., Int. Immunol. 12:1293, 2000; Mage, M. G., et al., Proc. Natl. Acad. Sci. USA 89:10658,1992; Toshitani, K., et al., Proc. Natl. Acad. Sci. USA 93:236,1996; Lee, L., et al., Eur. J. Immunol. 24:2633,1994; Chung, D. H., et al., J. Immunol. 163:3699,1999; Uger, R. A. and B. H. Barber, J. Immunol. 160:1598, 1998; Uger, R. A., et al., J. Immunol. 162:2671, 1999). Such constructs can have superior stability and overcome roadblocks in the processing-

presentation pathway. They can be used in the already described vaccines, reagents, and assays in similar fashion.

Tumor Associated Antigens

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Epitopes of the present invention are derived from the TuAAs tyrosinase (SEQ ID NO. 2), SSX-2, (SEQ ID NO. 3), PSMA (prostate-specific membrane antigen) (SEQ ID NO. 4), MAGE-1 (SEQ ID NO. 71), MAGE-2 (SEQ ID NO. 72), MAGE-3 (SEQ ID NO. 73), PRAME, (SEQ ID NO. 77), PSA, (SEQ ID NO. 78), PSCA, (SEQ ID NO. 79), CEA (carcinoembryonic antigen), (SEQ ID NO. 88), SCP-1 (SEQ ID NO. 92), GAGE-1, (SEQ ID NO. 96), survivin, (SEQ ID NO. 98), Melan-A/MART-1 (SEQ ID NO. 100), and BAGE (SEQ ID NO. 102). The natural coding sequences for these fifteen proteins, or any segments within them, can be determined from their cDNA or complete coding (cds) sequences, SEQ ID NOS. 5-7, 81-83, 85-87, 89, 93, 97, 99, 101, and 103, respectively.

Tyrosinase is a melanin biosynthetic enzyme that is considered one of the most specific markers of melanocytic differentiation. Tyrosinase is expressed in few cell types, primarily in melanocytes, and high levels are often found in melanomas. The usefulness of tyrosinase as a TuAA is taught in U.S. Patent 5,747,271 entitled "METHOD FOR IDENTIFYING INDIVIDUALS SUFFERING FROM A CELLULAR ABNORMALITY SOME OF WHOSE ABNORMAL CELLS PRESENT COMPLEXES OF HLA-A2/TYROSINASE DERIVED PEPTIDES, AND METHODS FOR TREATING SAID INDIVIDUALS."

GP100, also known as PMel17, also is a melanin biosynthetic protein expressed at high-levels in melanomas. GP100 as a TuAA is disclosed in U.S. Patent 5,844,075 entitled "MELANOMA ANTIGENS AND THEIR USE IN DIAGNOSTIC AND THERAPEUTIC METHODS."

Melan-A, also called MART-1 (Melanoma Antigen Recognized by T cells), is another melanin biosynthetic protein expressed at high levels in melanomas. The usefulness of Melan-A/MART-1 as a TuAA is taught in U.S. Patent Nos. 5,874,560 and 5,994,523 both entitiled "MELANOMA ANTIGENS AND THEIR USE IN DIAGNOSTIC AND THERAPEUTIC METHODS," as well as U.S. Patent No. 5,620,886, entitled "ISOLATED NUCLEIC ACID SEQUENCE CODING FOR A TUMOR REJECTION ANTIGEN PRECURSOR PROCESSED TO AT LEAST ONE TUMOR REJECTION ANTIGEN PRESENTED BY HLA-A2."

SSX-2, also know as Hom-Mel-40, is a member of a family of highly conserved cancertestis antigens (Gure, A.O. et al. *Int. J. Cancer* 72:965-971, 1997). Its identification as a TuAA is taught in U.S. Patent 6,025,191 entitled "ISOLATED NUCLEIC ACID MOLECULES WHICH ENCODE A MELANOMA SPECIFIC ANTIGEN AND USES THEREOF." Cancer-testis antigens are found in a variety of tumors, but are generally absent from normal adult tissues except testis. Expression of different members of the SSX family have been found variously in tumor cell

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lines. Due to the high degree of sequence identity among SSX family members, similar epitopes from more than one member of the family will be generated and able to bind to an MHC molecule, so that some vaccines directed against one member of this family can cross-react and be effective against other members of this family (see example 3 below).

MAGE-1, MAGE-2, and MAGE-3 are members of another family of cancer-testis antigens originally discovered in melanoma (MAGE is a contraction of melanoma-associated antigen) but found in a variety of tumors. The identification of MAGE proteins as TuAAs is taught in U.S. Patent 5,342,774 entitled NUCLEOTIDE SEQUENCE ENCODING THE TUMOR REJECTION ANTIGEN PRECURSOR, MAGE-1, and in numerous subsequent patents. Currently there are 17 entries for (human) MAGE in the SWISS Protein database. There is extensive similarity among these proteins so in many cases, an epitope from one can induce a cross-reactive response to other members of the family. A few of these have not been observed in tumors, most notably MAGE-H1 and MAGE-D1, which are expressed in testes and brain, and bone marrow stromal cells, respectively. The possibility of cross-reactivity on normal tissue is ameliorated by the fact that they are among the least similar to the other MAGE proteins.

GAGE-1 is a member of the GAGE family of cancer testis antigens (Van den Eynde, B., et al., *J. Exp. Med.* 182: 689-698, 1995; U.S Patent Nos. 5,610,013; 5648226; 5,858,689; 6,013,481; and 6,069,001). The PubGene database currently lists 12 distinct accessible members, some of which are synonymously known as PAGE or XAGE. GAGE-1 through GAGE-8 have a very high-degree of sequence identity, so most epitopes can be shared among multiple members of the family.

BAGE is a cancer-testis antigen commonly expressed in melanoma, particularly metastatic melanoma, as well as in carcinomas of the lung, breast, bladder, and squamous cells of the head and neck. It's usefulness as a TuAA is taught in U.S. Patent Nos. 5,683,88 entiltled "TUMOR REJECTION ANTIGENS WHICH CORRESPOND TO AMINO ACID SEQUENCES IN TUMOR REJECTION ANTIGEN PRECURSOR BAGE, AND USES THEREOF" and 5,571,711 entitled "ISOLATED NUCLEIC ACID MOLECULES CODING FOR BAGE TUMOR REJECTION ANTIGEN PRECURSORS."

NY-ESO-1, is a cancer-testis antigen found in a wide variety of tumors, also known as CTAG-1 (Cancer-Testis Antigen-1) and CAG-3 (Cancer Antigen-3). NY-ESO-1 as a TuAA is disclosed in U.S. Patent 5,804,381 entitled ISOLATED NUCLEIC ACID MOLECULE ENCODING AN ESOPHAGEAL CANCER ASSOCIATED ANTIGEN, THE ANTIGEN ITSELF, AND USES THEREOF. A paralogous locus encoding antigens with extensive sequence identity, LAGE-1a/s (SEQ ID NO. 75) and LAGE-1b/L (SEQ ID NO. 76), have been disclosed in publicly available assemblies of the human genome, and have been concluded to arise through alternate splicing. Additionally, CT-2 (or CTAG-2, Cancer-Testis Antigen-2) appears to be either an allele, a mutant, or a sequencing discrepancy of LAGE-1b/L. Due to the extensive sequence identity,

many epitopes from NY-ESO-1 can also induce immunity to tumors expressing these other antigens. See figure 1. The proteins are virtually identical through amino acid 70. From 71-134 the longest run of identities between NY-ESO-1 and LAGE is 6 residues, but potentially cross-reactive sequences are present. And from 135-180 NY-ESO and LAGE-1a/s are identical except for a single residue, but LAGE-1b/L is unrelated due to the alternate splice. The CAMEL and LAGE-2 antigens appear to derive from the LAGE-1 mRNA, but from alternate reading frames, thus giving rise to unrelated protein sequences. More recently, GenBank Accession AF277315.5, Homo sapiens chromosome X clone RP5-865E18, RP5-1087L19, complete sequence, reports three independent loci in this region which are labeled as LAGE1 (corresponding to CTAG-2 in the genome assemblies), plus LAGE2-A and LAGE2-B (both corresponding to CTAG-1 in the genome assemblies).

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PSMA (prostate-specific membranes antigen), a TuAA described in U.S. Patent 5,538,866 entitled "PROSTATE-SPECIFIC MEMBRANES ANTIGEN", is expressed by normal prostate epithelium and, at a higher level, in prostatic cancer. It has also been found in the neovasculature of non-prostatic tumors. PSMA can thus form the basis for vaccines directed to both prostate cancer and to the neovasculature of other tumors. This later concept is more fully described in U.S. Patent Publication No. 20030046714; PCT Publication No. WO 02/069907; and a provisional U.S. Patent application No. 60/274,063 entitled ANTI-NEOVASCULAR VACCINES FOR CANCER, filed March 7, 2001, and U.S. Application No. 10/094,699, attorney docket number CTLIMM.015A, filed on March 7, 2002, entitled "ANTI-NEOVASCULAR PREPARATIONS FOR CANCER." The teachings and embodiments disclosed in said publications and applications are contemplated as supporting principals and embodiments related to and useful in connection with the present invention. Briefly, as tumors grow they recruit ingrowth of new blood vessels. This is understood to be necessary to sustain growth as the centers of unvascularized tumors are generally necrotic and angiogenesis inhibitors have been reported to cause tumor regression. Such new blood vessels, or neovasculature, express antigens not found in established vessels, and thus can be specifically targeted. By inducing CTL against neovascular antigens the vessels can be disrupted, interrupting the flow of nutrients to (and removal of wastes from) tumors, leading to regression.

Alternate splicing of the PSMA mRNA also leads to a protein with an apparent start at Met₅₈, thereby deleting the putative membrane anchor region of PSMA as described in U.S. Patent 5,935,818 entitled "ISOLATED NUCLEIC ACID MOLECULE ENCODING ALTERNATIVELY SPLICED PROSTATE-SPECIFIC MEMBRANES ANTIGEN AND USES THEREOF." A protein termed PSMA-like protein, Genbank accession number AF261715, is nearly identical to amino acids 309-750 of PSMA and has a different expression profile. Thus the most preferred epitopes are those with an N-terminus located from amino acid 58 to 308.

PRAME, also know as MAPE, DAGE, and OIP4, was originally observed as a melanoma antigen. Subsequently, it has been recognized as a CT antigen, but unlike many CT antigens (e.g., MAGE, GAGE, and BAGE) it is expressed in acute myeloid leukemias. PRAME is a member of the MAPE family which consists largely of hypothetical proteins with which it shares limited sequence similarity. The usefulness of PRAME as a TuAA is taught in U.S. Patent 5,830,753 entitled "ISOLATED NUCLEIC ACID MOLECULES CODING FOR TUMOR REJECTION ANTIGEN PRECURSOR DAGE AND USES THEREOF."

PSA, prostate specific antigen, is a peptidase of the kallikrein family and a differentiation antigen of the prostate. Expression in breast tissue has also been reported. Alternate names include gamma-seminoprotein, kallikrein 3, seminogelase, seminin, and P-30 antigen. PSA has a high degree of sequence identity with the various alternate splicing products prostatic/glandular kallikrein-1 and -2, as well as kalikrein 4, which is also expressed in prostate and breast tissue. Other kallikreins generally share less sequence identity and have different expression profiles. Nonetheless, cross-reactivity that might be provoked by any particular epitope, along with the likelihood that that epitope would be liberated by processing in non-target tissues (most generally by the housekeeping proteasome), should be considered in designing a vaccine.

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PSCA, prostate stem cell antigen, and also known as SCAH-2, is a differentiation antigen preferentially expressed in prostate epithelial cells, and overexpressed in prostate cancers. Lower level expression is seen in some normal tissues including neuroendocrine cells of the digestive tract and collecting ducts of the kidney. PSCA is described in U.S. Patent 5,856,136 entitled "HUMAN STEM CELL ANTIGENS."

Synaptonemal complex protein 1 (SCP-1), also known as HOM-TES-14, is a meiosis-associated protein and also a cancer-testis antigen (Tureci, O., et al. Proc. Natl. Acad. Sci. USA 95:5211-5216, 1998). As a cancer antigen its expression is not cell-cycle regulated and it is found frequently in gliomas, breast, renal cell, and ovarian carcinomas. It has some similarity to myosins, but with few enough identities that cross-reactive epitopes are not an immediate prospect.

The ED-B domain of fibronectin is also a potential target. Fibronectin is subject to developmentally regulated alternative splicing, with the ED-B domain being encoded by a single exon that is used primarily in oncofetal tissues (Matsuura, H. and S. Hakomori *Proc. Natl. Acad. Sci. USA* 82:6517-6521, 1985; Carnemolla, B. et al. *J. Cell Biol.* 108:1139-1148, 1989; Loridon-Rosa, B. et al. *Cancer Res.*50:1608-1612, 1990; Nicolo, G. et al. *Cell Differ. Dev.* 32:401-408, 1990; Borsi, L. et al. *Exp. Cell Res.* 199:98-105, 1992; Oyama, F. et al. *Cancer Res.* 53:2005-2011, 1993; Mandel, U. et al. *APMIS* 102:695-702, 1994; Farnoud, M.R. et al. *Int. J. Cancer* 61:27-34, 1995; Pujuguet, P. et al. Am. J. Pathol. 148:579-592, 1996; Gabler, U. et al. *Heart* 75:358-362, 1996; Chevalier, X. *Br. J. Rheumatol.* 35:407-415, 1996; Midulla, M. *Cancer Res.* 60:164-169, 2000).

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The ED-B domain is also expressed in fibronectin of the neovasculature (Kaczmarek, J. et al. Int. J. Cancer 59:11-16, 1994; Castellani, P. et al. Int. J. Cancer 59:612-618, 1994; Neri, D. et al. Nat. Biotech. 15:1271-1275, 1997; Karelina, T.V. and A.Z. Eisen Cancer Detect. Prev. 22:438-444, 1998; Tarli, L. et al. Blood 94:192-198, 1999; Castellani, P. et al. Acta Neurochir. (Wien) 142:277-282, 2000). As an oncofetal domain, the ED-B domain is commonly found in the fibronectin expressed by neoplastic cells in addition to being expressed by the neovasculature. Thus, CTL-inducing vaccines targeting the ED-B domain can exhibit two mechanisms of action: direct lysis of tumor cells, and disruption of the tumor's blood supply through destruction of the tumor-associated neovasculature. As CTL activity can decay rapidly after withdrawal of vaccine, interference with normal angiogenesis can be minimal. The design and testing of vaccines targeted to neovasculature is described in Provisional U.S. Patent Application No. 60/274,063 entitled "ANTI-NEOVASCULATURE VACCINES FOR CANCER" and in U.S. Patent Application No. 10/094,699, attorney docket number CTLIMM.015A, entitled "ANTI-NEOVASCULATURE PREPARATIONS FOR CANCER, filed on date even with this application (March 7, 2002). A tumor cell line is disclosed in Provisional U.S. Application No. 60/363,131, filed on March 7, 2002, attorney docket number CTLIMM.028PR, entitled "HLA-TRANSGENIC MURINE TUMOR CELL LINE."

Carcinoembryonic antigen (CEA) is a paradigmatic oncofetal protein first described in 1965 (Gold and Freedman, J. Exp. Med. 121: 439-462, 1965. Fuller references can be found in the Online Medelian Inheritance in Man; record *114890). It has officially been renamed carcinoembryonic antigen-related cell adhesion molecule 5 (CEACAM5). Its expression is most strongly associated with adenocarcinomas of the epithelial lining of the digestive tract and in fetal colon. CEA is a member of the immunoglobulin supergene family and the defining member of the CEA subfamily.

Survivin, also known as Baculoviral IAP Repeat-Containing Protein 5 (BIRC5), is another protein with an oncofetal pattern of expression. It is a member of the inhibitor of apoptosis protein (IAP) gene family. It is widely overexpressed in cancers (Ambrosini, G. et al., *Nat. Med.* 3:917-921, 1997; Velculiscu V.E. et al., *Nat. Genet.* 23:387-388, 1999) and it's function as an inhibitor of apoptosis is believed to contribute to the malignant phenotype.

HER2/NEU is an oncogene related to the epidermal growth factor receptor (van de Vijver, et al., New Eng. J. Med. 319:1239-1245, 1988), and apparently identical to the c-ERBB2 oncogene (Di Fiore, et al., Science 237: 178-182, 1987). The over-expression of ERBB2 has been implicated in the neoplastic transformation of prostate cancer. As HER2 it is amplified and over-expressed in 25-30% of breast cancers among other tumors where expression level is correlated with the aggressiveness of the tumor (Slamon, et al., New Eng. J. Med. 344:783-792, 2001). A more detailed description is available in the Online Medelian Inheritance in Man; record *164870.

Useful epitopes were identified and tested as described in the following examples. However, these examples are intended for illustration purposes only, and should not be construed as limiting the scope of the invention in any way.

EXAMPLES

5 Example 1

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Manufacture of epitopes.

A. Synthetic production of epitopes

Peptides having an amino acid sequence of any of SEQ ID NO: 1, 8, 9, 11-23, 26-29, 32-44, 47-54, 56-63, 66-68, or 108-602 are synthesized using either FMOC or tBOC solid phase synthesis methodologies. After synthesis, the peptides are cleaved from their supports with either trifluoroacetic acid or hydrogen fluoride, respectively, in the presence of appropriate protective scavengers. After removing the acid by evaporation, the peptides are extracted with ether to remove the scavengers and the crude, precipitated peptide is then lyophilized. Purity of the crude peptides is determined by HPLC, sequence analysis, amino acid analysis, counterion content analysis and other suitable means. If the crude peptides are pure enough (greater than or equal to about 90% pure), they can be used as is. If purification is required to meet drug substance specifications, the peptides are purified using one or a combination of the following: reprecipitation; reverse-phase, ion exchange, size exclusion or hydrophobic interaction chromatography; or counter-current distribution.

20 Drug product formulation

GMP-grade peptides are formulated in a parenterally acceptable aqueous, organic, or aqueous-organic buffer or solvent system in which they remain both physically and chemically stable and biologically potent. Generally, buffers or combinations of buffers or combinations of buffers and organic solvents are appropriate. The pH range is typically between 6 and 9. Organic modifiers or other excipients can be added to help solubilize and stabilize the peptides. These include detergents, lipids, co-solvents, antioxidants, chelators and reducing agents. In the case of a lyophilized product, sucrose or mannitol or other lyophilization aids can be added. Peptide solutions are sterilized by membrane filtration into their final container-closure system and either lyophilized for dissolution in the clinic, or stored until use.

30 B. Construction of expression vectors for use as nucleic acid vaccines

The construction of three generic epitope expression vectors is presented below. The particular advantages of these designs are set forth in PCT Publication No. WO 01/82963 and U.S. Patent Application No. 09/561,572 entitled "EXPRESSION VECTORS ENCODING EPITOPES OF TARGET-ASSOCIATED ANTIGENS." Additional vectors strategies for their design are disclosed in PCT Publication WO 03/063770; U.S. Patent Application No. 10/292,413, filed on November 7, 2002; and Provisional U.S. Patent application No. 60/336,968 entitled

"EXPRESSION VECTORS ENCODING EPITOPES OF TARGET-ASSOCIATED ANTIGENS AND METHODS FOR THEIR DESIGN," filed on November 7, 2001. The teachings and embodiments disclosed in said PCT publications and applications are contemplated as supporting principals and embodiments related to and useful in connection with the present invention.

A suitable *E. coli* strain was then transfected with the plasmid and plated out onto a selective medium. Several colonies were grown up in suspension culture and positive clones were identified by restriction mapping. The positive clone was then grown up and aliquotted into storage vials and stored at -70°C.

A mini-prep (QIAprep Spin Mini-prep: Qiagen, Valencia, CA) of the plasmid was then made from a sample of these cells and automated fluorescent dideoxy sequence analysis was used to confirm that the construct had the desired sequence.

B.1 Construction of pVAX-EP1-IRES-EP2

Overview:

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The starting plasmid for this construct is pVAX1 purchased from Invitrogen (Carlsbad, CA). Epitopes EP1 and EP2 were synthesized by GIBCO BRL (Rockville, MD). The IRES was excised from pIRES purchased from Clontech (Palo Alto, CA).

Procedure:

- pIRES was digested with EcoRI and NotI. The digested fragments were separated by agarose gel electrophoresis, and the IRES fragment was purified from the excised band.
- 2. pVAX1 was digested with EcoRI and NotI, and the pVAX1 fragment was gel-purified.
- 3. The purified pVAX1 and IRES fragments were then ligated together.
- 4. Competent E. coli of strain DH5α were transformed with the ligation mixture.
- 5. Minipreps were made from 4 of the resultant colonies.
- Restriction enzyme digestion analysis was performed on the miniprep DNA. One
 recombinant colony having the IRES insert was used for further insertion of EP1 and
 EP2. This intermediate construct was called pVAX-IRES.
 - 7. Oligonucleotides encoding EP1 and EP2 were synthesized.
 - 8. EP1 was subcloned into pVAX-IRES between AfIII and EcoRI sites, to make pVAX-EP1-IRES;
 - EP2 was subcloned into pVAX-EP1-IRES between SalI and NotI sites, to make the final construct pVAX-EP1-IRES-EP2.
 - 10. The sequence of the EP1-IRES-EP2 insert was confirmed by DNA sequencing.

B 2. Construction of pVAX-EP1-IRES-EP2-ISS-NIS

Overview:

The starting plasmid for this construct was pVAX-EP1-IRES-EP2 (Example 1). The ISS (immunostimulatory sequence) introduced into this construct is AACGTT, and the NIS (standing for nuclear import sequence) used is the SV40 72bp repeat sequence. ISS-NIS was synthesized by GIBCO BRL. See Figure 2.

Procedure:

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- 1. pVAX-EP1-IRES-EP2 was digested with NruI; the linearized plasmid was gel-purified.
- 2. ISS-NIS oligonucleotide was synthesized.
- The purified linearized pVAX-EP1-IRES-EP2 and synthesized ISS-NIS were ligated together.
 - 4. Competent E. coli of strain DH5 α were transformed with the ligation product.
 - 5. Minipreps were made from resultant colonies.
 - 6. Restriction enzyme digestions of the minipreps were carried out.
- The plasmid with the insert was sequenced.

B3. Construction of pVAX-EP2-UB-EP1

Overview:

The starting plasmid for this construct was pVAX1 (Invitrogen). EP2 and EP1 were synthesized by GIBCO BRL. Wild type Ubiquitin cDNA encoding the 76 amino acids in the construct was cloned from yeast.

Procedure:

- 1. RT-PCR was performed using yeast mRNA. Primers were designed to amplify the complete coding sequence of yeast Ubiquitin.
- 2. The RT-PCR products were analyzed using agarose gel electrophoresis. A band with the predicted size was gel-purified.
- 3. The purified DNA band was subcloned into pZERO1 at EcoRV site. The resulting clone was named pZERO-UB.
- 4. Several clones of pZERO-UB were sequenced to confirm the Ubiquitin sequence before further manipulations.
- EP1 and EP2 were synthesized.
 - 6. EP2, Ubiquitin and EP1 were ligated and the insert cloned into pVAX1 between BamHI and EcoRI, putting it under control of the CMV promoter.
 - 7. The sequence of the insert EP2-UB-EP1 was confirmed by DNA sequencing.

WO 2004/022709

Example 2

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Identification of useful epitope variants.

The 10-mer FLPWHRLFLL (SEQ ID NO. 1) is identified as a useful epitope. Based on this sequence, numerous variants are made. Variants exhibiting activity in HLA binding assays (see Example 3, section 6) are identified as useful, and are subsequently incorporated into vaccines. Variants that increase the stability of binding, assayed can be particularly usefule, for example as described in WO 97/41440 entitled "Methods for Selecting and Producing T Cell Peptide Epitopes and Vaccines Incorporating Said Selected Epitopes." The teachings and embodiments disclosed in said PCT publication are contemplated as supporting principals and embodiments related to and useful in connection with the present invention.

PCT/US2003/027706

The HLA-A2 binding of length variants of FLPWHRLFLL have been evaluated. Proteasomal digestion analysis indicates that the C-terminus of the 9-mer FLPWHRLFL (SEQ ID NO. 8) is also produced. Additionally the 9-mer LPWHRLFLL (SEQ ID NO. 9) can result from N-terminal trimming of the 10-mer. Both are predicted to bind to the HLA-A*0201 molecule, however of these two 9-mers, FLPWHRLFL displayed more significant binding and is preferred (see Figs. 3A and B).

In vitro proteasome digestion and N-terminal pool sequencing indicates that tyrosinase₂₀₇₋₂₁₆ (SEQ ID NO. 1) is produced more commonly than tyrosinase₂₀₇₋₂₁₅ (SEQ ID NO. 8), however the latter peptide displays superior immunogenicity, a potential concern in arriving at an optimal vaccine design. FLPWHRLFL, tyrosinase₂₀₇₋₂₁₅ (SEQ ID NO. 8) was used in an in vitro immunization of HLA-A2⁺ blood to generate CTL (see CTL Induction Cultures below). Using peptide pulsed T2 cells as targets in a standard chromium release assay it was found that the CTL induced by tyrosinase₂₀₇₋₂₁₅ (SEQ ID NO. 8) recognize tyrosinase₂₀₇₋₂₁₆ (SEQ ID NO. 1) targets equally well (see fig. 3C). These CTL also recognize the HLA-A2⁺, tyrosinase⁺ tumor cell lines 624.38 and HTB64, but not 624.28 an HLA-A2⁻ derivative of 624.38 (fig. 3C). Thus the relative amounts of these two epitopes produced in vivo, does not become a concern in vaccine design. CTL induction cultures

PBMCs from normal donors were purified by centrifugation in Ficoll-Hypaque from buffy coats. All cultures were carried out using the autologous plasma (AP) to avoid exposure to potential xenogeneic pathogens and recognition of FBS peptides. To favor the in vitro generation of peptide-specific CTL, we employed autologous dendritic cells (DC) as APCs. DC were generated and CTL were induced with DC and peptide from PBMCs as described (Keogh et al., 2001). Briefly, monocyte-enriched cell fractions were cultured for 5 days with GM-CSF and IL-4 and were cultured for 2 additional days in culture media with 2 μg/ml CD40 ligand to induce maturation. 2 x10⁶ CD8+-enriched T lymphocytes/well and 2 x10⁵ peptide-pulsed DC/well were co-cultured in 24-well plates in 2 ml RPMI supplemented with 10% AP, 10 ng/ml IL-7 and 20

IU/ml IL-2. Cultures were restimulated on days 7 and 14 with autologous irradiated peptide-pulsed DC.

Sequence variants of FLPWHRLFL are constructed as follow. Consistent with the binding coefficient table (see Table 3) from the NIH/BIMAS MHC binding prediction program (see reference in example 3 below), binding can be improved by changing the L at position 9, an anchor position, to V. Binding can also be altered, though generally to a lesser extent, by changes at non-anchor positions. Referring generally to Table 3, binding can be increased by employing residues with relatively larger coefficients. Changes in sequence can also alter immunogenicity independently of their effect on binding to MHC. Thus binding and/or immunogenicity can be improved as follows:

By substituting F,L,M,W, or Y for P at position 3; these are all bulkier residues that can also improve immunogenicity independent of the effect on binding. The amine and hydroxylbearing residues, Q and N; and S and T; respectively, can also provoke a stronger, cross-reactive response.

By substituting D or E for W at position 4 to improve binding; this addition of a negative charge can also make the epitope more immunogenic, while in some cases reducing cross-reactivity with the natural epitope. Alternatively the conservative substitutions of F or Y can provoke a cross-reactive response.

By substituting F for H at position 5 to improve binding. H can be viewed as partially charged, thus in some cases the loss of charge can hinder cross-reactivity. Substitution of the fully charged residues R or K at this position can enhance immunogenicity without disrupting charge-dependent cross-reactivity.

By substituting I, L, M, V, F, W, or Y for R at position 6. The same caveats and alternatives apply here as at position 5.

By substituting W or F for L at position 7 to improve binding. Substitution of V, I, S, T, Q, or N at this position are not generally predicted to reduce binding affinity by this model (the NIH algorithm), yet can be advantageous as discussed above.

Y and W, which are equally preferred as the Fs at positions 1 and 8, can provoke a useful cross-reactivity. Finally, while substitutions in the direction of bulkiness are generally favored to improve immunogenicity, the substitution of smaller residues such as A, S, and C, at positions 3-7 can be useful according to the theory that contrast in size, rather than bulkiness per se, is an important factor in immunogenicity. The reactivity of the thiol group in C can introduce other properties as discussed in Chen, J.-L., et al. *J. Immunol.* 165:948-955, 2000.

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Table 3. 9-mer Coefficient Table for HLA-A*0201*

HLA Coeffic	ient table	for file "A	0201	standard"					
Amino Acid								T	T
Туре	1 st	2 nd	_3rd	4th	5th	6th	7th	8th	9th
A	1.000	1.000	1.000	1.000	1.000	1.000	1.000	1.000	1.000
C	1.000	0.470	1.000	1.000	1.000	1.000	1.000	1.000	1.000
D	0.075	0.100	0.400	4.100	1.000	1.000	0.490	1.000	0.003
E	0.075	1.400	0.064	4.100	1.000	1.000	0.490	1.000	0.003
F	4.600	0.050	3.700	1.000	3.800	1.900	5.800	5.500	0.015
G	1.000	0.470	1.000	1.000	1.000	1.000	0.130	1.000	0.015
H	0.034	0.050	1.000	1.000	1.000	1.000	1.000	1.000	0.015
I	1.700	9.900	1.000	1.000	1.000	2.300	1.000	0.410	2.100
K	3.500	0.100	0.035	1.000	1.000	1.000	1.000	1.000	0.003
L	1.700	72.000	3.700	1.000	1.000	2.300	1.000	1.000	4.300
M	1.700	52.000	3.700	1.000	1.000	2.300	1.000	1.000	1.000
N	1.000	0.470	1.000	1.000	1.000	1.000	1.000	1.000	0.015
P	0.022	0.470	1.000	1.000	1.000	1.000	1.000	1.000	0.003
Q	1.000	7.300	1.000	1.000	1.000	1.000	1.000	1.000	0.003
R	1.000	0.010	0.076	1.000	1.000	1.000	0.200	1.000	0.003
S	1.000	0.470	1.000	1.000	1.000	1.000	1.000	1.000	0.015
T	1.000	1.000	1.000	1.000	1.000	1.000	1.000	1.000	1.500
V	1.700	6.300	1.000	1.000	1.000	2.300	1.000	0.410	14.000
W	4.600	0.010	8.300	1.000	1.000	1.700	7.500	5.500	0.015
Y	4.600	0.010	3.200	1.000	1.000	1.500	1.000	5.500	0.015

^{*}This table and other comparable data that are publicly available are useful in designing epitope variants and in determining whether a particular variant is substantially similar, or is functionally similar.

Example 3

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Cluster Analysis (SSX-231-68).

1. Epitope cluster region prediction:

The computer algorithms: SYFPEITHI (internet http:// access at syfpeithi.bmi-heidelberg.com/Scripts/MHCServer.dll/EpPredict.htm), based on the book "MHC Ligands and Peptide Motifs" by H.G.Rammensee, J.Bachmann and S.Stevanovic; and HLA Peptide Binding Predictions (NIH) (internet http:// access at bimas.dcrt.nih.gov/molbio/hla_bin), described in Parker, K. C., et al., J. Immunol. 152:163, 1994; were used to analyze the protein sequence of SSX-2 (GI:10337583). Epitope clusters (regions with higher than average density of peptide fragments with high predicted MHC affinity) were defined as described fully in U.S. Patent Application No. 09/561,571 entitled "EPITOPE CLUSTERS," filed on April 28, 2000. Using a epitope density ratio cutoff of 2, five and two clusters were defined using the SYFPETHI and NIH algorithms, respectively, and peptides score cutoffs of 16 (SYFPETHI) and 5 (NIH). The highest scoring peptide with the NIH algorithm, SSX-241-49, with an estimated halftime of dissociation of

>1000 min., does not overlap any other predicted epitope but does cluster with SSX- 2_{57-65} in the NIH analysis.

2. <u>Peptide synthesis and characterization:</u>

SSX-2₃₁₋₆₈, YFSKEEWEKMKASEKIFYVYMKRKYEAMTKLGFKATLP (SEQ ID NO. 10) was synthesized by MPS (Multiple Peptide Systems, San Diego, CA 92121) using standard solid phase chemistry. According to the provided 'Certificate of Analysis', the purity of this peptide was 95%.

3. Proteasome digestion:

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Proteasome was isolated from human red blood cells using the proteasome isolation protocol described in PCT Publication No. WO 01/82963 and U.S. Patent Application No. 09/561,074 entitled "METHOD OF EPITOPE DISCOVERY," filed on April 28, 2000. The teachings and embodiments disclosed in said PCT publication and application are contemplated as supporting principals and embodiments related to and useful in connection with the present invention. SDS-PAGE, western-blotting, and ELISA were used as quality control assays. The final concentration of proteasome was 4 mg/ml, which was determined by non-interfering protein assay (Geno Technologies Inc.). Proteasomes were stored at -70°C in 25 µl aliquots.

SSX- 2_{31-68} was dissolved in Milli-Q water, and a 2 mM stock solution prepared and $20\mu L$ aliquots stored at -20°C.

1 tube of proteasome (25 μ L) was removed from storage at-70°C and thawed on ice. It was then mixed thoroughly with 12.5 μ L of 2mM peptide by repipetting (samples were kept on ice). A 5 μ L sample was immediately removed after mixing and transferred to a tube containing 1.25 μ L 10%TFA (final concentration of TFA was 2%); the T=0 min sample. The proteasome digestion reaction was then started and carried out at 37°C in a programmable thermal controller. Additional 5 μ L samples were taken out at 15, 30, 60, 120, 180 and 240 min respectively, the reaction was stopped by adding the sample to 1.25 μ L 10% TFA as before. Samples were kept on ice or frozen until being analyzed by MALDI-MS. All samples were saved and stored at -20°C for HPLC analysis and N-terminal sequencing. Peptide alone (without proteasome) was used as a blank control: 2 μ L peptide + 4 μ L Tris buffer (20 mM, pH 7.6) + 1.5 μ L TFA.

4. MALDI-TOF MS measurements:

For each time point 0.3 μ L of matrix solution (10mg/ml α -cyano-4-hydroxycinnamic acid in AcCN/H₂O (70:30)) was first applied on a sample slide, and then an equal volume of digested sample was mixed gently with matrix solution on the slide. The slide was allowed to dry at ambient air for 3-5 min. before acquiring the mass spectra. MS was performed on a Lasermat 2000 MALDI-TOF mass spectrometer that was calibrated with peptide/protein standards. To improve the accuracy of measurement, the molecular ion weight (MH) of the peptide substrate was used as

an internal calibration standard. The mass spectrum of the T=120 min. digested sample is shown in figure 4.

5. MS data analysis and epitope identification:

To assign the measured mass peaks, the computer program MS-Product, a tool from the UCSF Mass Spectrometry Facility (http:// accessible at prospector.ucsf.edu/ucsfhtml3.4/msprod.htm), was used to generate all possible fragments (N- and C-terminal ions, and internal fragments) and their corresponding molecular weights. Due to the sensitivity of the mass spectrometer, average molecular weight was used. The mass peaks observed over the course of the digestion were identified as summarized in Table 4.

Fragments co-C-terminal with 8-10 amino acid long sequences predicted to bind HLA by the SYFPEITHI or NIH algorithms were chosen for further study. The digestion and prediction steps of the procedure can be usefully practiced in any order. Although the substrate peptide used in proteasomal digest described here was specifically designed to include predicted HLA-A2.1 binding sequences, the actual products of digestion can be checked after the fact for actual or predicted binding to other MHC molecules. Selected results are shown in Table 5.

Table 4. SSX-231-68 Mass Peak Identification.

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MS PEAK	PEPTIDE	SEQUENCE	CALCULATED
(measured)	İ		MASS (MH ⁺)
988.23	31-37	YFSKEEW	989.08
1377.68±2.38	31-40	YFSKEEWEKM	1377.68
1662.45±1.30	31-43	YFSKEEWEKMKAS	1663.90
2181.72±0.85	31-47	YFSKEEWEKMKASEKIF	2181.52
2346.6	31-48	YFSKEEWEKMKASEKIFY	2344.71
1472.16±1.54	38-49	EKMKASEKIFYV	1473.77
2445.78±1.18	31-49*	YFSKEEWEKMKASEKIFYV	2443.84
2607.	31-50	YFSKEEWEKMKASEKIFYVY	2607.02
1563.3	50-61	YMKRKYEAMTKL	1562.93
3989.9	31-61	YFSKEEWEKMKASEKIFYVYMKRKYEAMTKL	3987.77
1603.74±1.53	51-63	MKRKYEAMTKLGF	1603.98
1766.45±1.5	50-63	YMKRKYEAMTKLGF	1767.16
1866.32±1.22	49-63	VYMKRKYEAMTKLGF	1866.29
4192.6	31-63	YFSKEEWEKMKASEKIFYVYMKRKYEAMTKLGF	4192.00
4392.1	31-65**	YFSKEEWEKMKASEKIFYVYMKRKYEAMTKLGFKA	4391.25

Boldface sequence correspond to peptides predicted to bind to MHC.

^{*} On the basis of mass alone this peak could also have been assigned to the peptide 32-50, however proteasomal removal of just the N-terminal amino acid is unlikely. N-terminal sequencing (below) verifies the assignment to 31-49.

^{**} On the basis of mass this fragment might also represent 33-68. N-terminal sequencing below is consistent with the assignment to 31-65.

Table 5. Predicted HLA binding by proteasomally generated fragments

SEQ ID NO.	PEPTIDE	HLA	SYFPEITHI	NIH
11	FSKEEWEKM	B*3501	NP†	90
12	KMKASEKIF	B*08	17	<5
13 & (14)	(K) MKASEKIFY	A1	19 (19)	<5
15 &(16)	(M) KASEKIFYV	A*0201	22 (16)	1017
	ĺ	B*08	17	<5
		B*5101	22 (13)	60
		B*5102	NP	133
		B*5103	NP	121
17 & (18)	(K) ASEKIFYVY	A1	34 (19)	14
19 & (20)	(K) RKYEAMTKL	A*0201	15	<5
		A26	15	NP
	İ	B14	NP	45 (60)
		B*2705	21	15
		B*2709	16	NP
		B*5101	15	<5
21	KYEAMTKLGF	A1	16	<5
22		A24	NP	300
22	YEAMTKLGF	B*4403	NP	80
23	EAMTKLGF	B*08	22	<5

†No prediction

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As seen in Table 5, N-terminal addition of authentic sequence to epitopes can generate epitopes for the same or different MHC restriction elements. Note in particular the pairing of (K)RKYEAMTKL (SEQ ID NOS 19 and (20)) with HLA-B14, where the 10-mer has a longer predicted halftime of dissociation than the co-C-terminal 9-mer. Also note the case of the 10-mer KYEAMTKLGF (SEQ ID NO. 21) which can be used as a vaccine useful with several MHC types by relying on N-terminal trimming to create the epitopes for HLA-B*4403 and -B*08.

6. <u>HLA-A0201 binding assay:</u>

Binding of the candidate epitope KASEKIFYV, SSX-2₄₁₋₄₉, (SEQ ID NO. 15) to HLA-A2.1 was assayed using a modification of the method of Stauss et al., (Proc Natl Acad Sci USA 89(17):7871-5 (1992)). Specifically, T2 cells, which express empty or unstable MHC molecules on their surface, were washed twice with Iscove's modified Dulbecco's medium (IMDM) and cultured overnight in serum-free AIM-V medium (Life Technologies, Inc., Rockville, MD) supplemented with human β2-microglobulin at 3μg/ml (Sigma, St. Louis, MO) and added peptide, at 800, 400, 200, 100, 50, 25, 12.5, and 6.25 μg/ml.in a 96-well flat-bottom plate at 3x10⁵ cells/200 μl (microliter)/well. Peptide was mixed with the cells by repipeting before distributing to the plate (alternatively peptide can be added to individual wells), and the plate was rocked gently for 2 minutes. Incubation was in a 5% CO₂ incubator at 37°C. The next day the unbound peptide was removed by washing twice with serum free RPMI medium and a saturating amount of anti-class I HLA monoclonal antibody, fluorescein isothiocyanate (FITC)-conjugated anti-HLA A2, A28 (One

Lambda, Canoga Park, CA) was added. After incubation for 30 minutes at 4°C, cells were washed 3 times with PBS supplemented with 0.5% BSA, 0.05%(w/v) sodium azide, pH 7.4-7.6 (staining buffer). (Alternatively W6/32 (Sigma) can be used as the anti-class I HLA monoclonal antibody the cells washed with staining buffer and then incubated with fluorescein isothiocyanate (FITC)-conjugated goat F(ab') antimouse-IgG (Sigma) for 30 min at 4°C and washed 3 times as before.) The cells were resuspended in 0.5 ml staining buffer. The analysis of surface HLA-A2.1 molecules stabilized by peptide binding was performed by flow cytometry using a FACScan (Becton Dickinson, San Jose, CA). If flow cytometry is not to be performed immediately the cells can be fixed by adding a quarter volume of 2% paraformaldehyde and storing in the dark at 4°C.

The results of the experiment are shown in Figure 5. SSX-2₄₁₋₄₉ (SEQ ID NO. 15) was found to bind HLA-A2.1 to a similar extent as the known A2.1 binder FLPSDYFPSV (HBV₁₈₋₂₇; SEQ ID NO: 24) used as a positive control. An HLA-B44 binding peptide, AEMGKYSFY (SEQ ID NO: 25), was used as a negative control. The fluoresence obtained from the negative control was similar to the signal obtained when no peptide was used in the assay. Positive and negative control peptides were chosen from Table 18.3.1 in *Current Protocols in Immunology* p. 18.3.2, John Wiley and Sons, New York, 1998.

7. <u>Immunogenicity:</u>

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A. In vivo immunization of mice.

HHD1 transgenic A*0201 mice (Pascolo, S., et al. *J. Exp. Med.* 185:2043-2051, 1997) were anesthetized and injected subcutaneously at the base of the tail, avoiding lateral tail veins, using 100 μ l containing 100 nmol of SSX-2₄₁₋₄₉ (SEQ ID NO. 15) and 20 μ g of HTL epitope peptide in PBS emulsified with 50 μ l of IFA (incomplete Freund's adjuvant).

B. <u>Preparation of stimulating cells (LPS blasts).</u>

Using spleens from 2 naive mice for each group of immunized mice, un-immunized mice were sacrificed and the carcasses were placed in alcohol. Using sterile instruments, the top dermal layer of skin on the mouse's left side (lower mid-section) was cut through, exposing the peritoneum. The peritoneum was saturated with alcohol, and the spleen was aseptically extracted. The spleen was placed in a petri dish with serum-free media. Splenocytes were isolated by using sterile plungers from 3 ml syringes to mash the spleens. Cells were collected in a 50 ml conical tubes in serum-free media, rinsing dish well. Cells were centrifuged (12000 rpm, 7 min) and washed one time with RPMI. Fresh spleen cells were resuspended to a concentration of 1x106 cells per ml in RPMI-10%FCS (fetal calf serum). 25g/ml lipopolysaccharide and 7 μg/ml Dextran Sulfate were added. Cell were incubated for 3 days in T-75 flasks at 37°C, with 5% CO₂. Splenic blasts were collected in 50 ml tubes pelleted (12000 rpm, 7 min) and resuspended to 3X10⁷/ml in RPMI. The blasts were pulsed with the priming peptide at 50 μg/ml, RT 4hr. mitomycin C-treated at 25μg/ml, 37⁰C, 20 min and washed three times with DMEM.

C. In vitro stimulation.

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3 days after LPS stimulation of the blast cells and the same day as peptide loading, the primed mice were sacrificed (at 14 days post immunization) to remove spleens as above. $3x10^6$ splenocytes were co-cultured with $1x10^6$ LPS blasts/well in 24-well plates at 37° C, with 5% CO₂ in DMEM media supplemented with 10% FCS, $5x10^{-5}$ M β -mercaptoethanol, $100\mu g/ml$ streptomycin and 100 IU/ml penicillin. Cultures were fed 5% (vol/vol) ConA supernatant on day 3 and assayed for cytolytic activity on day 7 in a 51Cr-release assay.

PCT/US2003/027706

D. <u>Chromium-release assay measuring CTL activity.</u>

To assess peptide specific lysis, $2x10^6$ T2 cells were incubated with 100 μ Ci sodium chromate together with 50 μ g/ml peptide at 37°C for 1 hour. During incubation they were gently shaken every 15 minutes. After labeling and loading, cells were washed three times with 10 ml of DMEM-10% FCS, wiping each tube with a fresh Kimwipe after pouring off the supernatant. Target cells were resuspended in DMEM-10% FBS $1x10^5$ /ml. Effector cells were adjusted to $1x10^7$ /ml in DMEM-10% FCS and 100 μ l serial 3-fold dilutions of effectors were prepared in U-bottom 96-well plates. 100 μ l of target cells were added per well. In order to determine spontaneous release and maximum release, six additional wells containing 100 μ l of target cells were prepared for each target. Spontaneous release was revealed by incubating the target cells with 100 μ l medium; maximum release was revealed by incubating the target cells with 100 μ l medium; maximum release was revealed by incubated for 4 hours at 37°C in 5% CO₂ and 80% humidity. After the incubation, plates were then centrifuged for 5 min at 1200 rpm. Supernatants were harvested and counted using a gamma counter. Specific lysis was determined as follows: % specific release = [(experimental release - spontaneous release)] x 100.

Results of the chromium release assay demonstrating specific lysis of peptide pulsed target cells are shown in figure 6.

8. <u>Cross-reactivity with other SSX proteins:</u>

SSX- 2_{41-49} (SEQ ID NO. 15) shares a high degree of sequence identity with the same region of the other SSX proteins. The surrounding regions have also been generally well conserved. Thus the housekeeping proteasome can cleave following V_{49} in all five sequences. Moreover, SSX₄₁₋₄₉ is predicted to bind HLA-A*0201 (see Table 6). CTL generated by immunization with SSX- 2_{41-49} cross-react with tumor cells expressing other SSX proteins.

Table 6. SSX₄₁₋₄₉ - A*0201 Predicted Binding

SEQ ID NO.	Family Member	Sequence	SYFPEITHI Score	NIH Score
15	SSX-2	KASEKIFYV	22	1017
26	SSX-1	KYSEKISYV	18	1017
27	SSX-3	KVSEKIVYV	24	1.7
28	SSX-4	KSSEKIVYV	20	1105
29	SSX-5	KASEKIIYV	22	82 175

Example 4

Cluster Analysis (PSMA₁₆₃₋₁₉₂).

[0227] A peptide, AFSPQGMPEGDLVYVNYARTEDFFKLERDM, PSMA₁₆₃₋₁₉₂, (SEQ ID NO. 30), containing an A1 epitope cluster from prostate specific membrane antigen, PSMA₁₆₈₋₁₉₀ (SEQ ID NO. 31) was synthesized using standard solid-phase F-moc chemistry on a 433A ABI Peptide synthesizer. After side chain deprotection and cleavage from the resin, peptide first dissolved in formic acid and then diluted into 30% Acetic acid, was run on a reverse-phase preparative HPLC C4 column at following conditions: linear AB gradient (5% B/min) at a flow rate of 4 ml/min, where eluent A is 0.1% aqueous TFA and eluent B is 0.1% TFA in acetonitrile. A fraction at time 16.642 min containing the expected peptide, as judged by mass spectrometry, was pooled and lyophilized. The peptide was then subjected to proteasome digestion and mass spectrum analysis essentially as described above. Prominent peaks from the mass spectra are summarized in Table 7.

Table 7. PSMA₁₆₃₋₁₉₂ Mass Peak Identification.

PEPTIDE	SEQUENCE	CALCULATED
163-177	AECDOCAPAGE	MASS (MH ⁺)
	AFSPQGMPEGDLVYV	1610.0
178-189	NYARTEDFFKLE	1533.68
170-189	PEGDLVYVNYARTEDFFKLE	2406.66
178-191	NYARTEDFFKLERD	1804.95
170-191	PEGDLVYVNYARTEDFFKLERD	
178-192	NYARTEDFFKLERDM	2677.93
163-176	AFSPQGMPEGDLVY	1936.17
177-192		1511.70
163-179	VNYARTEDFFKLERDM	2035.30
	AFSPQGMPEGDLVYVNY	1888.12
180-192	ARTEDFFKLERDM	1658.89
163-183	AFSPQGMPEGDLVYVNYARTE	2345.61
184-192	DFFKLERDM	
176-192	YVNYARTEDFFKLERDM	1201.40
167-185	QGMPEGDLVYVNYARTEDF	2198.48
178-186		2205.41
2.0 100	NYARTEDFF	1163.22

Boldface sequences correspond to peptides predicted to bind to MHC, see Table 8.

N-terminal Pool Sequence Analysis

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One aliquot at one hour of the proteasomal digestion (see Example 3 part 3 above) was subjected to N-terminal amino acid sequence analysis by an ABI 473A Protein Sequencer (Applied Biosystems, Foster City, CA). Determination of the sites and efficiencies of cleavage was based on consideration of the sequence cycle, the repetitive yield of the protein sequencer, and the relative yields of amino acids unique in the analyzed sequence. That is if the unique (in the analyzed sequence) residue X appears only in the nth cycle a cleavage site exists n-1 residues before it in the N-terminal direction. In addition to helping resolve any ambiguity in the assignment of mass to sequences, these data also provide a more reliable indication of the relative yield of the various fragments than does mass spectrometry.

For PSMA₁₆₃₋₁₉₂ (SEQ ID NO. 30) this pool sequencing supports a single major cleavage site after V_{177} and several minor cleavage sites, particularly one after Y_{179} . Reviewing the results presented in figures 7A-C reveals the following:

S at the 3rd cycle indicating presence of the N-terminus of the substrate.

Q at the 5th cycle indicating presence of the N-terminus of the substrate.

N at the 1st cycle indicating cleavage after V₁₇₇.

N at the 3^{rd} cycle indicating cleavage after V_{175} . Note the fragment 176-192 in Table 7.

T at the 5th cycle indicating cleavage after V₁₇₇.

T at the 1^{st} – 3^{rd} cycles, indicating increasingly common cleavages after R_{181} , A_{180} and Y_{179} . Only the last of these correspond to peaks detected by mass spectrometry; 163-179 and 180-192, see Table 7. The absence of the others can indicate that they are on fragments smaller than were examined in the mass spectrum.

K at the 4th, 8th, and 10th cycles indicating cleavages after E₁₈₃, Y₁₇₉, and V₁₇₇, respectively, all of which correspond to fragments observed by mass spectroscopy. See Table 7.

A at the 1st and 3rd cycles indicating presence of the N-terminus of the substrate and cleavage after V₁₇₇, respectively.

P at the 4th and 8th cycles indicating presence of the N-terminus of the substrate.

G at the 6th and 10th cycles indicating presence of the N-terminus of the substrate.

M at the 7th cycle indicating presence of the N-terminus of the substrate and/or cleavage after F₁₈₅.

M at the 15th cycle indicating cleavage after V₁₇₇.

The 1st cycle can indicate cleavage after D₁₉₁, see Table 7.

R at the 4th and 13th cycle indicating cleavage after V₁₇₇.

R at the 2nd and 11th cycle indicating cleavage after Y₁₇₉.

V at the 2nd, 6th, and 13th cycle indicating cleavage after V₁₇₅, M₁₆₉ and presence of the N-terminus of the substrate, respectively. Note fragments beginning at 176 and 170 in Table 7.

Y at the 1st, 2nd, and 14th cycles indicating cleavage after V_{175} , V_{177} , and presence of the N-terminus of the substrate, respectively.

L at the 11th and 12th cycles indicating cleavage after V_{177} , and presence of the N-terminus of the substrate, respectively, is the interpretation most consistent with the other data. Comparing to the mass spectrometry results we see that L at the 2nd, 5th, and 9th cycles is consistent with cleavage after F_{186} , E_{183} or M_{169} , and Y_{179} , respectively. See Table 7.

Epitope Identification

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Fragments co-C-terminal with 8-10 amino acid long sequences predicted to bind HLA by the SYFPEITHI or NIH algorithms were chosen for further analysis. The digestion and prediction steps of the procedure can be usefully practiced in any order. Although the substrate peptide used in proteasomal digest described here was specifically designed to include a predicted HLA-A1 binding sequence, the actual products of digestion can be checked after the fact for actual or predicted binding to other MHC molecules. Selected results are shown in Table 8.

15 Table 8. Predicted HLA binding by proteasomally generated fragments

SEQ ID NO	PEPTIDE	HLA	SYFPEITHI	NIH
32 & (33)	(G) MPEGDLVYV	A*0201	17 (27)	(2605)
		B*0702	20	<5
		B*5101	22	314
34 & (35)	(Q) GMPEGDLVY	A1	24 (26)	<5
		A3	16 (18)	36
		B*2705	17	25
36	MPEGDLVY	B*5101	15	NP†
37 & (38)	(P) EGDLVYVNY	A1	27 (15)	12
		A26	23 (17)	NP
39	LVYVNYARTE	A3	21	<5
40 & (41)	(Y) VNYARTEDF	A26	(20)	NP
		B*08	15	<5
		B*2705	12	50
42	NYARTEDFF	A24	NP†	100
		Cw*0401	NP	120
43	YARTEDFF	B*08	16	<5
44	RTEDFFKLE	A1	21	<5
4N7. 1		A26	15	NP

†No prediction

HLA-A*0201 binding assay:

HLA-A*0201 binding studies were preformed with PSMA₁₆₈₋₁₇₇, GMPEGDLVYV, (SEQ ID NO. 33) essentially as described in Example 3 above. As seen in figure 8, this epitope exhibits significant binding at even lower concentrations than the positive control peptides. The Melan-A peptide used as a control in this assay (and throughout this disclosure), ELAGIGILTV, is actually a variant of the natural sequence (EAAGIGILTV) and exhibits a high affinity in this assay.

Example 5

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Cluster Analysis (PSMA281-310).

Another peptide, RGIAEAVGLPSIPVHPIGYYDAQKLLEKMG, PSMA₂₈₁₋₃₁₀, (SEQ ID NO. 45), containing an A1 epitope cluster from prostate specific membrane antigen, PSMA₂₈₃₋₃₀₇ (SEQ ID NO. 46), was synthesized using standard solid-phase F-moc chemistry on a 433A ABI Peptide synthesizer. After side chain deprotection and cleavage from the resin, peptide in ddH2O was run on a reverse-phase preparative HPLC C18 column at following conditions: linear AB gradient (5% B/min) at a flow rate of 4 ml/min, where eluent A is 0.1% aqueous TFA and eluent B is 0.1% TFA in acetonitrile. A fraction at time 17.061 min containing the expected peptide as judged by mass spectrometry, was pooled and lyophilized. The peptide was then subjected to proteasome digestion and mass spectrum analysis essentially as described above. Prominent peaks from the mass spectra are summarized in Table 9.

Table 9. PSMA₂₈₁₋₃₁₀ Mass Peak Identification.

PEPTIDE	SEQUENCE	CALCULATE D MASS (MH ⁺)
281-297	RGIAEAVGLPSIPVHPI*	1727.07
286-297	AVGLPSIPVHPI**	1200.46
287-297	VGLPSIPVHPI	1129.38
288-297	GLPSIPVHPI [†]	1030.25
298-310	GYYDAQKLLEKMG‡	1516.5
298-305	GYYDAQKLS	958.05
281-305	RGIAEAVGLPSIPVHPIGYYDAQKL	2666.12
281-307	RGIAEAVGLPSIPVHPIGYYDAQKLLE	2908.39
286-307	AVGLPSIPVHPIGYYDAQKLLE¶	2381.78
287-307	VGLPSIPVHPIGYYDAQKLLE	2310.70
288-307	GLPSIPVHPIGYYDAQKLLE#	2211.57
281-299	RGIAEAVGLPSIPVHPIGY	1947
286-299	AVGLPSIPVHPIGY	1420.69
287-299	VGLPSIPVHPIGY	1349.61
288-299	GLPSIPVHPIGY	1250.48
287-310	VGLPSIPVHPIGYYDAQKLLEKMG	2627.14
288-310	GLPSIPVHPIGYYDAQKLLEKMG	2528.01

Boldface sequences correspond to peptides predicted to bind to MHC, see Table 10.

† By mass alone this peak could also have been 289-298.

None of these alternate assignments are supported N-terminal pool sequence analysis.

^{*}By mass alone this peak could also have been 296-310 or 288-303.

^{**}By mass alone this peak could also have been 298-307. Combination of HPLC and mass spectrometry show that at some later time points this peak is a mixture of both species.

[?] By mass alone this peak could also have been 281-295 or 294-306.

[§] By mass alone this peak could also have been 297-303.

[¶] By mass alone this peak could also have been 285-306.

[#] By mass alone this peak could also have been 288-303.

N-terminal Pool Sequence Analysis

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One aliquot at one hour of the proteasomal digestion (see Example 3 part 3 above) was subjected to N-terminal amino acid sequence analysis by an ABI 473A Protein Sequencer (Applied Biosystems, Foster City, CA). Determination of the sites and efficiencies of cleavage was based on consideration of the sequence cycle, the repetitive yield of the protein sequencer, and the relative yields of amino acids unique in the analyzed sequence. That is if the unique (in the analyzed sequence) residue X appears only in the nth cycle a cleavage site exists n-1 residues before it in the N-terminal direction. In addition to helping resolve any ambiguity in the assignment of mass to sequences, these data also provide a more reliable indication of the relative yield of the various fragments than does mass spectrometry.

For PSMA₂₈₁₋₃₁₀ (SEQ ID NO. 45) this pool sequencing supports two major cleavage sites after V₂₈₇ and I₂₉₇ among other minor cleavage sites. Reviewing the results presented in Fig. 9 reveals the following:

S at the 4^{th} and 11^{th} cycles indicating cleavage after V_{287} and presence of the N-terminus of the substrate, respectively.

H at the 8^{th} cycle indicating cleavage after V_{287} . The lack of decay in peak height at positions 9 and 10 versus the drop in height present going from 10 to 11 can suggest cleavage after A_{286} and E_{285} as well, rather than the peaks representing latency in the sequencing reaction.

D at the 2nd, 4th, and 7th cycles indicating cleavages after Y₂₉₉, I₂₉₇, and V₂₉₄, respectively. This last cleavage is not observed in any of the fragments in Table 10 or in the alternate assignments in the notes below.

Q at the 6th cycle indicating cleavage after I297.

M at the 10th and 12th cycle indicating cleavages after Y₂₉₉ and I₂₉₇, respectively. Epitope Identification

Fragments co-C-terminal with 8-10 amino acid long sequences predicted to bind HLA by the SYFPEITHI or NIH algorithms were chosen for further study. The digestion and prediction steps of the procedure can be usefully practiced in any order. Although the substrate peptide used in proteasomal digest described here was specifically designed to include a predicted HLA-A1 binding sequence, the actual products of digestion can be checked after the fact for actual or predicted binding to other MHC molecules. Selected results are shown in Table 10.

<u>Table 10.</u>

<u>Predicted HLA binding by proteasomally generated fragments: PSMA₂₈₁₋₃₁₀</u>

SEQ ID NO.	PEPTIDE	HLA	SYFPEITHI	NIH
47 & (48)	(G) LPSIPVH PI	A*0201	16 (24)	(24)
		B*0702/B7	23	12
		B*5101	24	572
		Cw*0401	NP†	20
49 & (50)	(P) IGYYDAQ KL	A*0201	(16)	<5
	[A26	(20)	NP
		B*2705	16	25
]	B*2709	15	NP
]	B*5101	21	57
		Cw*0301	NP	24
51 & (52)	(P)SIPVHPI GY	A1	21 (27)	<5 ·
		A26	22	NP
	[A3	16	<5
53	IPVHPIGY	B*5101	16	NP
54	YYDAQKLLE	A1	22	<5

†No prediction

As seen in Table 10, N-terminal addition of authentic sequence to epitopes can often generate still useful, even better epitopes, for the same or different MHC restriction elements. Note for example the pairing of (G)LPSIPVHPI with HLA-A*0201, where the 10-mer can be used as a vaccine useful with several MHC types by relying on N-terminal trimming to create the epitopes for HLA-B7, -B*5101, and Cw*0401.

10 <u>HLA-A*0201 binding assay:</u>

HLA-A*0201 binding studies were preformed with PSMA₂₈₈₋₂₉₇, GLPSIPVHPI, (SEQ ID NO. 48) essentially as described in Examples 3 and 4 above. As seen in figure 8, this epitope exhibits significant binding at even lower concentrations than the positive control peptides.

Example 6

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15 Cluster Analysis (PSMA₄₅₄₋₄₈₁).

Another peptide, SSIEGNYTLRVDCTPLMYSLVHLTKEL, PSMA₄₅₄₋₄₈₁, (SEQ ID NO. 55) containing an epitope cluster from prostate specific membrane antigen, was synthesized by MPS (purity >95%) and subjected to proteasome digestion and mass spectrum analysis as described above. Prominent peaks from the mass spectra are summarized in Table 11.

Table 11. PSMA₄₅₄₋₄₈₁ Mass Peak Identification.

MS PEAK	PEPTIDE	SEQUENCE	CALCULATED
(measured)			MASS (MH ⁺)
1238.5	454-464	SSIEGNYTLRV	1239.78

1768.38±0.60	454-469	SSIEGNYTLRVDCTPL	1768.99
1899.8	454-470	SSIEGNYTLRVDCTPLM	1900.19
1097.63±0.91	463-471	RVDCTPLMY	1098.32
2062.87±0.68	454-471*	SSIEGNYTLRVDCTPLMY	2063.36
1153	472-481**	SLVHNLTKEL	1154.36
1449.93±1.79	470-481	MYSLVHNLTKEL	1448.73

Boldface sequence correspond to peptides predicted to bind to MHC, see Table 12.

Epitope Identification

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Fragments co-C-terminal with 8-10 amino acid long sequences predicted to bind HLA by the SYFPEITHI or NIH algorithms were chosen for further study. The digestion and prediction steps of the procedure can be usefully practiced in any order. Although the substrate peptide used in proteasomal digest described here was specifically designed to include predicted HLA-A2.1 binding sequences, the actual products of digestion can be checked after the fact for actual or predicted binding to other MHC molecules. Selected results are shown in Table 12.

Table 12. Predicted HLA binding by proteasomally generated fragments

SEQ ID NO	PEPTIDE	HLA	SYFPEITHI	NIE
56 & (57)	(S) IEGNYTLRV	A1	(19)	<5
		A*0201	16 (22)	<5
58	EGNYTLRV	B*5101	15	NP†
59 & (60)	(Y) TLRVDCTPL	A*0201	20 (18)	(5)
		A26	16 (18)	NP
		B7	14	40
		B8	23	<5
		B*2705	12	30
		Cw*0301	NP	(30)
61	LRVDCTPLM	B*2705	20	600
		B*2709	20	NP
62 & (63)	(L) RVDCTPLMY	A1	32 (22)	125 (13.5)
		A3	25	<5
		A26	22	NP
		B*2702	NP	(200)
		B*2705	13 (NP)	(1000)

†No prediction

As seen in Table 12, N-terminal addition of authentic sequence to epitopes can often generate still useful, even better epitopes, for the same or different MHC restriction elements. Note for example the pairing of (L)RVDCTPLMY (SEQ ID NOS 62 and (63)) with HLA-B*2702/5, where the 10-mer has substantial predicted halftimes of dissociation and the co-C-

^{*} On the basis of mass alone this peak could equally well be assigned to the peptide 455-472 however proteasomal removal of just the N-terminal amino acid is considered unlikely. If the issue were important it could be resolved by N-terminal sequencing.

^{**}On the basis of mass this fragment might also represent 455-464.

terminal 9-mer does not. Also note the case of SIEGNYTLRV (SEQ ID NO 57) a predicted HLA-A*0201 epitope which can be used as a vaccine useful with HLA-B*5101 by relying on N-terminal trimming to create the epitope.

HLA-A*0201 binding assay

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HLA-A*0201 binding studies were preformed, essentially as described in Example 3 above, with PSMA₄₆₀₋₄₆₉, TLRVDCTPL, (SEQ ID NO. 60). As seen in figure 10, this epitope was found to bind HLA-A2.1 to a similar extent as the known A2.1 binder FLPSDYFPSV (HBV₁₈₋₂₇; SEQ ID NO: 24) used as a positive control. Additionally, PSMA₄₆₁₋₄₆₉, (SEQ ID NO. 59) binds nearly as well.

10 ELISPOT analysis: PSMA₄₆₃₋₄₇₁ (SEQ ID NO. 62)

The wells of a nitrocellulose-backed microtiter plate were coated with capture antibody by incubating overnight at 4°C using 50 μ l (microliter)/well of 4 μ g/ml murine anti-human γ (gamma)-IFN monoclonal antibody in coating buffer (35 mM sodium bicarbonate, 15 mM sodium carbonate, pH 9.5). Unbound antibody was removed by washing 4 times 5 min. with PBS. Unbound sites on the membrane then were blocked by adding 200µl (microliter)/well of RPMI medium with 10% serum and incubating 1 hr. at room temperature. Antigen stimulated CD8+ T cells, in 1:3 serial dilutions, were seeded into the wells of the microtiter plate using 100µl (microliter)/well, starting at 2x105 cells/well. (Prior antigen stimulation was essentially as described in Scheibenbogen, C. et al. Int. J. Cancer 71:932-936, 1997. PSMA₄₆₂₋₄₇₁ (SEQ ID NO. 62) was added to a final concentration of $10\mu g/ml$ and IL-2 to 100 U/ml and the cells cultured at 37°C in a 5% CO₂, watersaturated atmosphere for 40 hrs. Following this incubation the plates were washed with 6 times 200 µl (microliter)/well of PBS containing 0.05% Tween-20 (PBS-Tween). Detection antibody, 50μl (microliter)/well of 2g/ml biotinylated murine anti-human γ (gamma)-IFN monoclonal antibody in PBS+10% fetal calf serum, was added and the plate incubated at room temperature for 2 hrs. Unbound detection antibody was removed by washing with 4 times 200 μl of PBS-Tween. 100µl of avidin-conjugated horseradish peroxidase (Pharmingen, San Diego, CA) was added to each well and incubated at room temperature for 1 hr. Unbound enzyme was removed by washing with 6 times 200 µl of PBS-Tween. Substrate was prepared by dissolving a 20 mg tablet of 3-amino 9-ethylcoarbasole in 2.5 ml of N, N-dimethylformamide and adding that solution to 47,5 ml of 0.05 M phosphate-citrate buffer (pH 5.0). 25 μl of 30% H_2O_2 was added to the substrate solution immediately before distributing substrate at 100 µl (microliter)/well and incubating the plate at room temperature. After color development (generally 15-30 min.), the reaction was stopped by washing the plate with water. The plate was air dried and the spots counted using a stereomicroscope.

Figure 11 shows the detection of PSMA₄₆₃₋₄₇₁ (SEQ ID NO. 62)-reactive HLA-A1⁺ CD8⁺ T cells previously generated in cultures of HLA-A1⁺ CD8⁺ T cells with autologous dendritic cells

plus the peptide. No reactivity is detected from cultures without peptide (data not shown). In this case it can be seen that the peptide reactive T cells are present in the culture at a frequency between 1 in 2.2×10^4 and 1 in 6.7×10^4 . That this is truly an HLA-A1-restricted response is demonstrated by the ability of anti-HLA-A1 monoclonal antibody to block γ (gamma) IFN production; see figure 12. Example 7

Cluster Analysis (PSMA653-687).

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Another peptide, FDKSNPIVLRMMNDQLMFLERAFIDPLGLPDRP FY PSMA₆₅₃₋₆₈₇, (SEQ ID NO. 64) containing an A2 epitope cluster from prostate specific membrane antigen, PSMA₆₆₀₋₆₈₁ (SEQ ID NO 65), was synthesized by MPS (purity >95%) and subjected to proteasome digestion and mass spectrum analysis as described above. Prominent peaks from the mass spectra are summarized in Table 13.

Table 13. PSMA₆₅₃₋₆₈₇ Mass Peak Identification.

MS PEAK	PEPTIDE	SEQUENCE	CALCULATED
(measured)	ļ		MASS (MH ⁺)
906.17±0.65	681-687**	LPDRPFY	908.05
1287.73±0.76	677-687**	DPLGLPDRPFY	1290.47
1400.3±1.79	676-687	IDPLGLPDRPFY	1403.63
1548.0±1.37	675-687	FIDPLGLPDRPFY	1550.80
1619.5±1.51	674-687**	AFIDPLGLPDRPFY	1621.88
1775.48±1.32	673-687*	RAFIDPLGLPDRPFY	1778.07
2440.2±1.3	653-672	FDKSNPIVLRMMNDQLMFLE	2442.932
1904.63±1.56	672-687*	ERAFIDPLGLPDRPFY	1907.19
2310.6±2.5	653-671	FDKSNPIVLRMMNDQLMFL	2313.82
2017.4±1.94	671-687	LERAFIDPLGLPDRPFY	2020.35
2197.43±1.78	653-670	FDKSNPIVLRMMNDQLMF	2200.66

Boldface sequence correspond to peptides predicted to bind to MHC, see Table 13.

* On the basis of mass alone this peak could equally well be assigned to a peptide beginning at 654, however proteasomal removal of just the N-terminal amino acid is considered unlikely. If the issue were important it could be resolved by N-terminal sequencing.

** On the basis of mass alone these peaks could have been assigned to internal fragments, but given the overall pattern of digestion it was considered unlikely.

Epitope Identification

Fragments co-C-terminal with 8-10 amino acid long sequences predicted to bind HLA by the SYFPEITHI or NIH algorithms were chosen for further study. The digestion and prediction steps of the procedure can be usefully practiced in any order. Although the substrate peptide used in proteasomal digest described here was specifically designed to include predicted HLA-A2.1

binding sequences, the actual products of digestion can be checked after the fact for actual or predicted binding to other MHC molecules. Selected results are shown in Table 14.

Table 14. Predicted HLA binding by proteasomally generated fragments

SEQ ID NO	PEPTIDE	HLA	SYFPEITHI	NIB
66 & (67)	(R)MMNDQLMF L	A*0201	24 (23)	1360 (722)
		A*0205	NP†	71 (42)
		A26	15	NP
		B*2705	12	50
68	RMMNDQLMF	B*2705	17	75

†No prediction

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As seen in Table 14, N-terminal addition of authentic sequence to epitopes can generate still useful, even better epitopes, for the same or different MHC restriction elements. Note for example the pairing of (R)MMNDQLMFL (SEQ ID NOS. 66 and (67)) with HLA-A*02, where the 10-mer retains substantial predicted binding potential.

HLA-A*0201 binding assay

HLA-A*0201 binding studies were preformed, essentially as described in Example 3 above, with PSMA₆₆₃₋₆₇₁, (SEQ ID NO. 66) and PSMA₆₆₂₋₆₇₁, RMMNDQLMFL (SEQ NO. 67). As seen in figures 10, 13 and 14, this epitope exhibits significant binding at even lower concentrations than the positive control peptide (FLPSDYFPSV (HBV₁₈₋₂₇); SEQ ID NO: 24). Though not run in parallel, comparison to the controls suggests that PSMA₆₆₂₋₆₇₁ (which approaches the Melan A peptide in affinity) has the superior binding activity of these two PSMA peptides.

Example 8

Vaccinating with epitope vaccines.

20 1. Vaccination with peptide vaccines:

A. <u>Intranodal delivery</u>

A formulation containing peptide in aqueous buffer with an antimicrobial agent, an antioxidant, and an immunomodulating cytokine, was injected continuously over several days into the inguinal lymph node using a miniature pumping system developed for insulin delivery (MiniMed; Northridge, CA). This infusion cycle was selected in order to mimic the kinetics of antigen presentation during a natural infection.

B. Controlled release

A peptide formulation is delivered using controlled PLGA microspheres as is known in the art, which alter the pharmacokinetics of the peptide and improve immunogenicity. This formulation is injected or taken orally.

C. Gene gun delivery

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A peptide formulation is prepared wherein the peptide is adhered to gold microparticles as is known in the art. The particles are delivered in a gene gun, being accelerated at high speed so as to penetrate the skin, carrying the particles into dermal tissues that contain pAPCs.

D. <u>Aerosol delivery</u>

A peptide formulation is inhaled as an aerosol as is known in the art, for uptake into appropriate vascular or lymphatic tissue in the lungs.

2. <u>Vaccination with nucleic acid vaccines:</u>

A nucleic acid vaccine is injected into a lymph node using a miniature pumping system, such as the MiniMed insulin pump. A nucleic acid construct formulated in an aqueous buffered solution containing an antimicrobial agent, an antioxidant, and an immunomodulating cytokine, is delivered over a several day infusion cycle in order to mimic the kinetics of antigen presentation during a natural infection.

Optionally, the nucleic acid construct is delivered using controlled release substances, such as PLGA microspheres or other biodegradable substances. These substances are injected or taken orally. Nucleic acid vaccines are given using oral delivery, priming the immune response through uptake into GALT tissues. Alternatively, the nucleic acid vaccines are delivered using a gene gun, wherein the nucleic acid vaccine is adhered to minute gold particles. Nucleic acid constructs can also be inhaled as an aerosol, for uptake into appropriate vascular or lymphatic tissue in the lungs. Example 9

Assays for the effectiveness of epitope vaccines.

25 1. <u>Tetramer analysis:</u>

Class I tetramer analysis is used to determine T cell frequency in an animal before and after administration of a housekeeping epitope. Clonal expansion of T cells in response to an epitope indicates that the epitope is presented to T cells by pAPCs. The specific T cell frequency is measured against the housekeeping epitope before and after administration of the epitope to an animal, to determine if the epitope is present on pAPCs. An increase in frequency of T cells specific to the epitope after administration indicates that the epitope was presented on pAPC.

2. <u>Proliferation assay:</u>

Approximately 24 hours after vaccination of an animal with housekeeping epitope, pAPCs are harvested from PBMCs, splenocytes, or lymph node cells, using monoclonal antibodies against specific markers present on pAPCs, fixed to magnetic beads for affinity purification. Crude blood or splenoctye preparation is enriched for pAPCs using this technique. The enriched pAPCs are

then used in a proliferation assay against a T cell clone that has been generated and is specific for the housekeeping epitope of interest. The pAPCs are coincubated with the T cell clone and the T cells are monitored for proliferation activity by measuring the incorporation of radiolabeled thymidine by T cells. Proliferation indicates that T cells specific for the housekeeping epitope are being stimulated by that epitope on the pAPCs.

Chromium release assay:

A human patient, or non-human animal genetically engineered to express human class I MHC, is immunized using a housekeeping epitope. T cells from the immunized subject are used in a standard chromium release assay using human tumor targets or targets engineered to express the same class I MHC. T cell killing of the targets indicates that stimulation of T cells in a patient would be effective at killing a tumor expressing a similar TuAA.

Example 10

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Induction of CTL response with naked DNA is efficient by Intra-lymph node immunization.

In order to quantitatively compare the CD8⁺ CTL responses induced by different routes of immunization a plasmid DNA vaccine (pEGFPL33A) containing a well-characterized immunodominant CTL epitope from the LCMV-glycoprotein (G) (gp33; amino acids 33-41) (Oehen, S., et al.. *Immunology* 99, 163-169 2000) was used, as this system allows a comprehensive assessment of antiviral CTL responses. Groups of 2 C57BL/6 mice were immunized once with titrated doses (200-0.02μg) of pEGFPL33A DNA or of control plasmid pEGFP-N3, administered i.m. (intramuscular), i.d. (intradermal), i.spl. (intrasplenic), or i.ln. (intra-lymph node). Positive control mice received 500 pfu LCMV i.v. (intravenous). Ten days after immunization spleen cells were isolated and gp33-specific CTL activity was determined after secondary *in vitro* restimulation. As shown in Fig. 15, i.m. or i.d. immunization induced weakly detectable CTL responses when high doses of pEFGPL33A DNA (200μg) were administered. In contrast, potent gp33-specific CTL responses were elicited by immunization with only 2μg pEFGPL33A DNA i.spl. and with as little as 0.2μg pEFGPL33A DNA given i.ln. (figure 15; symbols represent individual mice and one of three similar experiments is shown). Immunization with the control pEGFP-N3 DNA did not elicit any detectable gp33-specific CTL responses (data not shown).

Example 11

30 Intra-lymph node DNA immunization elicits anti-tumor immunity.

To examine whether the potent CTL responses elicited following i.ln. immunization were able to confer protection against peripheral tumors, groups of 6 C57BL/6mice were immunized three times at 6-day intervals with 10µg of pEFGPL33A DNA or control pEGFP-N3 DNA. Five days after the last immunization small pieces of solid tumors expressing the gp33 epitope (EL4-33) were transplanted s.c. into both flanks and tumor growth was measured every 3-4d. Although the

EL4-33 tumors grew well in mice that had been repetitively immunized with control pEGFP-N3 DNA (figure 16), mice which were immunized with pEFGPL33A DNA i.ln. rapidly eradicated the peripheral EL4-33 tumors (figure 16).

Example 12

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5 <u>Differences in lymph node DNA content mirrors differences in CTL response following intra-</u>
<u>lymph node and intramuscular injection.</u>

pEFGPL33A DNA was injected i.ln. or i.m. and plasmid content of the injected or draining lymph node was assessed by real time PCR after 6, 12, 24, 48 hours, and 4 and 30 days. At 6, 12, and 24 hours the plasmid DNA content of the injected lymph nodes was approximately three orders of magnitude greater than that of the draining lymph nodes following i.m. injection. No plasmid DNA was detectable in the draining lymph node at subsequent time points (Fig. 17). This is consonant with the three orders of magnitude greater dose needed using i.m. as compared to i.ln. injections to achieve a similar levels of CTL activity. CD8^{-/-} knockout mice, which do not develop a CTL response to this epitope, were also injected i.ln. showing clearance of DNA from the lymph node is not due to CD8⁺ CTL killing of cells in the lymph node. This observation also supports the conclusion that i.ln. administration will not provoke immunopathological damage to the lymph node.

Example 13

Administration of a DNA plasmid formulation of a therapeutic vaccine for melanoma to humans.

A SYNCHROTOPETM TA2M melanoma vaccine encoding the HLA-A2-restricted tyrosinase epitope SEQ ID NO. 1 and epitope cluster SEQ ID NO. 69, was formulated in 1% Benzyl alcohol, 1% ethyl alcohol, 0.5mM EDTA, citrate-phosphate, pH 7.6. Aliquots of 80, 160, and 320 μg DNA/ml were prepared for loading into MINIMED 407C infusion pumps. The catheter of a SILHOUETTE infusion set was placed into an inguinal lymph node visualized by ultrasound imaging. The assembly of pump and infusion set was originally designed for the delivery of insulin to diabetics and the usual 17mm catheter was substituted with a 31mm catheter for this application. The infusion set was kept patent for 4 days (approximately 96 hours) with an infusion rate of about 25 μl (microliter)/hour resulting in a total infused volume of approximately 2.4 ml. Thus the total administered dose per infusion was approximately 200, and 400 μg; and can be 800 μg, respectively, for the three concentrations described above. Following an infusion subjects were given a 10 day rest period before starting a subsequent infusion. Given the continued residency of plasmid DNA in the lymph node after administration (as in example 12) and the usual kinetics of CTL response following disappearance of antigen, this schedule will be sufficient to maintain the immunologic CTL response.

35 <u>Example 14</u>

Evaluating Likelihood of Epitope Cross-reactivity on Non-target Tissues.

As noted above PSA is a member of the kallikrein family of proteases, which is itself a subset of the serine protease family. While the members of this family sharing the greatest degree of sequence identity with PSA also share similar expression profiles, it remains possible that individual epitope sequences might be shared with proteins having distinctly different expression profiles. A first step in evaluating the likelihood of undesirable cross-reactivity is the identification of shared sequences. One way to accomplish this is to conduct a BLAST search of an epitope sequence against the SWISSPROT or Entrez non-redundant peptide sequence databases using the "Search for short nearly exact matches" option; hypertext transfer protocol accessible on the world wide web (http://www) at "ncbi.nlm.nih.gov/blast/index.html". Thus searching SEQ ID NO. 104, WVLTAAHCI, against SWISSPROT (limited to entries for homo sapiens) one finds four exact matches, including PSA. The other three are from kallikrein 1 (tissue kallikrein), and elastase 2A and 2B. While these nine amino acid segments are identical, the flanking sequences are quite distinct, particularly on the C-terminal side, suggesting that processing may proceed differently and that thus the same epitope may not be liberated from these other proteins. (Please note that kallikrein naming is confused. Thus, the kallikrein 1 [accession number P06870] is a different protein than the one [accession number AAD13817] mentioned in the paragraph on PSA above in the section on tumor-associated antigens).

This possibility can be tested in several ways. Synthetic peptides containing the epitope sequence embedded in the context of each of these proteins can be subjected to *in vitro* proteasomal digestion and analysis as described above. Alternatively, cells expressing these other proteins, whether by natural or recombinant expression, can be used as targets in a cytotoxicity (or similar) assay using CD8⁺ T cells that recognize the epitope, in order to determine if the epitope is processed and presented.

Examples 15-67

25 Epitopes.

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The methodologies described above, and in particular in examples 3-7, have been applied to additional synthetic peptide substrates, as summarized in figures 18-70 leading to the identification of further epitopes as set forth the in tables 15-67 below. The substrates used here were generally designed to identify products of housekeeping proteasomal processing that give rise to HLA-A*0201 binding epitopes, but additional MHC-binding reactivities can be predicted, as discussed above. Many such reactivities are disclosed, however, these listings are meant to be exemplary, not exhaustive or limiting. As also discussed above, individual components of the analyses can be used in varying combinations and orders. N-terminal pool sequencing which allows quantitation of various cleavages and can resolve ambiguities in the mass spectrum where necessary, can also be used to identify cleavage sites when digests of substrate yield fragments that do not fly well in MALDI-TOF mass spectrometry. Due to these advantages it was routinely used.

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Although it is preferred to identify epitopes on the basis of the C-terminus of an observed fragment, epitopes can also be identified on the basis of the N-terminus of an observed fragment adjacent to the epitope.

Not all of the substrates necessarily meet the formal definition of an epitope cluster as referenced in example 3. Some clusters are so large that it was more convenient to use substrates spanning only a portion of the cluster. In other cases, substrates were extended beyond clusters meeting the formal definition to include neighboring predicted epitopes or were designed around predicted epitopes with no association with any cluster. In some instances, actual binding activity dictated what substrate was made when HLA binding activity was determined for a selection of peptides with predicted affinity, before synthetic substrates were designed.

Figures 18-70 show the results of proteasomal digestion analysis as a mapping of mass spectrum peaks onto the substrate sequence. Each figure presents an individual timepoint from the digestion judged to be respresentative of the overall data, however some epitopes listed in Tables 15-67 were identified based on fragments not observed at the particular timepoints illustrated. The mapping of peaks onto the sequence was informed by N-terminal pool sequencing of the digests, as noted above. Peaks possibly corresponding to more than one fragment are represented by broken lines. Nonetheless, epitope identifications are supported by unambiguous occurrence of the associated cleavage.

Example 15: Tyrosinase 171-203

<u>Table 15</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

Epitope	Sequence	Sequence ID No.	HLA type	HLA binding SYFPEITHI	predictions† NIH
			A0201	17	93.656
171-179	NIYDLFVWM	108	A26	25	N/A
		A3	18	<5	
173-182	YDLFVWMHYY	109	A1	17	<5
			A1	16	<5
174-182	DLFVWMHYY	110	A26	30	N/A
			A3	16	27
106 104	DALL COSET	111	A0201	17	<5
186-194	DALLGGSEI	111	B5101	26	440
191-200	GSEIWRDIDF	112	A1	18	67.5
192-200	SEIWRDIDF	113	B08	16	<5 .
193-201	EIWRDIDFA	114	A26	20	N/A

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 18.

Example 16: Tyrosinase 401-427

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<u>Table 16</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

10. 24	E-itana Camanaa		TIT A 4rms	HLA binding predictions†	
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIH
407-416	LQEVYPEANA:	115	A0203	18	N/A
400 410	EXAMPLANTADI	116	A26	19	N/A
409-418	418 EVYPEANAPI	110	A3	20	<5
410-418	VYPEANAPI	117	B5101	15	6.921
411-418	YPEANAPI	118	B5101	22	N/A
411-420	YPEANAPIGH	119	A1	16	<5
416 405	ADICIDIDECM	120	A1	18	<5
416-425	APIGHNRESY	120	A26	15	N/A
			Al	16	<5
417-425	PIGHNRESY	121	A26	21	N/A
			A3	17	<5
417-426	PIGHNRESYM	122	A26	19	N/A

†Scores are given from the two binding prediction programs referenced above (see example 3) See also figure 19.

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Example 17: Tyrosinase 415-449

Table 17

Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

		Sequence	eeping Flote	HLA binding predictions†		
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIH	
			A1	18	<5	
416 425	ADICIDIDECT	120	A26	15	N/A	
416-425	APIGHNRESY	120	A3	17	<5	
			B0702	15	N/A	
			A1	16	<5	
417-425	PIGHNRESY	124	A26	21	N/A	
1			A3	17	<5	
423-430	ESYMVPFI	125	B5101	17	N/A	
423-432	ESYMVPFIPL	126	A26	18	N/A	
424-432	SYMVPFIPL	127	B0702	16	N/A	
424-433	SYMVPFIPLY	120	Al	19	<5	
424-433		128	A26	15	N/A	
			A0201	18	<5	
425-433	YMVPFIPLY	129	Al	23	5	
			A26	17	N/A	
426-434	MVPFIPLYR	130	A3	18	<5	
426-435	MVPFIPLYRN	131	A26	16	N/A	
427-434	VPFIPLYR	132	B5101	18	N/A	
430-437	IPLYRNGD	133	B08	16	<5	
430-439	IPLYRNGDFF	134	B0702	18	N/A	
431-439	PLYRNGDFF	135	A26	18	N/A	
431-439	FLIKNODFF	133	A3	24	<5	
431-440	PLYRNGDFFI	136	A0201	16	23.43	
431-440	ILIMODITI	130	A3	17	<5	
434-443	RNGDFFISSK	137	A3	20	<5	
435-443	NGDEEISSK	138	A3	15	<5	
433-443	NGDFFISSK	138	B2705	15	5	

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 20.

Example 18: Tyrosinase 457-484

Table 18

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Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Enitone	Saguanaa	Sequence	TIT A trans	HLA binding predictions†	
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIH
463-471	YIKSYLEQA	139	A0201	18	<5
403-471	I IKS I LEQA	139	A26	17	N/A
466-474	SYLEQASRI	140	B5101	16	<5
469-478	EQASRIWSWL	141	A26	17	N/A
470-478	QASRIWSWL	142	B5101	16	55
471-478	ASRIWSWL	143	B08	16	<5
471-479	ASRIWSWLL	144	B08	16	<5
			A0201	19	13.04
473-481	RIWSWLLGA	145	A26	16	N/A
			A3	15	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 21.

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Example 19: CEA 92-118

Table 19

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Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

	Epitopes Revealed	Sequenc	_		HLA binding predictions†		
Epitope	Sequence	e ID No.	HLA type		NIH		
	•		B0702	18	8		
92-100	GPAYSGREI	146	B08	15	<5		
			B5101	22	484		
92-101	GPAYSGREII	147	B0702	18	12		
93-100	PAYSGREI	148	B5101	22	N.A.		
93-101	PAYSGREII	149	B5101	24	48.4		
93-102	PAYSGREIIY	150	A1	19	<5		
94-102	AYSGREIIY	151	A1	21	<5		
97-105	GREIIYPNA	152	B2705	17	200		
37-103	GKEHYPNA	132	B2709	16			
98-107	REIIYPNASL	153	A0201	16	<5		
			A0201	21	<5		
	*	l i	A26	28	N.A.		
99-107	EIIYPNASL	154	A3	16	<5		
99-107	EHITMASL		B0702	15	6		
			B08	18	<5		
			B2705	16	<5		
			A0201	16	<5		
99-108	EIIYPNASLL	155	A26	27	N.A.		
			A3	17	<5		
100-107	IIYPNASL	156	B08	15	<5		
		!	A0201	23	15.979		
			A26	21	N.A.		
			A24	N.A.	<5		
100-108	IIYPNASLL	157	A3	23	<5		
100-100	HIIINAGEL	137	B08	15	<5		
			B1510	15	N.A.		
			B2705	16	50		
			B2709	15			
100-109	IIYPNASLLI	158	A0201	22	7.804		
	·		A3	20	<5		
102-109	YPNASLLI	159	B5101	23	N.A.		
107-116	LLIQNIIQND	160	A0201	18	<5		
107-110	ENGIATION D	100	A26	17	N.A.		

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 22.

Example 20: CEA 131-159

Table 20

Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

		Sequenc		HLA binding predictions†	
Epitope	Sequence	e ID No.	HLA type	SYFPEITHI	NIH
122 141 PRACTICATIVA	161	A1	19	<5	
132-141	132-141 EEATGQFRVY	161	A26	21	N.A.
		162	A1	22	<5
133-141	EATGQFRVY		A26	23	N.A.
			B5101	16	<5
141 140	VADET DIADAL	163	B0702	20	<5
141-149	YPELPKPSI		B5101	22	572
142-149	PELPKPSI	164	B08	16	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 23.

Example 21: CEA 225-251

Table 21

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Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

		Sequenc	,	HLA binding	g predictions†
Epitope	Sequence	e ID No.	HLA type	SYFPEITHI	NIH
			A0201	15	<5
225-233	RSDSVILNV	165	Al	22	<5
			B2709	15	N.A.
225-234	RSDSVILNVL	166	A0201	15	<5
226-234	SDSVILNVL	167	A0201	17	<5
226-235	SDSVILNVLY	168	A1	20	<5
227 225	DOMESTICAL ST	1.00	A1	22	<5
227-235	DSVILNVLY	169	A26	18	N.A.
022 242	ATT MODE A DOT	170	A0201	25	56.754
233-242	VLYGPDAPTI	170	A3	23	<5
224 242	LYGPDAPTI	171	A0201	15	<5
234-242			B5101	15	5.72
235-242	YGPDAPTI	172	B5101	22	N.A.
026.045	CDD A DELIGIDA	172	A0201	15	<5
236-245	GPDAPTISPL	173	B0702	23	24
			A0201	15	<5
237-245	PDAPTISPL	174	A26	16	N.A.
			B2705	15	<5
238-245	DAPTISPL	175	B5101	25	N.A.
239-247	APTISPLNT	176	B0702	20	6
240 240	DTIGDI NTGA	122	A1	22	<5
240-249	PTISPLNTSY	177	A26	24	N.A.
			A1	20	5
241-249	TISPLNTSY	178	A26	24	N.A.
			A3	20	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 24.

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Example 22: CEA 239-270

<u>Table 22</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

	tope Sequence e HLA	Sequenc		HLA binding	predictions†
Epitope		HLA type	SYFPEITHI	NIH	
240 240	DTIGDI NTGV	179	A1	22	<5
240-249	PTISPLNTSY	179	A26	24	N.A.
			A1	20	5
241-249	TISPLNTSY	180	A26	24	N.A.
			A3	20	<5
246-255	NTSYRSGENL	181	A26	19	N.A.
247-255	TSYRSGENL	182	B2705	15	50
248-255	SYRSGENL	183	B08	18	<5
248-257	SYRSGENLNL	184	B0702	14	<5
			A.0201	15	<5
240 257	ADDOCEMENT.	185	B0702	16	<5
249-257	YRSGENLNL	165	B2705	27	2000
			B2709	22	N.A.
251-259	SGENLNLSC	186	A1	19	<5
253-262	ENLNLSCHAA	187	A0203	19	<5
254-262	NLNLSCHAA	188	A0201	17	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 25.

Example 23: CEA 259-286

10 Table 23

5

Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

		Sequenc			HLA binding predictions†	
Epitope	Sequence	e ID No.	HLA type	SYFPEITHI	NIH	
260-269	HAASNPPAQY	189	A1	15	<5	
261 260	A A CATOD A OV	190	A1	17	<5	
261-269	AASNPPAQY	190	A3	17	<5	
264-273	NPPAQYSWFV	191	B0702	18	∜	
265 272	DD V ONGMEN	192	B0702	18	<5	
265-273	PPAQYSWFV		B5101	19	20	
266-273	PAQYSWFV	193	B5101	18	N.A.	
272-280	EVNICTEOOG	104	A26	18	N.A.	
212-280	FVNGTFQQS	194	A3	15	<5	

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 26.

Example 24: CEA 309-336

Table 24

Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

		Sequenc		HLA binding	predictions†
Epitope	Epitope Sequence e H	HLA type	SYFPEITHI	NIH	
			A1	22	<5
310-319	RTTVTTITVY	195	A26	24	N.A.
			A3	15	<5
			A1	22	<5
311-319	TTVTTITVY	196	A26	24	N.A.
			B2705	15	5
			A0201	17	<5
319-327	YAEPPKPFI	197	A1	17	18
			B5101	22	286
319-328	YAEPPKPFIT	198	A1	16	45
320-327	AEPPKPFI	199	B08	16	<5
321-328	EPPKPFIT	200	B5101	16	N.A.
321-329	EPPKPFITS	201	B0702	16	<5
321-329	ELLECTI13	201	B5101	16	12.1
322-329	PPKPFITS	202	B08	16	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 27.

Example 25: CEA 381-408

Table 25

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10 Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

		Sequenc		HLA binding predictions†		
Epitope	Sequence	D No.	HLA type	SYFPEITHI	NIH	
			A1	18	<5	
382-391	SVTRNDVGPY	203	A26	24	N.A.	
			A3	21	<5	
383-391	VTRNDVGPY	204	A1	23	<5	
303-371			A26	24	N.A.	
389-397	GPYECGIQN	205	B5101	17	11	
391-399	YECGIQNEL	206	A0201	17	<5	
391-399	TECOIQNEL	200	B2705	17	30	
394-402	GIQNELSVD	207	A26	15	N.A.	
374-402	GIQIAELSAD		A3	16	<5	

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 28.

Example 26: CEA 403-429

Table 26

5 Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

		Sequenc		HLA binding	g predictions†
Epitope	Sequence	e ID No.	HLA type	SYFPEITHI	NIH
403-411	HSDPVILNV	208	A0201	17	<5
403-411	TIODI VILIV	200	A1	26	37.5
			A0201	17	<5
		1	A1	19	7.5
403-412	HSDPVILNVL	209	A26	15	N.A.
		ŀ	A24	N.A.	8.064
			B4402	17	N.A.
404-412	SDPVILNVL	210	A0201	17	<5
404-412	SDIVILIAND	210	B4402	16	N.A.
404-413	SDPVILNVLY	211	A1	20	<5
405-412	DPVILNVL	212	B08	16	<5
403-412	DI VILIVIL	212	B5101	24	N.A.
			A1	18	<5
405-413	DPVILNVLY	213	A26	18	N.A.
			B5101	16	7.26
408-417	ILNVLYGPDD	214	A3	15	<5
411-420	VLYGPDDPTI	215	A0201	25	56.754
711 720	VETGI DDI II	213	A3	20	<5
412-420	LYGPDDPTI	216	A0201	15	<5
			A24	N.A.	60
413-420	YGPDDPTI	217	B5101	22	N.A.
417-425	DPTISPSYT	218	B0702	16	<5
418-427	PTISPSYTYY	219	A1	.21	<5
		217	A26	27	N.A.
419-427	TISPSYTYY	220	A1	19	5
			A26	27	N.A.

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 29.

Example 27: CEA 416-448

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<u>Table 27</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

Epitope	Sequence	Sequence	HLA type	HLA binding	g predictions†
Epitope	Sequence .	ID No.	IIIA type	SYFPEITHI	NIH
418-427	418-427 PTISPSYTYY	221	Al	21	<5
110-127		221	A26	27	N.A.
			Al	19	5
419-427	TISPSYTYY	222	A26	27	N.A.
			A3	18	<5
419-428	TISPSYTYYR	223	A3	15	5.4
			A0201	18	<5
424-433	YTYYRPGVNL	224	A24	N.A.	<5
			A26	20	N.A.
	İ		A0201	14	<5
425-433	TYYRPGVNL	225	A24	N.A.	200
425-455			B0702	16	<5
L			B2705	16	5
426-433	YYRPGVNL	226	B08	16	<5
426-435	YYRPGVNLSL	227	A0201	17	<5
120 133	TIM OVIVESE		B0702	15	<5
			A0201	17	<5
427-435	YRPGVNLSL	228	B2705	26	2000
			B2709	21	N.A.
428-435	RPGVNLSL	229	B08	17	<5
		229	B5101	17	N.A.
428-437	RPGVNLSLSC	230	B0702	14	<5
430-438	GVNLSLSCH	231	A26	16	N.A.
			B2705	15	<5
431-440	VNLSLSCHAA	232	A0203	19	N.A.
432-440	NLSLSCHAA	233	A0201	16	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 30.

Example 28: CEA 437-464

Table 28

Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Epitope	Sequence	Sequence	DT A 4-	HI A binding	g predictions†
Phrohe	Sequence	ID No.	HLA type	SYFPEITHI	NIH
438-447	HAASNPPAQY	234	A1	15	<5
439-447	AASNPPAQY	235	A1	17	<5
433-447	AASNITAQI	233	A3	17	<5
442-451	NPPAQYSWLI	236	B0702	17	8
443-451	PPAQYSWLI	237	B0702	17	<5
773-731	TTAQISWLI	237	B5101	21	40
444-451	PAQYSWLI	238	B5101	20	N.A.
			A0201	17	<5
449-458	WLIDGNIQQH	239	A26	17	N.A.
			A3	21	<5
			A0201	16	<5
450-458	LIDGNIQQH	240	A26	19	N.A.
			A3	17	<5
450-459	LIDGNIQQHT	241	A0201	16	<5
130-139	LIDOMQQIII	241	A26	15	N.A.

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 31.

Example 29: CEA 581-607

Table 29

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Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Epitope	Sequence	Sequence		HLA binding	g predictions†
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIH
j			A0201	16	<5
581-590	RSDPVTLDVL	242	A1	19	7.5
301-370	RODI VILDVL	242	A26	15	N.A.
			A24	N.A.	9.6
582-590	SDPVTLDVL	243	A0201	16	<5
582-591	SDPVTLDVLY	244	A1	19	<5
583-590	DPVTLDVL	245	B08	16	<5
303-370	DIVILOVL	243	B5101	25	N.A.
			A1	17	<5
583-591	DPVTLDVLY	246	A26	18	N.A.
			B5101	16	6
588-597	DVLYGPDTPI	247	A26	16	N.A.
			A0201	25	56.754
589-597	VLYGPDTPI	248	A3	17	6.75
			B5101	17	11.44
			A1	15	<5
596-605	PIJSPPDSSY	249	A26	25	N.A.
J			A3	22	<5
		_	A1	20	5
597-605	HSPPDSSY	250	A26	24	N.A.
L			A3	24	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 32.

Example 30: CEA 595-622

<u>Table 30</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

Eniton.	Sagranas	Sequence	TOT A tyme	HLA binding	predictions†
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIH
			A0201	22	27.464
597-606	IISPPDSSYL	251	A26	21	N.A.
397-000	HSPPDSS IL	231	A3	16	<5
			B0702	14	<5
599-606	CODOCCIA	252	B08	18	<5_
399-000	SPPDSSYL	232	B5101	17	N.A.
600-608	PPDSSYLSG	253	A1	16	<5
600-609	PPDSSYLSGA	254	B0702	17	<5
602-611	DSSYLSGANL	255	A26	16	N.A.
603-611	SSYLSGANL	256	A0201	15	<5
003-011		250	B2705	17	50
604-613	SYLSGANLNL	257	A0201	15	<5
004-013	SILSGANULIL	231	A24	N.A.	300
		_	A0201	25	98.267
			A26	19	N.A.
605-613	YLSGANLNL	258	A3	15	<5
003-013	ILOUANLINL	230	B0702	16	<5
			B08	17	<5
			B2705	16	30
610-618	NLNLSCHSA	259	A0201	18	<5

†Scores are given from the two binding prediction programs referenced above (see example 3) See also figure 33.

Example 31: CEA 615-641

Table 31

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10 Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Enitona	G	Sequence	TTT A 4-m-s	HLA binding	HLA binding predictions†	
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIH	
620-629	NPSPQYSWRI	260	B0702	19	8	
622-629	SPQYSWRI	261	B08	15	<5	
022-029	SPQISWKL	201	B5101	20	N.A.	
627-635	WRINGIPQQ	262	B2705	19	20	
628-636	RINGIPQQH	263	A3	22	<5	
020-030	KUNGIPQQH	203	B2705	16	<5	
628-637	RINGIPQQHT	264	A0201	15	<5	
631-639	GIPQQHTQV	265	A0201	19	9.563	
632-639	IPQQHTQV	266	B5101	20	N.A.	

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 34.

Example 32: CEA 643-677

<u>Table 32</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

T7	C	Sequence	777 4 4	HLA binding	predictions†
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIH
			A1	20	5
644-653	KITPNNNGTY	267	A26	22	N.A.
	: H=:		A3	25	<5
			A1	22	<5
645-653	ITPNNNGTY	268	A26	21	N.A.
			A3	14	<5
647-656	PNNNGTYACF	269	A26	15	N.A.
648-656	NNNGTYACF	270	A26_	17	N.A.
650-657	NGTYACFV	271	B5101	15	N.A.
661-670	ATGRNNSIVK	272	A3	20	<5
662-670	TGRNNSIVK	273	A3	18	<5
664-672	RNNSIVKSI	274	B2709	15	N.A.
666-674	NSIVKSITV	275	A0201	16	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 35.

Example 33: GAGE-1 6-32

10 <u>Table 33</u>

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Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

173 24	9	Sequence	HLA	HLA binding	g predictions†
Epitope	Sequence	ID No.	type	SYFPEITHI	NIH
			Al	23	<5
7-16	STYRPRPRRY	276	A26	21	N/A
			A3	15	<5
0.16	TYTERENTAL	077	A1	19	<5
8-16	TYRPRPRRY	277	A3	15	<5
	RPRPRRYVE	RYVE 278	A3	17	<5
10-18			B0702	16	N/A
			B08	20	<5
16-23	YVEPPEMI	279	B5101	15	N/A
22.21	AGCDA GODEOE	200	A26	23	N/A
22-31	MIGPMRPEQF	280	A3	19	<5
23-31	IGPMRPEQF	281	B08	15	<5
24-31	GPMRPEQF	282	B5101	16	N/A

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 36.

Example 34: GAGE-1 105-131

Table 34

5 Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

TF-:4	S	Sequence	TIT A 4	HLA binding	predictions†	
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NUH	
105-114	KTPEEEMRSH	283	A26	18	N/A	
106-115	TPEEEMRSHY	284	A1	26	11.25	
107-115	PEEEMRSHY	285	A1	26	<5	
110-119	EMRSHYVAQT	286	A0201	15	<5	
113-121	SHYVAQTGI	287	B5101	15	<5	
			A0201	23	108.769	
115-124	YVAQTGILWL	288	A26	24	N/A	
	•		A3	15	<5	
	VAQTGILWL			A0201	22	6.381
116-124		289	B08	16	<5	
110-124		289	B2705	16	10	
			B5101	20	78.65	
116-125	VAQTGILWLL	290	A0201	19	8.701	
117-125	A OTOT TITLE	291	A0201	17	37.362	
11/-123	AQTGILWLL	291	B2705	16	200	
118-126	QTGILWLLM	292	A26	19	N/A	
118-127	QTGILWLLMN	293	A26	15	N/A	
120-129	GILWLLMNNC	294	A26	15	N/A	
121-129	ILWLLMNNC	295	A0201	15	161.227	

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 37.

Example 35: GAGE-1 112-137

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<u>Table 35</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

T-:4 C		Sequence	HLA	HLA bindin	g predictions†
Epitope	Sequence	ID No.	type	SYFPEITHI	NIH
124-131	LLMNNCFL	296	B08	16	<5
			A0201	22	1999.734
123-131	WLLMNNCFL	297	A26	16	N/A
			B08	17	<5
122-130	LWLLMNNCF	298	B2705	15	<5
121-130	ILWLLMNNCF	299	A26	18	N/A.
121-130	ILW LLIMINICE	299	A3	17	10
121-129	ILWLLMNNC	295	A0201	15	161.227
120-129	GILWLLMNNC	294	A26	15	N/A
118-127	QTGILWLLMN	293	A26	15	N/A
118-126	QTGILWLLM	292	A26	19	N/A
			A0201	17	37.362
117-125	AQTGILWLL	291	B2705	16	200
			B4402	17	N/A
116-125	VAQTGILWLL	290	A0201	19	8.701
			A0201	22	6.381
			B08	16	<5
116-124	VAQTGILWL	289	B2705	16	10
			B4402	15	N/A
			B5101	20	78.65
_			A0201	23	108.769
115-124	YVAQTGILWL	288	A26	24	N/A
			A3	15	<5
113-121	SHYVAQTGI	287	B5101	15	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 38.

Example 36 MAGE-1 51-77

Table 36

Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Epitope	Converse	Sequence ID	HLA type	HLA binding predictions†		
	Sequence	No.	HLA type	SYFPEITHI	NIH	
62-70 SAFPTTINF		A26	15	N/A		
	SAFPTTINF	309	B4402	18	N/A	
			B2705	17	25	
61-70	ASAFPTTINF	310	B4402	15	N/A	
60.60	CACAEDTEL	311	A0201	16	<5	
60-68	GASAFPTTI	311	B5101	25	220	
57-66	SPQGASAFPT	312	B0702	19	N/A	

†Scores are given from the two binding prediction programs referenced above. See also figure 39.

Example 37: Mage-1 126-153

Table 37

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Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

		Sequence		HLA binding	·
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIH
144-151	FGKASESL	313	B08	21	<5
143-151	IFGKASESL	314	A26	16	N/A
143-131	II'UKASESE	314	B2705	15	<5
			A0201	20	<5
142-151	EIFGKASESL	315	A26	29	N/A
			B4402	15	N/A
142-149	EIFGKASE	316	B08	16	<5
133-140	IKNYKHCF	317	B08	18	<5
132-140	VIKNYKHCF	318	A26	21	N/A
132-140	VIKIVI KHCF	310	B08	21	<5
		319	A26	23	N/A
131-140	SVIKNYKHCF		A3	18	<5
			B4402	15	N/A
132-139	VIKNYKHC	320	B08	15	<5
131-139	SVIKNYKHC	321	A26	18	N/A
			A1	28	45
128-136	MLESVIKNY	322	A26	24	N/A
120-130	MILESVIKIVI	322	A3	. 17	<5
			B4402	15	N/A
			A1	15	<5
127-136	EMLESVIKNY	323	A26	23	N/A
			B4402	18	N/A
			A3	18	<5
126-134	AEMLESVIK	324	B2705	15	30
			B4402	16	N/A

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 40.

Example 38: MAGE-2 272-299

Table 38

5 Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

T		Sequence	TT 4 4	HLA binding	predictions†
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIH
274-283	GPRALIETSY	325	A1	15	<5
075 002	DD AT ITYTOX	200	A1	15	<5
275-283	PRALIETSY	326	B2705	23	100
076 004	DAT TOTOLOGI	207	A0201	18	19.658
276-284	RALIETSYV	327	B5101	20	55
			A0201	30	427.745
277-286	ALIETSYVKV	328	A26	18	N/A
	1		A3	21	<5
			A0201	23	<5
278-286	LIETSYVKV	329	A26	17	N/A
	1		B5101	15	<5
070 007	r reseason over	ETSYVKVL 330	A0201	22	<5
278-287	LIEISYVKVL		A26	22	N/A
			A0201	15	<5
279-287	IETSYVKVL	331	B1510	15	N/A
			B5101	15	<5
280-289	ETSYVKVLH H	332	A26	21	N/A
282-291	SYVKVLHHT L	333	A0201	15	<5
			A0201	19	<5
283-291	YVKVLHHTL	334	A26	20	N/A
283-291	IVEATURIT	334	A3	15	<5
	<u> </u>		B08	21	<5
			A0201	20	11.822
285-293	KVLHHTLKI	335	A3	18	<5
			B5101	15	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). | See also figure 41.

Example 39 MAGE-2 287-314

<u>Table 39</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

		Sequence	TTT A 4	HLA binding	predictions†
Epitope	Sequence	HLA typ		SYFPEITHI	NIH
		226	A3	19	<5
303-311	PLHERALRE	336	B08	16	<5
	**************************************	227	B08	16	<5
302-309	2-309 PPLHERAL	337	B5101	18	N/A
			B0702	21	N/A
201 200	YPPLHERAL	PPLHERAL 338	B08	18	<5
301-309			B4402	15	N/A
			B5101	20	143
222 222	GI MAY TITTO A T	220	A0201	15	<5
300-309	SYPPLHERAL	339	B4402	18	N/A
299-307	ISYPPLHER	340	B2705	17	25
298-307	HISYPPLHER	341	A26	15	N/A
292-299	KIGGEPHI	342	B5101	15	N/A
291-299	LKIGGEPHI	343	A0201	17	<5
290-299	TLKIGGEPHI	344	A0201	18	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 42.

Example 40 Mage-3 287-314

Table 40

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Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Epitope	Sequence	Sequence	HLA type	HLA bi	_
• •	-	ID No.		SYFPEITHI	NIH
303-311	PLHEWVLRE	345	A26	15	N/A
200 200	309 PPLHEWVL	246	B08	16	<5
302-309		346	B5101	19	N/A
			B0702	21	N/A
301-309	YPPLHEWVL	347	B08	17	<5
			B5101	22	130
301-308	YPPLHEWV	348	B5101	22	N/A
300-308	SYPPLHEWV	349	A0201	15	<5
299-308	ISYPPLHEWV	350	A0201	15	6.656
298-307	HISYPPLHEW	351	A26	15	N/A
293-301	ISGGPHISY	352	A1	25	<5
			A1	20	<5
292-301	KISGGPHISY	353	A26	23	N/A
			A3	21	5.4

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 43.

Example 41: Melan-A 44-71

Table 41

5 Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

777-14		Sequence	TT 4 4		oinding ctions†
Epitope	Sequence	ID No.	HLA type	SYFPEITH I	NIH
45-54	CWYCRRRNG Y	354	A1	16	<5
46-54	WYCRRRNGY	355	A1	16	<5
47-55	YCRRRNGYR	356	B08	15	<5
	RRRNGYRAL	-	B08	17	<5
49-57		357	B2705	26	1800
			B2709	24	N/A
51-60	RNGYRALMD K	358	A3	15	<5
52-60	NGYRALMDK	359	A3	18	<5
55-63	RALMDKSLH	360	B2705	16	<5
56-63	ALMDKSLH	361	B08	16	<5
55-64	RALMDKSLH V	362	A0201	17	<5
			A0201	26	1055.104
56-64	ALMDKSLHV	363	A3	18	<5
			B08	16	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 44.

Example 42: PRAME 274-301

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<u>Table 42</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

177. *4	Sequence	Sequence		l	oinding ctions†
Epitope		ID No.	HLA type	SYFPEITH	
				I	NIH
			A1	21	5
275-284	YISPEKEEQY	364	A26	23	N/A
213-204	IISPEKEEQI	30 4	A3	20	<5
			B4402	15	N/A
276-284	ISPEKEEQY	365	A1	19	<5
270-204	ISPEKEEQI	303	A26	15	N/A
277-285	SPEKEEQYI	266	B0702	17	N/A
211-203	SPEKEEQII	366	B5101	21	484
278-285	PEKEEQYI	367	B08	18	<5
279-288	ENEEOMYOE	260	A26	24	N/A
2/9-200	-288 EKEEQYIAQF	368	B4402	16	N/A
	KEEQYIAQF	369	A26	17	N/A
280-288			B2705	19	45
			B4402	25	N/A
283-292	OMACETECE	270	A3	17	<5
265-292	QYIAQFTSQF	370	B4402	15	N/A
		TIAQFTSQF 371	A0201	15	<5
284-292	YIAQFTSQF		A26	24	N/A
			A3	19	<5
284-293	YIAQFTSQFL	372	A0201	22	74.314
204-233	TIAQFISQFL	312	A26	21	N/A
			A0201	15	<5
285-293	IAQFTSQFL	373	B08	15	<5
			B5101	19	78.65
			A0201	16	15.226
286-295	AQFTSQFLSL	374	A26	15	N/A
200-233	TOT. ISOLESE	3/4	B0702	15	N/A
			A4402	18	N/A
287-295	QFTSQFLSL	375	A26	21	N/A
			A0201	17	18.432
290-298	SOET ST OCT	376	A26	16	N/A
27U-270	SQFLSLQCL	3/0	B2705	16	1000
			B4402	15	N/A

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 45.

Example 43: PRAME 434-463

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<u>Table 43</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

Epitope	Sequence	Sequence ID No.	HLA type	HLA binding	predictions† NIH
			A0201	20	<5
420 440	VLYPVPLESY	277	A1	21	5
439-448	VLYPVPLEST	377	A26	25	N/A
			A3	25	67.5
440-448	LYPVPLESY	378	A1	16	<5
446-455	ESYEDIHGTL	379	A26	16	N/A
448-457	YEDIHGTLHL	380	Al	18	<5
449-457	EDIHGTLHL	381	B2705	15	<5
451-460	IHGTLHLERL	382	A0201	16	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 46.

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Example 44: PRAME 452-480

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<u>Table 44</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

	Sequence	Sequence		7.	oinding ctions†
Epitope		ID No.	HLA type	SYFPETTH I	NIH
454 460	CT TTT TDT 43/7	202	A0201	26	270.234
454-463	TLHLERLAYL	383	A26	21	N/A
			A0201	22	<5
155 160	T 777 DD 7 4377	204	B08	20	<5
455-463	LHLERLAYL	384	B1510	21	N/A
			B2705	15	<5
456-463	HLERLAYL	385	B08	17	<5
	HLERLAYLH	206	A3	16	<5
456-465	A	386	A1	17	<5
458-467	ERLAYLHARL	387	A26	16	N/A
	RLAYLHARL		A0201	24	21.362
150 155		388	B08	17	<5
459-467			B2705	18	90
			B2709	15	N/A
459-468	RLAYLHARL R	389	A3	22	<5
160 167	* ******	200	B08	15	<5
460-467	LAYLHARL	390	B5101	20	N/A
460-468	LAYLHARLR	391	B5101	18	<5
461 470	ATTIADIDE	200	A0201	20	<5
461-470	AYLHARLREL	392	B4402	16	N/A
460 470	X/I XX A DX DESI	202	A0201	28	45.203
462-470	YLHARLREL	393	B08	25	8
460 471	VI II A DI DELLI	204	A0201	22	48.151
462-471	YLHARLRELL	394	A26	16	N/A
462 471	THADIDET	206	A0201	15	<5
463-471	LHARLRELL	395	B1510	22	N/A
164 471	TIADIDETT	206	B08	30	320
464-471	HARLRELL	396	B5101	17	N/A
464-472	HARLRELLC	397	B08	20	16
469-478	ELLCELGRPS	398	A3	15	<5
470-478	LLCELGRPS	399	A0201	15	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 47.

Example 45: PSA 143-169

Table 45

Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

77		Sequence	TTT A 4	HLA bindin SYFPEITHI	g predictions†
Epitope	Sequence	ID No.	ньа туре	SYFPEITHI	HIN
144-153	QEPALGTTCY	400	A1	15	<5
145 150	DD 41 CONCOV	401	A1	17	<5
145-153	EPALGTTCY	401	A26	17	N/A

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 48.

Example 46: PSA 156-1883

Table 46

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Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

T7	6	Sequence	HLA type	HLA binding predictions†		
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIH	
162-171	PEEFLTPKKL	402	B4402	24	N.A.	
162 171	EEFLTPKKL	403	A26	17	N.A.	
163-171	EEFLIPKKL	403	B4402	29	N.A.	
165-173	EL TREET OC	404	A3	20	<5	
103-173	FLTPKKLQC	404	B08	17	<5	
165 174	FLTPKKLOCV	405	A0201	26	735.86	
165-174	FLIPKKLQCV	403	A26	15	N.A.	
166-174	LTPKKLQCV	406	A0201	21	<5	
100-174		LITERALQCV	400	A26	18	N.A.
167-174	TPKKLQCV	407	B08	16	<5	
107-174	IPAKLQCV	407	B5101	22	N.A.	
167-175	TPKKLQCVD	408	B5101	15	<5	
170 170	KI OCADI IIXA	409	A0201	24	34.433	
1/0-1/9	170-179 KLQCVDLHVI 4		A3	17	<5	
171-179	LOCVDLHVI	410	A0201	15	<5	
1/1-1/9	INCADPRAI	410	B5101	16	6.292	

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 49.

Example 47: PSCA 67-94

<u>Table 47</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

Epitope	Sequence	Sequence	HLA type	HLA binding	predictions†
	Sequence	ID No.	ньа туре	SYFPEITHI	NIH
73-81	DSQDYYVGK	411	A3	15	<5
74-82	SQDYYVGKK	412	Al	16	<5
74-83	SQDYYVGKK N	413	Al	15	<5
76-84	DYYVGKKNI	414	B5101	19	23.426
77-84	YYVGKKNI	415	B08	16	<5
78-86	YVGKKNITC	416	A3	15	<5
78-87	YVGKKNITCC		A26	15	N/A

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 50.

Example 48: PSMA 378-405

Table 48

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10 Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Enitone	Sequence	Sequence	HLA type	HLA binding predictions†	
Epitope		ID No.	HLA type	SYFPEITHI	NIH
381-390	WVFGGIDPQS	418	A26	16	N/A
361-390	WALGOIDLOS	410	A3	15	<5
			A0201	24	<5
1			A0203	17	N/A
385-394	GIDPQSGAAV	419	A1	15	10
			A26	15	N/A
		<u> </u>	A3	18	<5
386-394	IDPQSGAAV	420	A0201	15	<5
387-394	DPQSGAAV	421	B5101	22	N/A
387-395	DPQSGAAVV	422	B0702	18	N/A
307-393			B5101	26	440
387-396	DPQSGAAVVH	423	A3	15	<5
388-396	PQSGAAVVH	424	A3	17	<5
389-398	QSGAAVVHEI	425	A0201	15	<5
390-398	SGAAVVHEI	426	A0201	19	<5
390-396	3GAAV VIIEI	426	B5101	21	88
391-398	GAAVVHEI	427	B5101	23	N/A
391-399	CA ANDUERI	420	A0201	17	<5
391-399	GAAVVHEIV	428	B5101	20	133.1
392-399	AAVVHEIV	429	B5101	19	N/A

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 51.

Example 49: PSMA 597-623

Table 49

Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Enitone	Sequence	Sequence	TIT A true	HLA binding predictions†		
Epitope		ID No.	HLA type	SYFPEITHI	NIH	
597-605	CRDYAVVLR	430	B2705	22	N/A	
	DDM YMM DM		A1	17	<5	
598-607	RDYAVVLRK Y	431	A26	15	N/A	
	1		A3	16	<5	
500-607	DYAVVLRKY	432	A1	19	<5	
399-007	DIAVVEREI	732	A26	22	N/A	
600-607	YAVVLRKY	433	B5101	17	N/A	
602-611	VVLRKYADKI	434	A0201	17	<5	
002-011	VVLKKTADKI	454	A3	18	<5	
	VLRKYADKI	435	A0201	22	<5	
603-611			A3	16	<5	
003-011			B08	19	<5	
			B5101	16	5.72	
İ			A 1	17	<5	
603-612	VLRKYADKIY	436	A26	19	N/A	
			A3	19	<5	
604-611	LRKYADKI	437	B08	17	<5	
604-612	LRKYADKIY	438	A1	15	<5	
004-012	Didtimbidi	450	B2705	19	N/A	
605-614	RKYADKIYSI	439	A0201	16	<5	
606-614	KYADKIYSI	440	A0201	20	<5	
		——————————————————————————————————————	B08	17	<5	
607-614	YADKIYSI	441	B5101	. 27	N/A	

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 52.

Example 50: PSMA 615-642

<u>Table 50</u>

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Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Epitope	Sequence	Sequence	HLA type	HLA binding	predictions†
	Sequence	ID No.	ma type	SYFPEITHI	NIH
616 625	MKHPQEMKT	442	A1	19	<5
010-023	Y		A26	16	N/A
617 625	KHPQEMKTY	443	A1	15	<5
017-023	KHIYEMKI I		A26	16	N/A
618 627	HPQEMKTYSV	444	A0201	15	<5
016-027			B0702	17	N/A

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 53.

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Example 51: SCP-1 57-86

<u>Table 51</u>

Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Epitope	Sequence	Sequence	HLA type	HLA binding predictions†	
Epitope	Sequence	ID No.	ших суре	SYFPEITHI	NIH
62-71	IDSDPALQKV	445	A0201	19	<5
			A0201	17	<5
63-71	DSDPALQKV	446	A1	20	7.5
03-71	DSDFALQKV	440	A26	15	N/A
			B5101	15	5.324
67-76	ALQKVNFLPV	447	A0201	23	132.149
07-70		447	A3	16	<5
70-78	KVNFLPVLE	448	A3	18	<5
71-80	VNFLPVLEQV	449	A0201	16	<5
72-80	NFLPVLEQV	450	A0201	18	<5
75-84	PVLEQVGNSD	451	A3	18	<5
76-84	VLEQVGNSD	452	Al	15	<5
/0-0-4	Arred Agusp	432	A3	16	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 54.

Example 52: SCP-1 201-227

Table 52

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Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Epitope	Sequence	Sequence	HLA type	HLA binding predictions†	
	Бефиенее	ID No.	ILA type	SYFPEITHI	NIH
202-210	YEREETRQV	453	A0201	16	<5
			Al	19	<5
202-211	YEREETRQVY	454	A3	15	<5
			A4402	22	N/A
			A1	27	<5
203-211	EREETRQVY	455	A26	19	N/A
			B2705	20	N/A
203-212	EREETRQVYM	456	A26	17	N/A
204-212	REETRQVYM	457	B2705	15	N/A
211-220	YMDLNSNIEK	458	A1	17	25
213-221	DLNSNIEKM	459	A0201	20	<5
213-221			A26	28	N/A
216-226	SNIEKMITAF	460	A26	19	N/A
210-220	SNIEKMITAF	460	B4402	19	N/A
217-225	NIEKMITAF		A26	26	N/A
		461	B2705	17	N/A
			B4402	16	N/A
218-225	IEKMITAF	462	B08	17	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 55.

Example 53: SCP-1 395-424

<u>Table 53</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

Epitope	Sequence	Sequence	HLA type	HLA binding predictions†	
Бриоре	Sequence	ID No.	HLA type	SYFPEITHI	NIH
307 406	RLENYEDOLI	463	A0201	17	<5
397-400	KLEN I EDQLI	403	A3	15	<5
398-406	LENYEDQLI	464	B4402	19	N/A
398-407	LENYEDQLII	465	B4402	19	N/A
399-407	ENYEDQLII	466	B5101	17	19.36
399-408	ENYEDQLIIL	467	A26	20	N/A
400-408	NYEDQLIIL	468	A1	16	<5
400-409	NYEDQLILT	469	A1	16	<5
401-409	YEDQLIILT	470	A1	18	<5
401-403	1 EDQLILL	4/0	B4402	16	N/A
401 410	YEDQLIILTM	471	<u>A</u> 1	18	<5
401-410	TEDQUILIM	4/1	B4402	16	N/A
402-410	EDOLIILTM	472	A26	18	N/A
402-410	EDQLILIM	412	B2705	15	<5
406-415	IILTMELQKT	473	A0201	22	14.824
	IIL I MELQKI	4/3	A26	16	N/A
407-415	ILTMELQKT	474	A0201	21	29.137

†Scores are given from the two binding prediction programs referenced above (see example 3).. See also figure 56.

Example 54: SCP-1 416-442

<u>Table 54</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

Epitope	Sequence	Sequence	HLA type	HLA bindin	g predictions†		
	Sequence	ID No.	HLA type	SYFPEITHI	NIH		
424-432	KLTNNKEVE	475	A3	18	<5		
			A0201	24	74.768		
424-433	KLTNNKEVEL	476	A26	18	N/A		
			A3	18	<5		
			A0201	22	<5		
425-433	LTNNKEVEL	477	A26	21	N/A		
			B08	22	<5		
429-438	KEVELEELKK	478	A3	17	<5		
			A1	18	90		
430-438	EVELEELKK	479	A26	17	N/A		
430-130	EVELEELKK	479	A3	24	<5		
			B2705	15	<5		
430-430	EVELEELKKV	480	A0201	15	<5		
430-137	D V DECERENCE	400	A26	21	N/A		
			A0201	20	80.217		
431-439	VELEELKKV	481	A4402	15	N/A		
			B5101	17	ঠ		

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 57.

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Example 55: SCP-1 518-545

Table 55

Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Epitope	Sequence	Sequence	HLA type	HLA binding predictions†	
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIH
530-539	ETSDMTLELK	482	A26	21	N/A
531-539	TSDMTLELK	483	Al	16	15

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 58.

Example 56: SCP-1 545-578

Table 56

5

10 Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Epitope	Sequence	Sequence	HLA type	HLA binding	predictions†
Бриорс		ID No.	IIIA type	SYFPEITHI	NIH
548-556	NKKQEERML	484	B08	20	<5
553-562	ERMLTQIENL	485	A26	19'`	N/A
333-302	EKWET QIENE	403	B4402	· 17	N/A
			A0201	24	64.335
			B2705	, 21	150
554-562	RMLTQIENL	486	B2709	17	N/A
			B4402	15	N/A
555-562	MLTQIENL	487	" B08	16	<5
555-564	MLTQIENLQE	488	A3	16	<5
560-569	ENLQETETQL	489	A26	16	N/A
			A0201	22	87.586
561-569	NLQETETQL	490	A26	19	N/A
301-309	MEQUIEIQE	490	A3	15	<5
			B08	18	<5
<u>561-570</u>	NLQETETQLR	491	A3	15	6

†Scores are given from the two binding prediction programs referenced above (see example 3).. See also figure 59.

Example 57: SCP-1 559-585

Table 57

Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Epitope	Sequence	Sequence	HLA type	HLA binding predictions†		
Thrope	эециенсе	ID No.	ma type	SYFPEITHI	NIH	
567-576	TQLRNELEYV	492	A0201	16	161.729	
568-576	QLRNELEYV	493	A0201	24	32.765	
308-376	QLKIVELE I V	493	A3	16	<5	
571-580	NELEYVREEL	494	A0201	16	<5	
3/1-300		454	B4402	23	N/A	
			A0201	17	<5	
572-580	ELEYVREEL	495	A26	23	N/A	
			B08	20	<5	
573-580	LEYVREEL	496	B08	19	<5	
574-583	EYVREELKQK	497	A3	16	<5	
575-583	YVREELKQK	400	A26	17	N/A	
373-383		498	A3	27	<5	

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 60.

Example 58: SCP-1 665-701

Table 58

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Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Epitope	Saguenes	Sequence	HLA type	HLA binding	HLA binding predictions†		
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIH		
675-684	LLEEVEKAK V	499	A0201	27	31.026		
676-684	LEEVEKAKV	500	A0201	15	<5		
676-685	LEEVEKAKVI	501	A4402	22	N/A		
			B08	21	<5		
677-685	EEVEKAKVI	502	B4402	24	N/A		
			B5101	18	<5		
681-690	KAKVIADEA V	503	A0201	15	<5		
	KVIADEAVK L		A0201	21	6.542		
683 602		504	A26	22	N/A		
003-032			A3	25	<5		
			B4402	17	N/A		
			A0201	26	20.473		
			A26	22	N/A		
684-692	VIADEAVKL	505	A3	17	<5		
			B08	16	<5		
			B2705	15	N/A		
685-692	IADEAVKL	506	B08	17	<5		
005-092	INDEAVEL	200	B5101	21	N/A		

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 61.

Example 59: SCP-1 694-720

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<u>Table 59</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

Enitar a	C	Sequence	TIT A from a	HLA binding	predictions†
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	HIN
604 702	KEIDKRCQH	507	A3	16	<5
094-702	KEIDKKCQH	307	A4402	17	N/A
694-703	KEIDKRCQH	508	A3	17	_<5
094-703	K	308	B4402	15	N/A
695-703	EIDKRCOHK	509	A26	20	N/A
093-703	EDRICQIR	303	A3	20	<5
605 704	EIDKRCOHKI	510	A0201	16	<5
093-704	EDARCORA	310	A26	19	N/A
696-704	IDKRCQHKI	511	B08	17	<5
697-704	DKRCQHKI	512	B5101	16	N/A
698-706	KRCQHKIAE	513	B2705	16	60
698-707	KRCQHKIAE M	514	A26	15	N/A
500 505	D. GOTTUTA D. C	61.5	A26	15	N/A
	RCQHKIAEM	515	B2705	18	9
701-710	QHKIAEMVA L	516	A26	15	N/A
			A0201	15	<5
702-710	HKIAEMVAL	517	A26	16	N/A
			B4402	16	N/A
703-710	KIAEMVAL	518	B08	16	<5

†Scores are given from the two binding prediction programs referenced

[0386] above (see example 3)

10 **[0387]** See also figure 62.

Example 60: SCP-1 735-769

Table 60

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Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Enitone	Commence	Sequence	HLA type	HLA binding	predictions†	
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIH	
737-746	QEQSSLRASL	519	B4402	21	N.A.	
729 746	EQSSLRASL	520	A26	22	N.A.	
			B0702	15	6	
739-746	QSSLRASL	521	B08	19	<5	
			A0201	24	<5	
741-750	SLRASLEIEL	522	A26	17	N.A.	
			A3	16	<5	
			A0201	17	<5	
742-750	LRASLEIEL	523	B2705	23	2000	
			B2709	21	N.A.	
743-750	RASLEIEL	524	B5101	17	N.A.	
744_753	ASLEIELSNL	525	A0201	20	<5	
744-755	AUDEILEUNIE	J2J	A26	16	N.A.	
			A0201	25	<5	
745-753	SLEIELSNL	526	A26	22 .	N.A.	
745-755	BLEELBINE	320	A3	15	<5	
			B08	18	<5	
745-754	SLEIELSNLK	527	A1	15	18	
7 15 751	SEELESIVER	J21	A3	22	20	
746-754	LEIELSNI K	LEIELSNLK	528	B2705	16	30
			B4402	15	N.A.	
747-755	EIELSNLKA	529	Al	19	<5	
141-155	EIEEBIVEKA	329	A26	18	N.A.	
749-758	ELSNLKAELL	530	A0201	17	<5	
747-730	LLGIVLICALLL	330	A26	22	N.A.	
750-758	LSNLKAELL	531	B08	21	<5	
751-760	SNLKAELLSV	532	A0201	21	<5	
			A0201	26	5.599	
752-760	NLKAELLSV	533	A3	18	<5	
			B08	16	<5	
752-761	NLKAELLSV K	534	A3	30	30	
753-761	LKAELLSVK	535	A3	19	<5	
	LKAELLSVK K	536	A3	16	<5	
751.762	KAELLSVKK	527	A3	18	<5	
		537	B2705	18	30	
755-763	AELLSVKKQ	538	B4402	19	N.A.	

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 63.

Example 61: SCP-1 786-816

5 <u>Table 61</u> <u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

Epitope	Sequence	Sequence	HLA type	HLA binding	predictions†
Epitope	Sequence	ID No.	III.A type	SYFPEITHI	NIH
787-796	EKKDKKTQT	539	A26	19	N/A
707-190	F	333	B4402	15	N/A
788-796	KKDKKTQTF	540	B08	16	<5
700 750	idd)dc/Q11	340	B2705	16	<5
789-796		541	B08	16	<5
797-806	LLETPDIYW	542	A0201	16	<5
727 000	K	372	A3	21	90
798-806	LETPDIYWK	543	B2705	15	30
770-000	CETTOTTWK	343	B4402	16	N/A
	LETPDIYWK		A0201	15	7.944
798-807	CEILDIIWK	544	A26	15	N/A
	L.		A4402	24	N/A
799-807	ETPDIYWKL	545	A26	31	N/A
,,,,	ZII ZII WKL	343	B4402	16	N/A
800-807	TPDIYWKL	546	B08	16	<5
300-807	111)II WKL		B5101	19	N/A

†Scores are given from the two binding prediction programs referenced

[0390] above (see example 3)

10 [0391] See also figure 64.

Example 62: SCP-1 806-833

Table 62

Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Fritana	Saguanca	Sequence	HLA type	HLA binding predictions†		
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIH	
809-817	SKAVPSQTV	547	A0201	17	<5	
810-817	KAVPSQTV	548	B5101	19	N/A	
812-821	VPSQTVSRNF	549	B0702	18	N/A	
815-824	OTVSRNFTSV	550	A0201	16	<5	
013-024	QIVBRITISV	330	A26	16	N/A	
			A0201	16	11.426	
816-824	TVSRNFTSV	551	A26	15	N/A	
			A3	16	<5	
816-825	TVSRNFTSVD	552	A3	20	<5	
823-832	SVDHGISKDK	553	A3	21	<5	

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 65.

Example 63: SCP-1 826-853

Table 63

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Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Epitope	Sequence	Sequence	TOT A 4	HLA binding predictions†		
Epitope	ID No.		HLA type	SYFPEITHI	NIH	
829-838	SKDKRDYLWT	554	A1	18	<5	
832-840	KRDYLWTSA	555	B2705	16	600	
832-841	KRDYLWTSAK	556	A3	17	<5	
833-841	RDYLWTSAK	557	A3	23	<5	
033-041		337	B2705	18	15	
835-843	YLWTSAKNT	558	A0201	16	284.517	
835-844	YLWTSAKNTL	559	A0201	26	815.616	
033-044	TEWISARNIE	339	A26	16	N/A	
837-844	WTSAKNTL	560	B08	20	<5	
841-850	KNTLSTPLPK	561	A3	18	<5	
842-850	NTLSTPLPK	562	A3	16	<5	

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 66.

Example 64: SCP-1 832-859

<u>Table 64</u>
<u>Preferred Epitopes Revealed by Housekeeping Proteasome Digestion</u>

1 5. •4.	0	Sequence	TOT 4 4	HLA binding predictions†		
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIH	
832-840	KRDYLWTSA	563	B2705	16	600	
832-841	KRDYLWTSA K	564	A3	17	<5	
000 041	DDY TIME AT	5.65	A3	23	<5	
833-841	RDYLWTSAK	565	B2705	18	15	
835-843	YLWTSAKNT	566	A0201	16	284.517	
839-846	SAKNTLST	567	B08	16	<5	
841-850	KNTLSTPLPK	568	A3	18	<5	
842-850	NTLSTPLPK	569	A3	16	<5	
			A1	16	<5	
042.050	TO COUNT DIV A SZ	570	A26	19	N/A	
843-852	TLSTPLPKAY	TLSTPLPKAY 570	A3	18	<5	
			B4402	17	N/A	
	I		A 1	22	7.5	

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 67.

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Example 65: SSX-2 1-27

844-852 LSTPLPKAY

Table 65

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Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

TD	G	Sequence	TTT A 4	HILA binding	predictions†
Epitope	Sequence	DD No.	HLA type	SYFPEITHI	NIH
5-12	DAFARRPT	572	B5101	18	N/A
7-15	FARRPTVGA	573	A0201	15	<5
8-17	ARRPTVGAQI	574	A3	18	<5
9-17	DDDTVC A OI	575	B2705	23	1800
9-17	RRPTVGAQI	3/3	B2709	23	N/A
10-17	RPTVGAQI	576	B5101	20	N/A
13-21	VGAQIPEKI	577	B5101	20	125.84
14-21	GAQIPEKI	578	B5101	25	N/A
15-24	AQIPEKIQKA	579	A0201	16	<5
			A0201	21	6.442
16-24	QIPEKIQKA	580	A26	20	N/A
			B08	17	<5
16-25	OMENIONAE	581	A26	24	N/A
10-23	QIPEKIQKAF	201	_A3	16	<5
17-24	IPEKIQKA	582	B5101	19	N/A
			B0702	19	N/A
17-25	PEKIQKAF	583	B08	15	<5
			B2705	16	<5
18-25	PEKIQKAF	584	B08	16	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 68.

Example 66: Survivin 116-142

Table 66

Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Epitope	Sequence	Sequence	HLA type	HLA bindin	g predictions†
Epitope	Sequence	ID No.	III.A type	SYFPEITHI	NIH
116-124	ETNNKKKEF	585	A26	28	N/A
110-124	E IMMAKKET.	363	B08	20	<5
117-124	TNNKKKEF	586	B08	16	<5
122-131	KEFEETAKKV	587	A0201	15	71.806
123-131	EFEETAKKV	588	A26	15	N/A
123-131	EFECTARRY	366	B5101	15	5.324
127-134	TAKKVRRA	589	B5101	17	N/A
126-134	ETAKKVRRA	590	A26	24	N/A
128-136	AKKVRRAIE	591	B08	19	<5
129-138	KKVRRAIEQL	592	A0201	15	<5
			A0201	19	<5
			A26	23	N/A
130-138	KVRRAIEQL	593	A3	22	<5
			B08	17	<5
			B2705	16	30
130-139	KVRRAIEQLA	594	A3	19	<5
131-138	VRRAIEQL	595	B08	17	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 69.

Example 67: BAGE 1-35

Table 67

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Preferred Epitopes Revealed by Housekeeping Proteasome Digestion

Epitope	Sequence	Sequence	TIT A 4	HLA binding	predictions†
Epitope	Sequence	ID No.	HLA type	SYFPEITHI	NIE
24-31	SPVVSWRL	596	B08	19	<5
24-31	DI A ARMICI	390	B5101	17	N/A
21-29	KEESPVVSW	597	B4402	23	N/A
10.27	9-27 LMKEESPVV	598	A0201	22	5.024
19-27		390	B5101	15 .	<5
18-27	RLMKEESPVV	599	A0201	22	105.51
10-27			A3	18	<5
18-26	RLMKEESPV	600	A0201	21	257.342
16-20	KLWIKEESP V		A3	17	<5
14-22	LLQARLMKE	601	A0201	18	<5
17-22		001	A3	15	<5
İ			A0201	18	<5
13-22	QLLQARLMKE	602	A26	15	N/A
		1 [A3	15	<5

†Scores are given from the two binding prediction programs referenced above (see example 3). See also figure 70.

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Example 68

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Epitope Clusters.

Known and predicted epitopes are generally not evenly distributed across the sequences of protein antigens. As referred to above, we have defined segments of sequence containing a higher than average density of (known or predicted) epitopes as epitope clusters. Among the uses of epitope clusters is the incorporation of their sequence into substrate peptides used in proteasomal digestion analysis as described herein, or to otherwise inform the selection and design of such substrates. Epitope clusters can also be useful as vaccine components. Fuller discussions of the definition and uses of epitope clusters is found in PCT Publication No. WO 01/82963; PCT Publication No. WO 03/057823; and U.S. Patent Application No. 09/561,571 entitled EPITOPE CLUSTERS and in U.S. Patent Application No. 10/026,066 entitled "EPITOPE SYNCHRONIZATION IN ANTIGEN PRESENTING CELLS." Epitopes and epitope clusters for many of the TAA mentioned herein have been previously disclosed in PCT Publication No. WO 02/081646; in Patent Application No. 09/561,571; in U.S. Patent Application No. 10/117,937; U.S. Provisional Application Nos. 60/337,017 filed on November 7, 2001, and 60/363,210 filed on March 7, 2002, all entitled EPITOPE SEQUENCES. The teachings and embodiments disclosed in said publications and applications are contemplated as supporting principals and embodiments related to and useful in connection with the present invention.

For the TuAAs survivin (SEQ ID NO. 98) and GAGE-1 (SEQ ID NO. 96) the following tables (68-73) present 9-mer epitopes predicted for HLA-A2 binding using both the SYFPEITHI and NIH algorithms and the epitope density of regions of overlapping epitopes, and of epitopes in the whole protein, and the ratio of these two densities. (The ratio must exceed one for there to be a cluster by the above definition; requiring higher values of this ratio reflect preferred embodiments). Individual 9-mers are ranked by score and identified by the position of their first amino in the complete protein sequence. Each potential cluster from a protein is numbered. The range of amino acid positions within the complete sequence that the cluster covers is indicated, as are the rankings of the individual predicted epitopes it is made up of.

Table 68

HLA-A2 Epitope cluster analysis for Survivin (NIH algorithm)

Length of protein sequence: 142 amino acids

Number of 9-mers: 134

Number of 9-mers with NIH score = 5: 2

Cluster	AA	Peptide	Start	Score	Pepti	des/AAs	Ratio
		Rank	Position		Cluster	Whole Pro.	
1	13-28	1	13	10.26	0.125	0.014	8.875
SEQ ID NO:603		2	20	4.919			

Table 69

HLA-A2 Epitope cluster analysis for Survivin (SYFPEITHI algorithm)

Length of protein sequence: 142 amino acids

Number of 9-mers: 134

Number of 9-mers with SYFPEITHI score = 15: 10

Cluster	AA	Peptide	Start	Score	Peptie	les/AAs	Ratio
		Rank	Position		Cluster	Whole Pro.	
1	13-28	5	13	17	0.125	0.070	1.775
SEQ ID NO:603		4	20	18			
2	79-111	8	79	15	0.182	0.070	2.597
SEQ ID NO:604		9	81	15			
		6	88	17			
		1	96	23			
		7	97	16			
_		10	103	15			
3	130-141	2	130	19	0.167	0.070	2.381
SEQ ID NO:605		3	133	19			

Table 70

HLA-A2 Epitope cluster analysis for GAGE-1 (NIH algorithm)

Length of protein sequence: 138 amino acids

Number of 9-mers: 130

Number of 9-mers with NIH score = 5:5

Cluster	AA	Peptide	Start	Score	Peptio	les/AAs	Ratio
		Rank	Position		Cluster	Whole Pro.	
1	116-133	1	123	1999.734	0.278	0.036	7.667
SEQ ID NO:606		2	121	161.227			
		3	125	49.834			
		4	117	37.362			
		5	116	6.381			

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<u>Table 71</u>

HLA-A2 Epitope cluster analysis for GAGE-1 (SYFPEITHI algorithm)

Length of protein sequence: 138 amino acids

Number of 9-mers: 130

Number of 9-mers with SYFPEITHI score = 5: 6

Cluster	AA	Peptide	Start	Score	Peptie	des/AAs	Ratio
		Rank	Position		Cluster	Whole Pro.	
1	116-133	1	116	22	0.333	0.043	7.667
SEQ ID NO:606		2	123	22			
		3	125	22			
		4	117	17			
		5	120	16			4
		6	121	15			

<u>Table 72</u> **HLA-A2** Epitope cluster analysis for BAGE (NIH algorithm)

Length of protein sequence: 43 amino acids

Number of 9-mers included: 35

Number of 9-mers with NIH score = 5: 4

Cluster	AA	Peptide	Start	Score	Peptid	les/AAs	Ratio
		Rank	Position		Cluster	Whole Pro.	
1	7-17	2	7	98.267	0.182	0.093	1.955
SEQ ID NO:607		3	9	11.426			
2	18-27	1	. 18	257.342	0.200	0.093	2.151
SEQ ID NO:608		4	19	5.024			

5 <u>Table 73</u>

15

HLA-A2 Epitope cluster analysis for BAGE (SYFPEITHI algorithm)

Length of protein sequence: 43 amino acids

Number of 9-mers included: 35

Number of 9-mers with SYFPEITHI score = 15: 10

Cluster	AA	Peptide	Start	Score	Pepti	ides/AAs	Ratio
		Rank	Position		Cluster	Whole Pro.	
1	2-27	6	2	18	0.308	0.233	1.323
SEQ ID NO:609		9	6	16			
		1	7	23			
		3	9	21			
1		5	11	19			
ł		7	14	18			
		4	18	21			
		2	19	22			
2	30-39	8	30	17	0.200	0.233	0.858
SEQ ID NO:610		10	31	15			

[0406] The embodiments of the invention are applicable to and contemplate variations in the sequences of the target antigens provided herein, including those disclosed in the various databases that are accessible by the world wide web. Specifically for the specific sequences disclosed herein, variation in sequences can be found by using the provided accession numbers to access information for each antigen.

TYROSINASE PROTEIN; SEQ ID NO 2

1 MLLAVLYCLL WSFQTSAGHF PRACVSSKNL MEKECCPPWS GDRSPCGQLS GRGSCQNILL

61 SNAPLGPQFP FTGVDDRESW PSVFYNRTCQ CSGNFMGFNC GNCKFGFWGP NCTERRLLVR

121 RNIFDLSAPE KDKFFAYLTL AKHTISSDYV IPIGTYGQMK NGSTPMFNDI NIYDLFVWMH

- 181 YYVSMDALLG GSEIWRDIDF AHEAPAFLPW HRLFLLRWEQ EIQKLTGDEN FTIPYWDWRD
- 5 241 AEKCDICTDE YMGGQHPTNP NLLSPASFFS SWQIVCSRLE EYNSHQSLCN GTPEGPLRRN
 - 301 PGNHDKSRTP RLPSSADVEF CLSLTQYESG SMDKAANFSF RNTLEGFASP LTGIADASQS
- 361 SMHNALHIYM NGTMSQVQGS ANDPIFLLHH AFVDSIFEQW LRRHRPLQEV 10 YPEANAPIGH
- 421 NRESYMVPFI PLYRNGDFFI SSKDLGYDYS YLQDSDPDSF QDYIKSYLEQ
 ASRIWSWLLG
 - 481 AAMVGAVLTA LLAGLVSLLC RHKRKQLPEE KOPLLMEKED YHSLYOSHL
- 15 SSX-2 PROTEIN; SEQ ID NO 3
 - 1 MNGDDAFARR PTVGAQIPEK IQKAFDDIAK YFSKEEWEKM KASEKIFYVY MKRKYEAMTK
- 61 LGFKATLPPF MCNKRAEDFQ GNDLDNDPNR GNQVERPQMT FGRLQGISPK 20 IMPKKPAEEG
 - 121 NDSEEVPEAS GPONDGKELC PPGKPTTSEK IHERSGPKRG EHAWTHRLRE RKOLVIYEEI
 - 181 SDPEEDDE
- 25 PSMA PROTEIN; SEQ ID NO 4
 - 1 MWNLLHETDS AVATARRPRW LCAGALVLAG GFFLLGFLFG WFIKSSNEAT NITPKHNMKA
- 61 FLDELKAENI KKFLYNFTQI PHLAGTEQNF QLAKQIQSQW KEFGLDSVEL
- 30 AHYDVLLSYP
 - 121 NKTHPNYISI INEDGNEIFN TSLFEPPPPG YENVSDIVPP FSAFSPQGMP EGDLVYVNYA
 - 181 RTEDFFKLER DMKINCSGKI VIARYGKVFR GNKVKNAQLA GAKGVILYSD PADYFAPGVK
- 35 241 SYPDGWNLPG GGVQRGNILN LNGAGDPLTP GYPANEYAYR RGIAEAVGLP SIPVHPIGYY
 - 301 DAQKLLEKMG GSAPPDSSWR GSLKVPYNVG PGFTGNFSTQ KVKMHIHSTN EVTRIYNVIG
- 361 TLRGAVEPDR YVILGGHRDS WVFGGIDPQS GAAVVHEIVR SFGTLKKEGW 40 RPRRTILFAS
 - 421 WDAEEFGLLG STEWAEENSR LLQERGVAYI NADSSIEGNY TLRVDCTPLM YSLVHNLTKE
 - 481 LKSPDEGFEG KSLYESWTKK SPSPEFSGMP RISKLGSGND FEVFFQRLGI ASGRARYTKN
- 45 541 WETNKFSGYP LYHSVYETYE LVEKFYDPMF KYHLTVAQVR GGMVFELANS IVLPFDCRDY
 - 601 AVVLRKYADK IYSISMKHPQ EMKTYSVSFD SLFSAVKNFT EIASKFSERL ODFDKSNPIV
- 661 LRMMNDQLMF LERAFIDPLG LPDRPFYRHV IYAPSSHNKY AGESFPGIYD 50 ALFDIESKVD
 - 721 PSKAWGEVKR QIYVAAFTVO AAAETLSEVA
- Homo sapiens tyrosinase (oculocutaneous albinism IA) (TYR), mRNA.;
- 55 ACCESSION NM_000372
 - VERSION NM 000372.1 GI:4507752
 - SEQ ID NO 2
 - /translation="MLLAVLYCLLWSFQTSAGHFPRACVSSKNLMEKECCPPWSGDRS

	PCGQLSGRGSCQNILLSNAP	LGPQFPFTGVD	DRESWPSVFYNI	RTCQCSGNFMG	FNCGN					
5	CKFGFWGPNCTERRLLVRRN	IFDLSAPEKDK	FFAYLTLAKHT:	ISSDYVIPIGT	YGQMK					
5	NGSTPMFNDINIYDLFVWMH	YYVSMDALLGG	SEIWRDIDFAHI	EAPAFLPWHRL	FLLRW					
	EQEIQKLTGDENFTIPYWDW	RDAEKCDICTDI	EYMGGQHPTNPI	NLLSPASFFSS	NQIVC					
10	SRLEEYNSHQSLCNGTPEGP	LRRNPGNHDKSI	RTPRLPSSADVI	EFCLSLTQYES	GSMDK					
	AANFSFRNTLEGFASPLTGI	ADASQSSMHNAI	LHI YMNGTMSQ'	VQGSANDPIFL	LHHAF					
15	VDSIFEQWLRRHRPLQEVYPEANAPIGHNRESYMVPFIPLYRNGDFFISSKDLGYDYS									
13	YLQDSDPDSFQDYIKSYLEQ		AMVGAVLTALLI EDYHSLYQSHL'		RKQLP					
20	SEQ ID NO 5 ORIGIN									
	1 atcactgtag ttcctgcaga	tagtagctgg	aaagagaaat	ctgtgactcc	aattagccag					
	61 ccttgtgagg tgtggagttt	actagaggaa	gaatgctcct	ggctgttttg	tactgcctgc					
25	121 ccagacctcc tgatggagaa	gctggccatt	tccctagagc	ctgtgtctcc	tctaagaacc					
	181 ggaatgctgt	ccaccgtgga	gcggggacag	gagtccctgt	ggccagcttt					
20	caggcagagg 241 ttcctgtcag	aatatccttc	tgtccaatgc	accacttggg	cctcaatttc					
30	301 ggtggatgac	cgggagtcgt	ggccttccgt	cttttataat	aggacctgcc					
	agtgctctgg 361 caacttcatg	ggattcaact	gtggaaactg	caagtttggc	ttttggggac					
35	caaactgcac 421 agagagacga	ctcttggtga	gaagaaacat	cttcgatttg	agtgccccag					
	agaaggacaa 481 attttttgcc	tacctcactt	tagcaaagca	taccatcagc	tcagactatg					
	tcatccccat 541 agggacctat	ggccaaatga	aaaatggatc	aacacccatg	tttaacgaca					
40	tcaatattta 601 tgacctcttt	gtctggatgc	attattatgt	gtcaatggat	gcactgcttg					
	ggggatctga 661 aatctggaga									
45	ggcatagact 721 cttcttgttg									
	acttcactat 781 tccatattgg									
	agtacatggg 841 aggtcagcac									
50	cctcttggca 901 gattgtctgt									
	atggaacgcc 961 cgagggacct									
55	caaggctccc									
55	1021 ctcttcagct gttccatgga .									
	1081 taaagctgcc	aatttcagct	ttagaaatac	actggaagga	tttgctagtc					

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- 5 541 gattcacgag agatctggac ccaaaagggg ggaacatgcc tggacccaca gactgcgtga
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- Homo sapiens folate hydrolase (prostate-specific membrane antigen) $1 \ \, \text{(FOLH1), mRNA.}$ ACCESSION NM 004476

VERSION NM_004476 VERSION NM_004476.1 GI:4758397

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- 35 AVVHEIVRSFGTLKKEGWRPRRTILFASWDAEEFGLLGSTEWAEENSRLLQERGVAYI NADSSIEGNYTLRVDCTPLMYSLVHNLTKELKSPDEGFEGKSLYESWTKKSPSPEFSG
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SEQ ID NO 7 ORIGIN

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- 121 gcgaattcca gcctgcaggg ctgataagcg aggcattagt gagattgaga 55 gagactttac
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Human melanocyte-specific (pmel 17) gene, exons 2-5, and complete cds.

ACCESSION U20093

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30 VERSION U20093.1 GI:1142634 SEQ ID NO 70

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Homo sapiens kallikrein 3, (prostate specific antigen) (KLK3), mRNA.

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    VERSION
                P08217 GI:119255
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    PRAME Homo sapiens preferentially expressed antigen in melanoma
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                NM 006115
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40 //

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CEA Homo sapiens carcinoembryonic antigen-related cell adhesion molecule 5 (CEACAM5), mRNA.

ACCESSION NM_004363

45 VERSION NM_004363.1 GI:11386170

SEQ ID NO 88

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10

SEQ ID NO 89 ORIGIN 1 ctcagggcag agggaggaag gacagcagac cagacagtca cagcagcctt gacaaaacgt 15 61 tcctggaact caagctcttc tccacagagg aggacagagc agacagcaga gaccatggag 121 tetecetegg ecceteceea cagatggtge atcecetgge agaggeteet gctcacagcc 181 tcacttctaa ccttctggaa cccgcccacc actgccaagc tcactattga 20 atccacqccq 241 ttcaatgtcg cagaggggaa ggaggtgctt ctacttgtcc acaatctgcc ccagcatctt 301 tttggctaca gctggtacaa aggtgaaaga gtggatggca accgtcaaat tataggatat 25 361 gtaataggaa ctcaacaagc taccccaggg cccqcataca gtggtcgaga gataatatac 421 cccaatgcat ccctgctgat ccagaacatc atccagaatg acacaggatt ctacacccta 481 cacgtcataa agtcagatct tgtgaatgaa gaagcaactg gccagttccg 30 ggtatacccg 541 gagctgccca agccctccat ctccagcaac aactccaaac ccgtggagga caaggatgct 601 gtggccttca cctgtgaacc tgagactcag gacgcaacct acctgtggtg ggtaaacaat 35 661 cagageetee eggteagtee caggetgeag etgtecaatg geaacaggae cctcactcta 721 ttcaatgtca caagaaatga cacagcaagc tacaaatgtg aaacccagaa cccagtgagt

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Her2/Neu Human tyrosine kinase-type receptor (HER2) mRNA, complete
cds.

ACCESSION M11730

VERSION M11730.1 GI:183986

SEQ ID NO 90

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SEQ ID NO 91

ORIGIN Chromosome 17q21-q22.

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	ctgacctgca	~~~~~~~~~	*****		
	oogaoocgca	geeecagee	tgaatatgtg	aaccagccag	atgttcggcc
	togococgan	agggccctct	acctactace	caacetast~	ataggaatat
	ייייייייייייייייייייייייייייייייייייי		sootgotgee	cyacciyolg	gugodadidi
	aagactctct	ccccagggaa	gaatggggtc	gtcaaagacg	tttttacctt
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	gcagattgco 2641 cgctcggaac 2701 tcggctgctc 2761 gtggatggcg 2821 ttatggtgtg 2881 agcccgggag 2941 caccattgat 3001 aagattccgg 3061 ggtcatccag 3121 actgctggag 3181 gcaggcttc 3241 ccgcagctca 3301 agaggaggcc 3361 tggtgacctg 3421 ccctctacag 3481 cgttgcccc 3541 ccagcccct 3601 ggaaaggcc 3661	gcagattgcc 2641 aaggggatga cgctcggaac 2701 gtgctggtca tcggctgctg 2761 gacattgacg gtggatggcg 2821 ctggagtcca ttatggtgtg 2881 actgtgtggg agcccgggag 2941 atccctgacc caccattgat 3001 gtctacatga aagattccgg 3061 gagttggtgt ggtcatccag 3121 aatgaggact actgctggag 3181 gacgatgaca gcagggettc 3241 ttctgtccag ccgcagctca 3301 tctaccagga agaggagcc 3361 cccaggtctc tggtgacctg 3421 ggaatgggg ccctctacag 3481 cggtacagtg cgttgcccc 3541 ctgacctgca ccagcccct 3601 tcgccccgag ggaaagggcc 3661 aagactctct	gcagattgcc 2641 aaggggatga gctacctgga cgctcggaac 2701 gtgctggtca agagtccaa tcggctgctg 2761 gacattgacg agacagagta gtggatggcg 2821 ctggagtcca ttctccgccg ttatggtgg 2881 actgtgtggg agctgatgac agccgggag 2941 atccctgacc tgctggaaaa caccattgat 3001 gtctacatga tcatggtcaa aagattccgg 3121 aatgaggact tgggcccagc actgctggag 3181 gacgatgaca tgggggacct gcagggettc 3241 ttctgtccag accctgccc ccgcagctca 3301 tctaccagga gtggcggtgg agaggaggcc 3421 ggaatgggg cagccaaggg cctctacag 3421 ggaatgggg cagccaaggg ccctctacag 3481 cggtacagtg aggacccac cgttgcccc 3541 ctgacctgca gccccagcc ccagcccct 3601 tcgccccgag agggccctct ggaaagggcc 3601 tcgccccgag agggccctct ggaaagggcc 3601 aagactctct ccccagggaa	gcagattgcc 2641 aaggggatga gctacctgga ggatgtgcgg cgctcggaac 2701 gtgctggtca agagtcccaa ccatgtcaaa tcggctgctg 2761 gacattgacg agacagagta ccatgcagat gtggatggcg 2821 ctggagtcca ttctccgccg gcggttcacc ttatggttg 2881 actgtgtggg agctgatgac ttttggggcc agcccgggag 2941 atccctgacc tgctggaaaa gggggagggc caccattgat 3001 gtctacatga tcatggtcaa atgttggatg aggtcaccag 3121 aatgaggat tgggccagc cagtccctg ggtcatccag 3181 gacgatgaca tgggggacct ggtggatgct gcagggctc 3241 ttctgtccag accctgccc gggcgttggc ccgcaggctc 3301 tctaccagga gtggcggtgg ggacctgac agaggaggcc 3341 tcccaggtct cactggcac ctccgaaggg tggtgacctg 3361 cccaggtct cactggcac ctccgaaggg cccctctacag 3421 ggaatgggg cagccaaggg gctgcaaagc ccctctcacag 3481 cggtacagtg aggacccac agtaccctg ccgttgcccc 3541 ctgacctgca gccccagcc tgaatatgtg ccagccccc 3541 ctgacctgca gccccagcc tgaatatgtg ccagcccccc 3541 ctgacctgca gccccagcc tgaatatgtg ccagcccccc 3541 ctgacctgca gccccagcc tgaatatgtg ccagcccccc 3541 ctgacctga agggccctct gcctgccc ggaaagggcc 3601 tcgccccag agggccctct gcctgccc ggaaagggcc 3601 tcgccccag agggccctct gcctgccc ggaaagggcc 3601 aagactctc ccccagggaa gaatggggtc	gcagattgcc 2641 aaggggatga gctacctgga ggatgtgcgg ctcgtacaca cgctcggaac 2701 gtgctggtca agagtccaa ccatgtcaaa attacagact tcggctgctg 2761 gacattgacg agacagagta ccatgcagat gggggatggcg 2821 ctggagtcca ttctccgccg gcggttcacc caccagagtg ttatggtgg 2881 actgtgtgg agctgatgac ttttggggcc aaaccttacg agcccggggg 2941 atccctgacc tgctggaaaa gggggagggg ctgcccagccacttgat 3001 gctacatga tcatggtcaa atgttggatg attgactcggggaggagggg ctgcaccaggggtcaccagggggagggggggggg

3721 gtggagaacc ccgagtactt ga	caccccag ggaggagctg cccctcagcc											
ccaccctcct												
3781 cetgeettea geecageett eg	acaacete tattactggg accaggacec											
accagagcgg												
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gtacctgggt												
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tgtcctcagg												
3961 gagcagggaa ggcctgactt ct	gctggcat caagaggtgg gagggccctc											
10 cgaccacttc												
4021 caggggaacc tgccatgcca gg	aacctgtc ctaaggaacc ttccttcctg											
cttgagttcc												
4081 cagatggctg gaaggggtcc ag	cctcgttg gaagaggaac agcactgggg											
agtctttgtg												
15 4141 gattctgagg ccctgcccaa tg	agactcta gggtccagtg gatgccacag											
cccagcttgg												
	tactgaaa gccttaggga agctggcctg											
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	aacaaaag cgacccattc agagactgtc											
20 cctgaaacct												
	gaacagca atggtgtcag tatccaggct											
ttgtacagag												
	ettttttg ttttgttttt ttaaagacga											
aataaagacc 25 4441 caggggagaa tgggtgttgt at	ggggaggc aagtgtgggg ggtccttctc											
cacacccact	ggggagge aagegegggg ggeeeeeee											
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30												
W comicans mDNA for CCD1 washein												
n.sapiens mkNA for SCPT protein.	H.sapiens mRNA for SCP1 protein.											
H.sapiens mRNA for SCP1 protein. ACCESSION X95654 VERSION X95654.1 GI:1212982												
ACCESSION X95654												

/translation="MEKQKPFALFVPPRSSSSQVSAVKPQTLGGDSTFFKSFNKCTED DLEFPFAKTNLSKNGENIDSDPALQKVNFLPVLEQVGNSDCHYQEGLKDSDLENSEGL SRVFSKLYKEAEKIKKWKVSTEAELRQKESKLQENRKIIEAQRKAIQELQFGNEKVSL KLEEGIQENKDLIKENNATRHLCNLLKETCARSAEKTKKYEYEREETRQVYMDLNNNI

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KQFEKIAEELKGTEQELIGLLQAREKEVHDLEIQLTAITTSEQYYSKEVKDLKTELEN
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ETETQLRNELEYVREELKQKRDEVKCKLDKSEENCNNLRKQVENKNKYIEELQQENKA
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EKAKVIADEAVKLQKEIDKRCQHKIAEMVALMEKHKHQYDKIIEERDSELGLYKSKEQ

EQSSLRASLEIELSNLKAELLSVKKQLEIEREEKEKLKREAKENTATLKEKKDKKTQT
FLLETPEIYWKLDSKAVPSQTVSRNFTSVDHGISKDKRDYLWTSAKNTLSTPLPKAYT
VKTPTKPKLQQRENLNIPIEESKKKRKMAFEFDINSDSSETTDLLSMVSEEETLKTLY
RNNNPPASHLCVKTPKKAPSSLTTPGPTLKFGAIRKMREDRWAVIAKMDRKKKLKEAE
KLFV"

15

SEQ ID NO 93 ORIGIN

agaccctggg

- 1 gccctcatag accgtttgtt gtagttcgcg tgggaacagc aacccacggt
 ttcccgatag

 20 61 ttcttcaaag atatttacaa ccgtaacaga gaaaatggaa aagcaaaagc
 cctttgcatt

 121 gttcgtacca ccgagatcaa gcagcagtca ggtgtctgcg gtgaaacctc
- 181 aggcgattcc actttcttca agagtttcaa caaatgtact gaagatgatt
 25 tggagtttcc
 - 241 atttgcaaag actaatctct ccaaaaatgg ggaaaacatt gattcagatc ctgctttaca
 - 301 aaaagttaat ttcttgcccg tgcttgagca ggttggtaat tctgactgtc actatcagga
- 30 361 aggactaaaa gactctgatt tggagaattc agagggattg agcagagtgt tttcaaaact
 - 421 gtataaggag gctgaaaaga taaaaaaatg gaaagtaagt acagaagctg aactgagaca
- 481 gaaagaaagt aagttgcaag aaaacagaaa gataattgaa gcacagcgaa 35 aagccattca
 - 541 ggaactgcaa tttggaaatg aaaaagtaag tttgaaatta gaagaaggaa tacaagaaaa
 - 601 taaagattta ataaaagaga ataatgccac aaggcattta tgtaatctac tcaaagaaac

	661	ctgtgctaga	tctgcagaaa	agacaaagaa	atatgaatat	gaacgggaag
	aaaccaggca					
	721	agtttatatg	gatctaaata	ataacattga	gaaaatgata	acageteate
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5	781	tgtgcaagct	gagaattcca	gactggaaat	gcattttaag	ttaaaggaag
	attatgaaaa					
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	agcaggtatc					
	901	actactattg	atccaaatca	ctgagaaaga	aaataaaatg	aaagatttaa
10	catttctgct					
	961	agaggaatcc	agagataaag	ttaatcaatt	agaggaaaag	acaaaatta
	agagtgaaaa					
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	ctaataaagc					
	1201	tagagctgct	cattcgtttg	tggttactga	atttgaaact	actgtctgca
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	1261	attattgaga	acagaacagc	aaagattgga	aaaaaatgaa	gatcaattga
	aaatacttac					
			caaaagaaat	caagtgagct	ggaagagatg	actaagctta
	caaataacaa					
25			cttgaagaat	tgaaaaaagt	cttgggagaa	aaggaaacad
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	aactaattgg					
20			gccagagaga	aagaagtaca	tgatttggaa	atacagttaa
30	ctgccattac					
			cagtattatt	caaaagaggt	taaagatcta	aaaactgago
	ttgaaaacga			h b b		- 4- 4- 4 4
		_	aatactgaat	taacttcaca	Cigcaacaag	ctttcactag
35	aaaacaaaga		~~~~~~	2+2+42444	20220+	
,,	1681		yaaacaagug	atatgaccct	ayaactcadg	aaccagcaag
	aagatattaa		2246224224	aaaggatgtt	assaussats	
			aaycaayaay	aaayyatytt	yaaacaada	yaaaatutti
	aagaaacaga					

	1801	aacccaatta	agaaatgaac	tagaatatgt	gagagaagag	ctaaaacaga
	aaagagatga	ı				
	1861	agttaaatgt	aaattggaca	agagtgaaga	aaattgtaac	aatttaagga
	aacaagttga	L				
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	aaaaaaagg					
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	tagagttaga					
	2041	actagaaagt	gccaaacaga	aatttggaga	aatcacagac	acctatcaga
10	aagaaattga			•		
	2101	ggacaaaaag	atatcagaag	aaaatctttt	ggaagaggtt	gagaaagcaa
	aagtaatagc					
	2161	tgatgaagca	gtaaaattac	agaaagaaat	tgataagcga	tgtcaacata
	aaatagctga					
15	2221	aatggtagca	cttatggaaa	aacataagca	ccaatatgat	aagatcattg
	aagaaagaga					
	2281	ctcagaatta	ggactttata	agagcaaaga	acaagaacag	tcatcactga
	gagcatcttt					
	2341	ggagattgaa	ctatccaatc	tcaaagctga	acttttgtct	gttaagaagc
20	aacttgaaat					
	2401	agaaagagaa	gagaaggaaa	aactcaaaag	agaggcaaaa	gaaaacacag
	ctactcttaa					
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25	2521	attggattct	aaagcagttc	cttcacaaac	tgtatctcga	aatttcacat
	cagttgatca					
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	ctttatctac					
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30	agcaaagaga					
	2701	aaacttgaat	atacccattg	aagaaagtaa	aaaaaagaga	aaaatggcct
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	aagaagagac					
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	gagctataag				-	2 2 3

2941 aaaaatgcgg gaggaccgtt gggctgtaat tgctaaaatg gatagaaaaa aaaaactaaa

- 3001 agaagctgaa aagttatttg tttaatttca gagaatcagt gtagttaagg agcctaataa
- 5 3061 cgtgaaactt atagttaata ttttgttctt atttgccaga gccacatttt atctggaagt
 - 3121 tgagacttaa aaaatacttg catgaatgat ttgtgtttct ttatattttt agcctaaatg
- 3181 ttaactacat attgtctgga aacctgtcat tgtattcaga taattagatg 10 attatatatt
 - 3241 gttgttactt tttcttgtat tcatgaaaac tgtttttact aagttttcaa atttgtaaag
 - 3301 ttagcctttg aatgctagga atgcattatt gagggtcatt ctttattctt tactattaaa
- 3361 atattttgga tgcaaaaaaa aaaaaaaaaa aaa
 //

Homo sapiens synovial sarcoma, X breakpoint 4 (SSX4), mRNA.

20 ACCESSION NM 005636

• VERSION NM_005636.1 GI:5032122

SEQ ID NO 94

/translation="MNGDDAFARRPRDDAQISEKLRKAFDDIAKYFSKKEWEKMKSSEKIVY

25 VYMKLNYEVMTKLGFKVTLPPFMRSKRAADFHGNDFGNDRNHRNQVERPQMTFG
SLQRIFPKIMPKKPAEEENGLKEVPEASGPQNDGKQLCPPGNPSTLEKINKTSGPKRG
KHAWTHRLRERKQLVVYEEISDPEEDDE"

SEQ ID NO 95

- 30 ORIGIN
 - 1 atgaacggag acgacgcctt tgcaaggaga cccagggatg atgctcaaat atcagagaag
 - 61 ttacgaaagg ccttcgatga tattgccaaa tacttctcta agaaagagtg ggaaaagatg
- 35 121 aaatcctcgg agaaaatcgt ctatgtgtat atgaagctaa actatgaggt catgactaaa
 - 181 ctaggtttca aggtcaccct cccacctttc atgcgtagta aacgggctgc agacttccac

241 gggaatgatt ttggtaacga tcgaaaccac aggaatcagg ttgaacgtcc tcagatgact

- 301 ttcggcagcc tccagagaat cttcccgaag atcatgccca agaagccagc agaggaagaa .
- 5 361 aatggtttga aggaagtgcc agaggcatct ggcccacaaa atgatgggaa acagctgtgc
 - 421 cccccgggaa atccaagtac cttggagaag attaacaaga catctggacc caaaaggggg
- 481 aaacatgcct ggacccacag actgcgtgag agaaagcagc tggtggttta 10 tgaagagatc
 - 541 agcgaccctg aggaagatga cgagtaactc ccctcg

U19142. Human GAGE-1 prot...[gi:914898]

15 LOCUS HSU19142 646 bp mRNA linear

DEFINITION Human GAGE-1 protein mRNA, complete cds.

ACCESSION U19142

VERSION U19142.1 GI:914898

20 SEQ ID No. 96

/translation="MSWRGRSTYRPRPRRYVEPPEMIGPMRPEQFSDEVEPATPEEGE

$\label{eq:patorope} {\tt PATORODPAAAQEGEDEGASAGQGPKPEADSQEQGHPQTGCECEDGPDGQEMDPPNPE} \\ {\tt EVKTPEEEMRSHYVAQTGILWLLMNNCFLNLSPRKP"}$

25

SEQ ID NO. 97

- 1 ctgccgtccg gactcttttt cctctactga gattcatctg tgtgaaatat gagttggcga
- 61 ggaagatcga cctatcggcc tagaccaaga cgctacgtag agcctcctga 30 aatgattggg
 - 121 cctatgcggc ccgagcagtt cagtgatgaa gtggaaccag caacacctga agaaggggaa
 - 181 ccagcaactc aacgtcagga tcctgcagct gctcaggagg gagaggatga gggagcatct
- 241 gcaggtcaag ggccgaagcc tgaagctgat agccaggaac agggtcaccc acagactggg
 - 301 tgtgagtgtg aagatggtcc tgatgggcag gagatggacc cgccaaatcc agaggaggtg

361 aaaacgcctg aagaagagat gaggtctcac tatgttgccc agactgggat tctctggctt

- 421 ttaatgaaca attgcttctt aaatctttcc ccacggaaac cttgagtgac tgaaatatca
- 5 481 aatggcgaga gaccgtttag ttcctatcat ctgtggcatg tgaagggcaa tcacagtgtt
 - 541 aaaagaagac atgctgaaat gttgcaggct gctcctatgt tggaaaattc ttcattgaag
- 601 ttctcccaat aaagctttac agccttctgc aaagaaaaaa aaaaaa 10 //

NM 001168. Homo sapiens bacu...[gi:4502144]

LOCUS BIRC5 1619 bp mRNA linear

DEFINITION Homo sapiens baculoviral IAP repeat-containing 5

15 (survivin) (BIRC5), mRNA.

ACCESSION NM 001168

VERSION NM_001168.1 GI:4502144

SEQ ID NO. 98

20 /translation="MGAPTLPPAWQPFLKDHRISTFKNWPFLEGCACTPERMAEAGFI

HCPTENEPDLAQCFFCFKELEGWEPDDDPIEEHKKHSSGCAFLSVKKQFEELTLGEFL KLDRERAKNKIAKETNNKKKEFEETAKKVRRAIEOLAAMD"

- 25 SEQ ID NO. 99
 - 1 ccgccagatt tgaatcgcgg gacccgttgg cagaggtggc ggcggcggca tgggtgccc
 - 61 gacgttgccc cctgcctggc agccctttct caaggaccac cgcatctcta cattcaagaa
- 30 121 ctggcccttc ttggagggct gcgcctgcac cccggagcgg atggccgagg ctggcttcat
 - 181 ccactgcccc actgagaacg agccagactt ggcccagtgt ttcttctgct tcaaggagct
- 241 ggaaggctgg gagccagatg acgaccccat agaggaacat aaaaagcatt 35 cgtccggttg
 - 301 cgctttcctt tctgtcaaga agcagtttga agaattaacc cttggtgaat
 - 361 ggacagagaa agagccaaga acaaaattgc aaaggaaacc aacaataaga agaaagaatt

421 tgaggaaact gcgaagaaag tgcgccgtgc catcgagcag ctggctgcca tggattgagg 481 cctctggccg gagctgcctg gtcccagagt ggctgcacca cttccagggt ttattccctg 5 541 gtgccaccag ccttcctgtg ggccccttag caatgtctta ggaaaggaga tcaacatttt 601 caaattagat gtttcaactg tgctcctgtt ttgtcttgaa agtggcacca gaggtgcttc 661 tgcctgtgca gcgggtgctg ctggtaacag tggctgcttc tctctctct tctcttttt 10 721 gggggctcat ttttgctgtt ttgattcccg ggcttaccag gtgagaagtg agggaggaag 781 aaggcagtgt cccttttgct agagctgaca gctttgttcg cgtgggcaga gccttccaca 15 841 gtgaatgtgt ctggacctca tgttgttgag gctgtcacag tcctgagtgt ggacttggca 901 ggtgcctgtt gaatctgagc tgcaggttcc ttatctgtca cacctgtgcc tcctcagagg 20 tgacttgtgt 1021 gtgatgagag aatggagaca gagtccctgg ctcctctact gtttaacaac atggctttct 1081 tattttgttt gaattgttaa ttcacagaat agcacaaact acaattaaaa ctaagcacaa 25 1141 agccattcta agtcattggg gaaacggggt gaacttcagg tggatgagga gacagaatag 1201 agtgatagga agcgtctggc agatactcct tttgccactg ctgtgtgatt agacaggccc 1261 agtgagccgc ggggcacatg ctggccgctc ctccctcaga aaaaggcagt 30 ggcctaaatc 1321 ctttttaaat gacttggctc gatgctgtgg gggactggct gggctgctgc aggccgtgtg 1381 tetgteagee caacetteae atetgteaeg ttetecaeae gggggagaga cgcagtccgc 35 1441 ccaggtcccc gctttctttg gaggcagcag ctcccgcagg gctgaagtct ggcgtaagat 1501 gatggatttg attcgccctc ctccctgtca tagagctgca gggtggattg ttacagcttc

1561 gctggaaacc tctggaggtc atctcggctg ttcctgagaa ataaaaagcc tgtcatttc

11

5

U06452. Human melanoma an...[gi:476131]

LOCUS HSU06452 1524 bp mRNA linear

DEFINITION Human melanoma antigen recognized by T-cells (MART-1) mRNA.

10 ACCESSION U06452

VERSION U06452.1 GI:476131

SEQ ID NO.100

/translation="MPREDAHFIYGYPKKGHGHSYTTAEEAAGIGILTVILGVLLLIG

15

CWYCRRRNGYRALMDKSLHVGTQCALTRRCPQEGFDHRDSKVSLQEKNCEPVVPNAPP AYEKLSAEQSPPPYSP"

SEQ ID NO. 101

- 20 1 agcagacaga ggactctcat taaggaaggt gtcctgtgcc ctgaccctac aagatgccaa
 - 61 gagaagatgc tcacttcatc tatggttacc ccaagaaggg gcacggccac tcttacacca
- 121 cggctgaaga ggccgctggg atcggcatcc tgacagtgat cctgggagtc 25 ttactgctca
 - 181 tcggctgttg gtattgtaga agacgaaatg gatacagagc cttgatggat aaaagtcttc
 - 241 atgttggcac tcaatgtgcc ttaacaagaa gatgcccaca agaagggttt gatcatcggg
- 30 301 acagcaaagt gtctcttcaa gagaaaaact gtgaacctgt ggttcccaat gctccacctg
 - 361 cttatgagaa actctctgca gaacagtcac caccacctta ttcaccttaa gagccagcga
- 421 gacacctgag acatgctgaa attatttctc tcacactttt gcttgaattt 35 aatacagaca
 - 481 totaatgtto tootttggaa tggtgtagga aaaatgcaag coatototaa taataagtoa
 - 541 gtgttaaaat tttagtaggt ccgctagcag tactaatcat gtgaggaaat gatgagaaat

601 attaaattgg gaaaactcca tcaataaatg ttgcaatgca tgatactatc tgtgccagag 661 gtaatgttag taaatccatg gtgttatttt ctgagagaca gaattcaagt gggtattctg 5 721 gggccatcca atttctcttt acttgaaatt tggctaataa caaactagtc aggttttcga 781 accttgaccg acatgaactg tacacagaat tgttccagta ctatqqaqtq ctcacaaagg 841 atacttttac aggttaagac aaagggttga ctqqcctatt tatctqatca 10 agaacatgtc 901 agcaatgtct ctttgtgctc taaaattcta ttatactaca ataatatatt gtaaagatcc 961 tatagetett tttttttgag atggagttte gettttgttg cecaggetgg agtgcaatgg 15 1021 cgcgatcttg gctcaccata acctccgcct cccaggttca agcaattctc ctgccttagc 1081 ctcctgagta gctgggatta caggcgtgcg ccactatgcc tgactaattt tgtagtttta 1141 gtagagacgg ggtttctcca tgttggtcag gctggtctca aactcctgac 20 ctcaggtgat 1201 ctgcccgcct cagcctccca aagtgctgga attacaggcg tgagccacca cgcctggctg 1261 gatcctatat cttaggtaag acatataacg cagtctaatt acatttcact tcaaggctca 25 1321 atgctattct aactaatgac aagtattttc tactaaacca gaaattggta gaaggattta 1381 aataagtaaa agctactatg tactgcctta gtgctgatgc ctgtgtactg ccttaaatgt 1441 acctatggca atttagctct cttgggttcc caaatccctc tcacaagaat 30 gtgcagaaga 1501 aatcataaag gatcagagat tctg // U19180. Human B melanoma ...[gi:726039] 35 LOCUS HSU19180 1004 bp mRNA linear DEFINITION Human B melanoma antigen (BAGE) mRNA, complete cds. ACCESSION U19180 VERSION U19180.1 GI:726039

SEQ IS NO. 102

/translation="MAARAVFLALSAQLLQARLMKEESPVVSWRLEPEDGTALCFIF"

SEQ ID NO. 103

35

- 5 1 cgccaattta gggtctccgg tatctcccgc tgagctgctc tgttcccggc ttagaggacc
 - 61 aggagaaggg ggagctggag gctggagcct gtaacaccgt ggctcgtctc actctggatg
- 121 gtggtggcaa cagagatggc agcgcagctg gagtgttagg agggcggcct 10 gagcggtagg
 - 181 agtggggctg gagcagtaag atggcggcca gagcggtttt tctggcattg tctgccagc
 - 241 tgctccaagc caggctgatg aaggaggagt cccctgtggt gagctggagg ttggagcctg
- 301 aagacggcac agctetgtge tteatettet gaggttgtgg cagecaeggt gatggagaeg
 - 361 gcagctcaac aggagcaata ggaggagatg gagtttcact gtgtcagcca ggatggtctc
- 421 gatctcctga cctcgtgatc cgcccgcctt ggccttccaa agtgccgaga 20 ttacagcgat
 - 481 gtgcattttg taagcacttt ggagccacta tcaaatgctg tgaagagaaa tgtacccaga
 - 541 tgtatcatta tccttgtgct gcaggagccg gctcctttca ggatttcagt cacatcttcc
- 25 601 tgctttgtcc agaacacatt gaccaagctc ctgaaagatg taagtttact acgcatagac
 - 661 ttttaaactt caaccaatgt atttactgaa aataacaaat gttgtaaatt ccctgagtgt
- 721 tattctactt gtattaaaag gtaataatac ataatcatta aaatctgagg 30 gatcattgcc
 - 781 agagattgtt ggggagggaa atgttatcaa cggtttcatt gaaattaaat ccaaaaagtt
 - 841 atttcctcag aaaaatcaaa taaagtttgc atgtttttta ttcttaaaac attttaaaaa
 - 901 ccactgtaga atgatgtaaa tagggactgt gcagtatttc tgacatatac tataaaatta
 - 961 ttaaaaagtc aatcagtatt caacatcttt tacactaaaa agcc

The teachings and embodiments disclosed in any of the publications, including patents, patent publications and non-patent publications, disclosed herein are contemplated as supporting principals and embodiments related to and useful in connection with the present invention.

The invention illustratively described herein suitably may be practiced in the absence of any element or elements, limitation or limitations which is not specifically disclosed herein. The terms and expressions which have been employed are used as terms of description and not of limitation, and there is no intention that in the use of such terms and expressions indicates the exclusion of equivalents of the features shown and described or portions thereof. It is recognized that various modifications are possible within the scope of the invention claimed. Thus, it should be understood that although the present invention has been specifically disclosed by preferred embodiments and optional features, modification and variation of the concepts herein disclosed may be resorted to by those skilled in the art, and that such modifications and variations are considered to be within the scope of the embodiments of this invention.

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WHAT IS CLAIMED IS:

1. A polypeptide, comprising a component selected from the group consisting of:

- (i) a polypeptide epitope having the sequence as disclosed in TABLE 1B;
- (ii) an epitope cluster comprising the polypeptide of (i);
- (iii) a polypeptide having substantial similarity to (i) or (ii);
- (iv) a polypeptide having functional similarity to any of (i) through (iii); and
- (v) a nucleic acid encoding the polypeptide of any of (i) through (iv).
- 2. The polypeptide of claim 1, wherein the polypeptide is immunologically active.
- 3. The polypeptide of claim 1, wherein the polypeptide is less than about 30 amino acids in length.
- 4. The polypeptide of claim 1, wherein the polypeptide is 8 to 10 amino acids in length.
- 5. The polypeptide of claim 1, wherein the substantial or functional similarity comprises addition of at least one amino acid.
- 6. The polypeptide of claim 5, wherein the at least one additional amino acid is at an N-terminus of the polypeptide.
- 7. The polypeptide of claim 1, wherein the substantial or functional similarity comprises a substitution of at least one amino acid.
- 8. The polypeptide of claim 1, the polypeptide having affinity to an HLA-A2 molecule.
- 9. The polypeptide of claim 8, wherein the affinity is determined by an assay of binding.
- 10. The polypeptide of claim 8, wherein the affinity is determined by an assay of restriction of epitope recognition.
- 11. The polypeptide of claim 8, wherein the affinity is determined by a prediction algorithm.
- 12. The polypeptide of claim 1, the polypeptide having affinity to an HLA-B7 or HLA-B51 molecule.
 - 13. The polypeptide of claim 1, wherein the polypeptide is a housekeeping epitope.
- 14. The polypeptide of claim 1, wherein the polypeptide corresponds to an epitope displayed on a tumor cell.
- 15. The polypeptide of claim 1, wherein the polypeptide corresponds to an epitope displayed on a neovasculature cell.
 - 16. The polypeptide of claim 1, wherein the polypeptide is an immune epitope.
 - 17. The polypeptide of claim 1, wherein the polypeptide is encoded by a nucleic acid.

18. A composition comprising the polypeptide of claim 1 and a pharmaceutically acceptable adjuvant, carrier, diluent, or excipient.

- 19. The composition of claim 18, where the adjuvant is a polynucleotide.
- 20. The composition of claim 19 wherein the polynucleotide comprises a dinucleotide.
- 21. The composition of claim 20 wherein the dinucleotide is CpG.
- 22. The composition of claim 18, wherein the adjuvant is encoded by a polynucleotide.
- 23. The composition of claim 18 wherein the adjuvant is a cytokine.
- 24. The composition of claim 23 wherein the cytokine is GM-CSF.
- 25. The composition of claim 18 further comprising a professional antigen-presenting cell (pAPC).
 - 26. The composition of claim 25, wherein the pAPC is a dendritic cell.
 - 27. The composition of claim 18, further comprising a second epitope.
 - 28. The composition of claim 27, wherein the second epitope is a polypeptide.
 - 29. The composition of claim 27, wherein the second epitope is a nucleic acid.
- 30. The composition of claim 27, wherein the second epitope is a housekeeping epitope.
 - 31. The composition of claim 27, wherein the second epitope is an immune epitope.
- 32. A composition comprising the nucleic acid of claim 1 and a pharmaceutically acceptable adjuvant, carrier, diluent, or excipient.
 - 33. A recombinant construct comprising the nucleic acid of Claim 1.
- 34. The construct of claim 33, further comprising a plasmid, a viral vector, a bacterial vector, or an artificial chromosome.
- 35. The construct of claim 33, further comprising a sequence encoding at least one feature selected from the group consisting of a second epitope, an IRES, an ISS, an NIS, and ubiquitin.
 - 36. A purified antibody that specifically binds to the polypeptide of claim 1.
- 37. A purified antibody that specifically binds to a peptide-MHC protein complex comprising the polypeptide of claim 1.
- 38. The antibody of claim 36 or claim 37, wherein the antibody is a monoclonal antibody.
 - 39. A multimeric MHC-peptide complex comprising the polypeptide of claim 1.
- 40. An isolated T cell expressing a T cell receptor specific for an MHC-peptide complex, the complex comprising the polypeptide of claim 1.
 - 41. The T cell of claim 40, produced by an in vitro immunization.
 - 42. The T cell of claim 40, isolated from an immunized animal.
 - 43. A T cell clone comprising the T cell of claim 40.

- 44. A polyclonal population of T cells comprising the T cell of claim 40.
- 45. A pharmaceutical composition comprising the T cell of claim 40 and a pharmaceutically acceptable adjuvant, carrier, diluent, or excipient.
- 46. An isolated protein molecule comprising the binding domain of a T cell receptor specific for an MHC-peptide complex, the complex comprising the epitope of claim 1.
 - 47. The protein of claim 46, wherein the protein is multivalent.
 - 48. An isolated nucleic acid encoding the protein of claim 46.
 - 49. A recombinant construct comprising the nucleic acid of claim 48.
- 50. A host cell expressing a recombinant construct, the construct comprising the nucleic acid of claim 1, or the construct encoding a protein molecule comprising the binding domain of a T cell receptor specific for an MHC-peptide complex.
- 51. The host cell of claim 50, wherein the host cell is a dendritic cell, macrophage, tumor cell, or tumor-derived cell.
- 52. The host cell of claim 50, wherein the host cell is a bacterium, fungus, or protozoan.
- 53. A composition comprising the host cell of claim 50 and a pharmaceutically acceptable adjuvant, carrier, diluent, or excipient.
- 54. A composition comprising at least one component selected from the group consisting of the epitope of claim 1; the composition of claim 18, 32, or 45, the construct of claim 33; the T cell of claim 40, a host cell expressing a recombinant construct comprising a nucleic acid encoding a T cell receptor binding domain specific for an MHC-peptide complex and a composition comprising the same, and a host cell expressing a recombinant construct comprising the nucleic acid of claim 1 and a composition comprising the same.
 - 55. A method of treating an animal, comprising: administering to an animal the composition of claim 54.
- 56. The method of claim 55, wherein the administering step comprises a mode of delivery selected from the group consisting of transdermal, intranodal, perinodal, oral, intravenous, intradermal, intramuscular, intraperitoneal, mucosal, aerosol inhalation, and instillation.
- 57. The method of claim 55, further comprising a step of assaying to determine a characteristic indicative of a state of a target cell or target cells.
- 58. The method of claim 57, comprising a first assaying step and a second assaying step, wherein the first assaying step precedes the administering step, and wherein the second assaying step follows the administering step.
- 59. The method of claim 58, further comprising a step of comparing the characteristic determined in the first assaying step with the characteristic determined in the second assaying step to obtain a result.

60. The method of claim 59, wherein the result is selected from the group consisting of: evidence of an immune response, a diminution in number of target cells, a loss of mass or size of a tumor comprising target cells, a decrease in number or concentration of an intracellular parasite infecting target cells.

- 61. A method of evaluating immunogenicity of an immunogenic composition, comprising:
 - administering to an animal the composition of claim 54; and evaluating immunogenicity based on a characteristic of the animal.
 - 62. The method of claim 61, wherein the animal is MHC-transgenic.
 - 63. A method of evaluating immunogenicity, comprising:

 in vitro stimulation of a T cell with the composition of claim 54; and evaluating immunogenicity based on a characteristic of the T cell.
 - 64. The method of claim 63, wherein the stimulation is a primary stimulation.
 - 65. A method of making a passive/adoptive immunotherapeutic, comprising:

 combining the T cell of claim 40, or a host cell expressing a recombinant construct
 comprising a nucleic acid encoding a T cell receptor binding domain specific for an MHCpeptide complex, or a host cell expressing a recombinant construct comprising the nucleic
 acid of claim 1 with a pharmaceutically acceptable adjuvant, carrier, diluent, or excipient.
- 66. A method of determining specific T cell frequency comprising the step of contacting T cells with a MHC-peptide complex comprising the polypeptide of claim 1.
- 67. The method of claim 66, wherein the contacting step comprises at least one feature selected from the group consisting of immunization, restimulation, detection, and enumeration.
- 68. The method of Claim 66, further comprising ELISPOT analysis, limiting dilution analysis, flow cytometry, in situ hybridization, the polymerase chain reaction or any combination thereof.
- 69. A method of evaluating immunologic response, comprising the method of claim 66 carried out prior to and subsequent to an immunization step.
 - 70. A method of evaluating immunologic response, comprising:

 determining frequency, cytokine production, or cytolytic activity of T cells, prior
 to and subsequent to a step of stimulation with MHC-peptide complexes comprising the
 polypeptide of claim 1.
 - 71. A method of diagnosing a disease comprising:

 contacting a subject tissue with at least one component selected from the group
 consisting of the T cell of claim 40, the host cell of claim 50, the antibody of claim 36, and
 the protein of claim 46; and

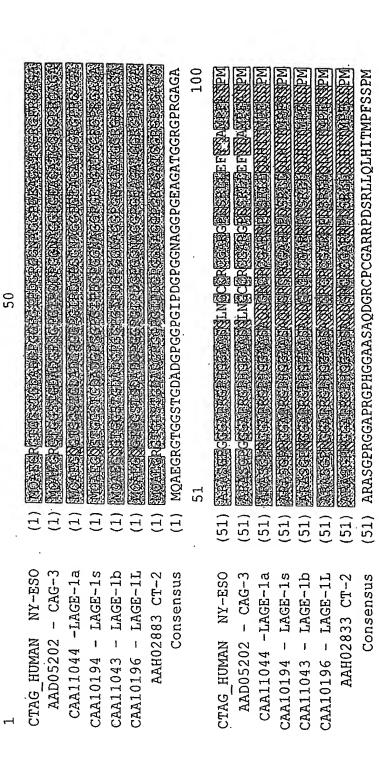
diagnosing the disease based on a characteristic of the tissue or of the component.

- 72. The method of claim 71, wherein the contacting step takes place in vivo.
- 73. The method of claim 71, wherein the contacting step takes place in vitro.
- 74. A method of making a vaccine, comprising:

combining at least one component selected from the group consisting of the polypeptide of claim 1; the composition of claim 18, 32, 45, or 53; the construct of claim 33; the T cell of claim 40, and the host cell of claim 50, with a pharmaceutically acceptable adjuvant, carrier, diluent, or excipient.

- 75. A computer readable medium having recorded thereon the sequence of any one of SEQ ID NOS: 108-610, in a machine having a hardware or software that calculates the physical, biochemical, immunologic, or molecular genetic properties of a molecule embodying said sequence.
- 76. A method of treating an animal comprising combining the method of claim 55 combined with at least one mode of treatment selected from the group of radiation therapy, chemotherapy, biochemotherapy, and surgery.
- 77. An isolated polypeptide comprising an epitope cluster from a target-associated antigen having the sequence as disclosed in Tables 68-73, wherein the amino acid sequence consists of not more than about 80% of the amino acid sequence of the antigen.
 - 78. A vaccine or immunotherapeutic product comprising the polypeptide of claim 77.
 - 79. An isolated polynucleotide encoding the polypeptide of claim 77.
- 80. A vaccine or immunotherapeutic product comprising the polynucleotide of claim 79.
 - 81. The polynucleotide of claim 79 or 80, wherein the polynucleotide is DNA.
 - 82. The polynucleotide of claim 79 or 80, wherein the polynucleotide is RNA.

FIG. 1A



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•	(101)	(101)	(101)	(101)	(101)	(101)	(101)	(101)		(151)	(151)	(151)	(151)	(151)	(151)	(151)	(151)
	CTAG_HUMAN NY-ESO	AAD05202 - CAG-3	CAA11044 -LAGE-1a	CAA10194 - LAGE-1s	CAA11043 - LAGE-1b	CAA10196 - LAGE-1L	AAH02833 CT-2	Consensus		CTAG HUMAN NY-ESO	AAD05202 - CAG-3	CAA11044 -LAGE-1a	CAA10194 - LAGE-1s.	CAA11043 - LAGE-1b	CAA10196 - LAGE-1L	AAH02833 CT-2	Consensus

FIG. 1B

FNVMFSAPHI FNVMESAPHI FNVMFSAPHI (201) (201) (201) (201)(181)(181)(181)(181)CTAG_HUMAN' NY-ESO AAD05202 - CAG-3 AAH02833 CT-2 CAA11044 -LAGE-1a CAA10194 - LAGE-1s CAA11043 - LAGE-1b CAA10196 - LAGE-1L Consensus

FIG. 1C

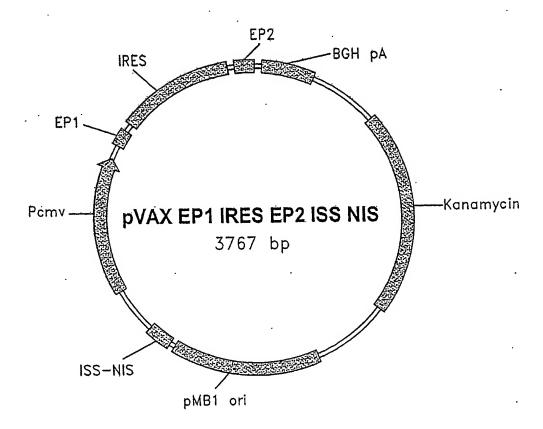
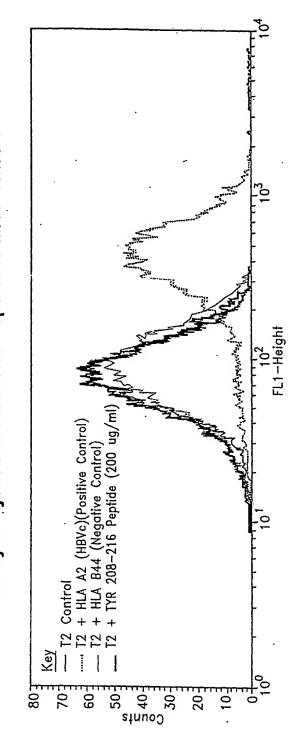


FIG. 2

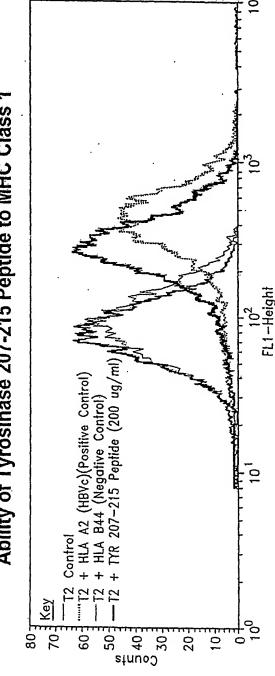
FACscan Analysis of Binding Assay to Determine the Binding Ability of Tyrosinase 208-216 Peptide to MHC Class 1

FIG. 3A



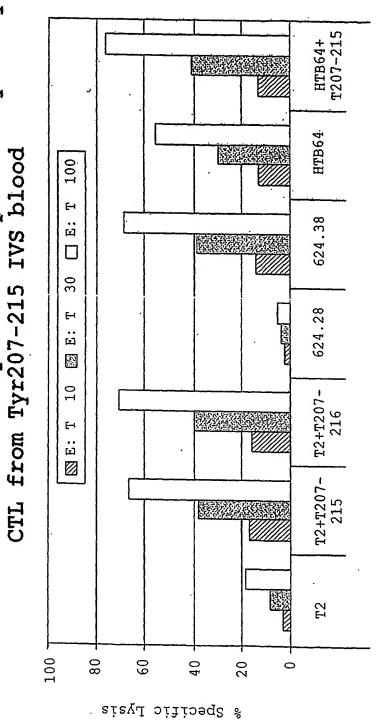
F1 (HLA A2 Peptide) = 3.13F1 (TYR 208-216 Peptide) = 0.01

FACscan Analysis of Binding Assay to Determine the Binding Ability of Tyrosinase 207-215 Peptide to MHC Class 1



F1 (HLA A2 Peptide) = 3.13F1 (TYR 207-215 Peptide) = 2.00

HLA A2 restricted and tyrosinase specific lysis by



CTL from Tyr 207-215 IVS blood

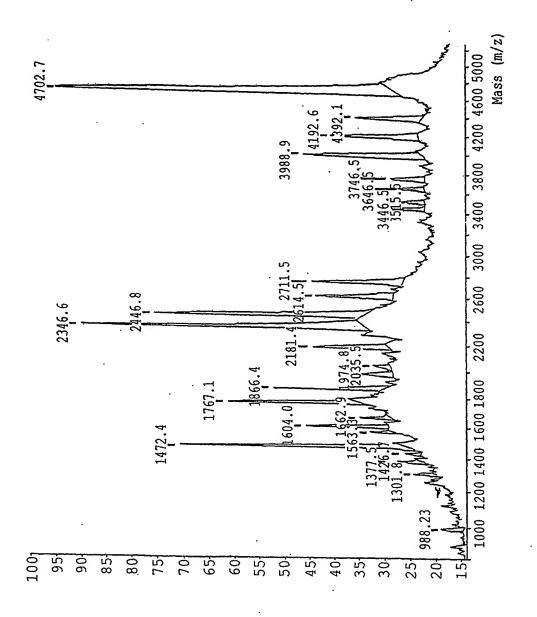
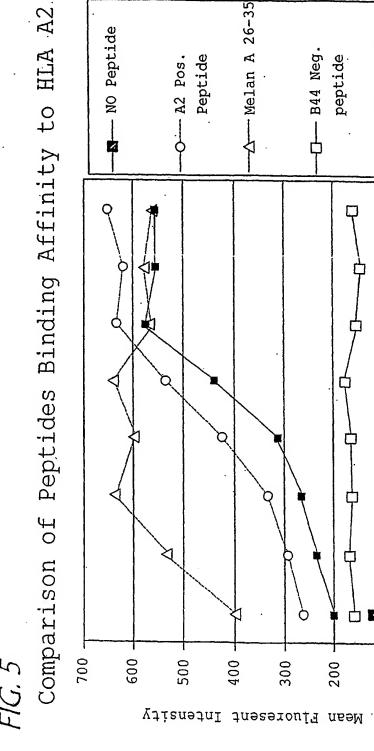


FIG. 4

SSX 41-49

100



Concentration of Peptides (ug/ml)

800

400

200

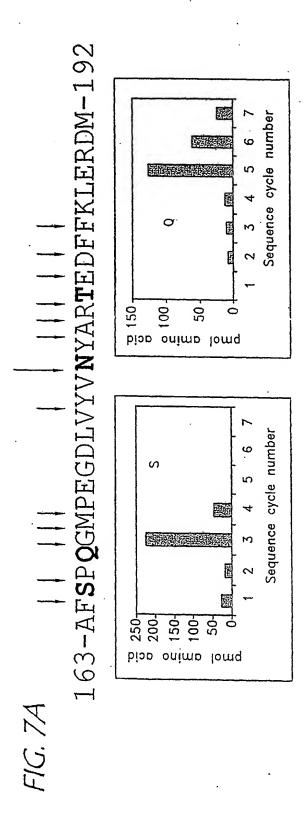
10,0

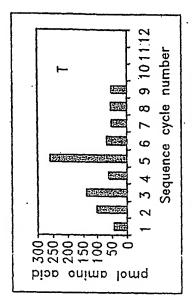
50

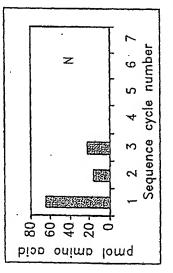
SSX241-49 specific lysis by CTL from peptide injected HHD1 mice ET 100 + SSX2 41-49 12 ET 30 -@-CTL (SSX2 41-49) CTL (SSX2 41-49) T2 ET ET 80-100. 120 90 40-

FIG. C

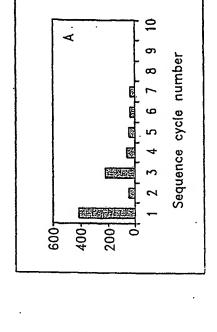
% Specific Lysis

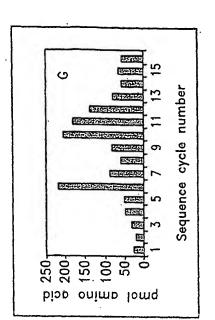


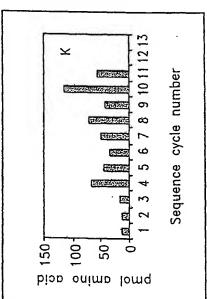


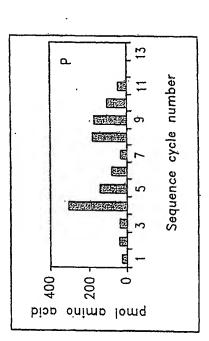


Pool sequencing of PSMA_163-192 Digested for 60 min by proteasome

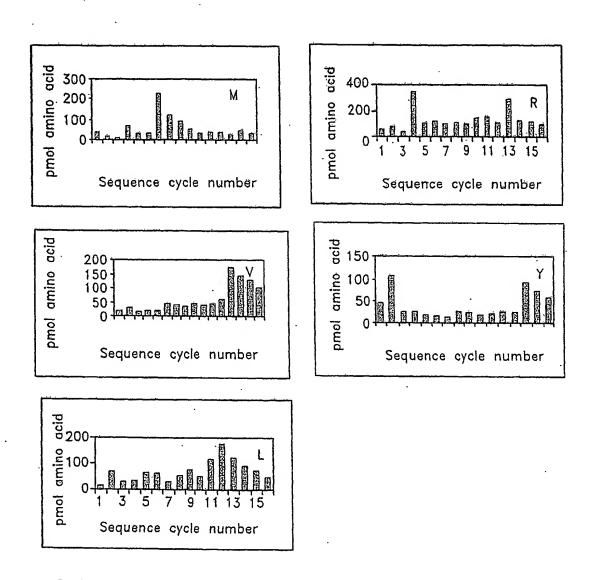








Pool sequencing of PSMA_163-192 Digested for 60 min by proteasome



Pool sequencing of PSMA_163-192 Digested for 60 min by proteasome

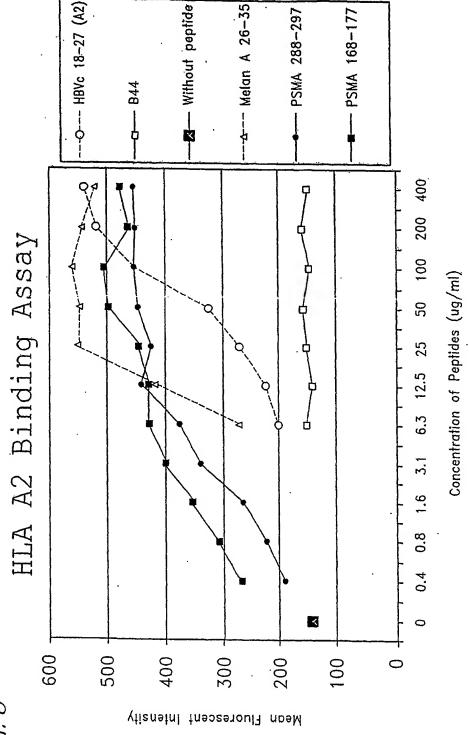
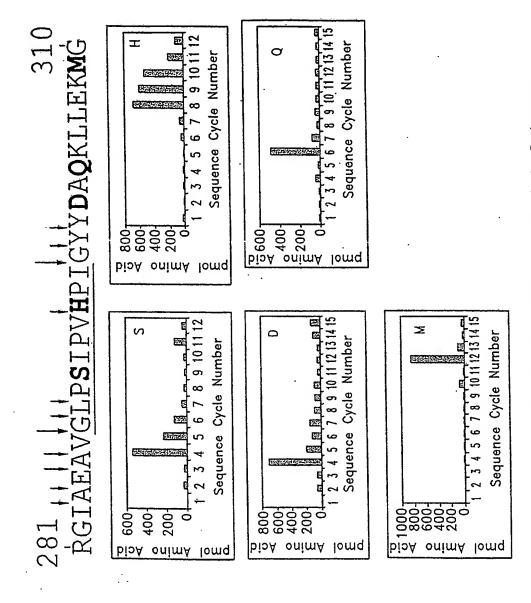
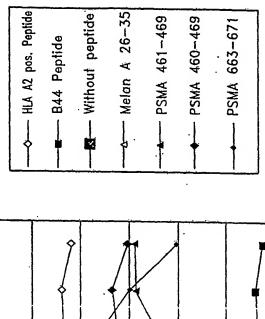


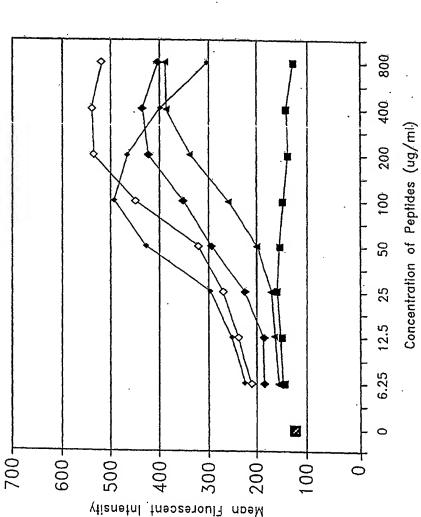
FIG. 8



Pool sequencing of PSMA_281_310 Digested for 60 min by Proteasome

Comparison of Peptides Binding Affinity to HLA A2 Assay by Binding





Autologous DC Present Al Peptide to CD8 T cell

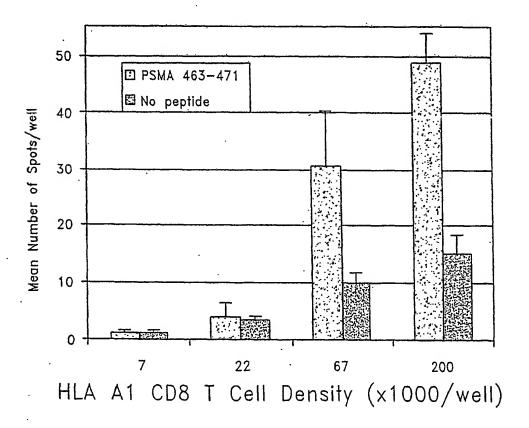
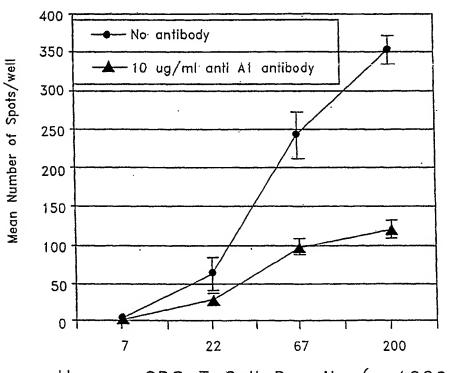


FIG. 11

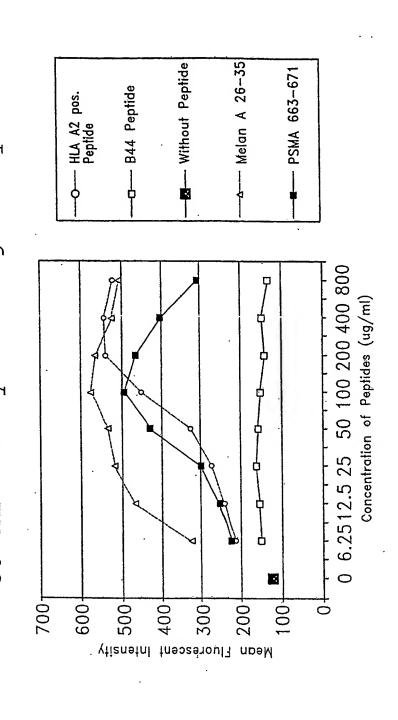
Secretion of IFNgama Was Blocked by Anti-A1 Antibody



Human CD8 T Cell Density (x 1000/well)

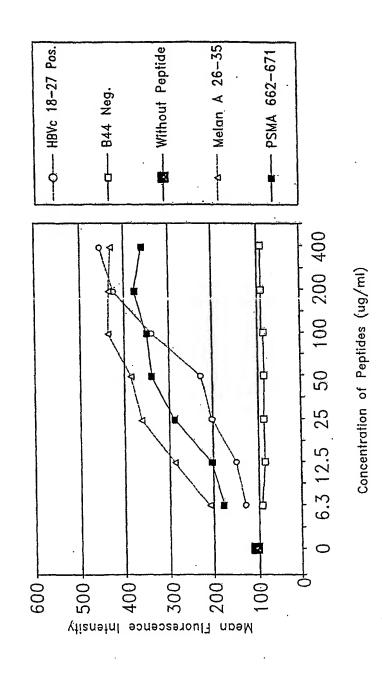
FIG. 12

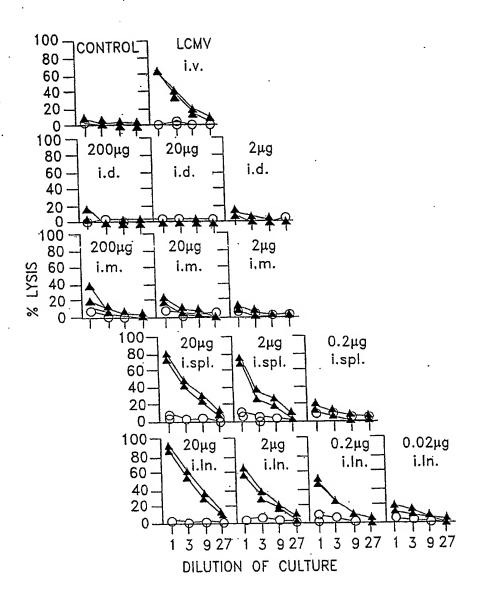
Comparison of Peptides Binding Affinity to HLA A2 by Binding Assay



Comparison of Peptides Binding Affinity to HLA A2 by Binding Assay

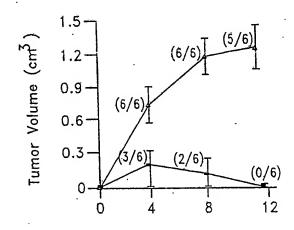
FIG. 14





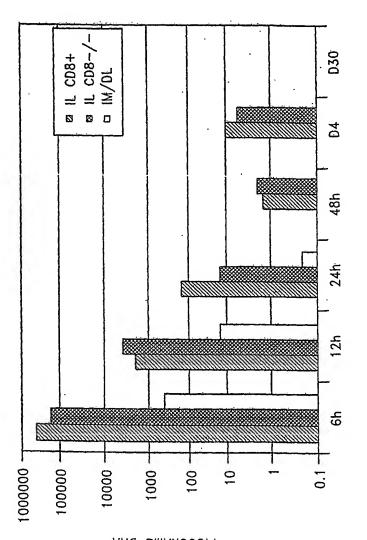
Graphs show lysis of unpulsed EL4 cells (open circles) and EL4 cells pulsed with gp33 peptide (solid triangles). Symbols represent individual mice and one of three similar experiments is shown.

FIG. 15

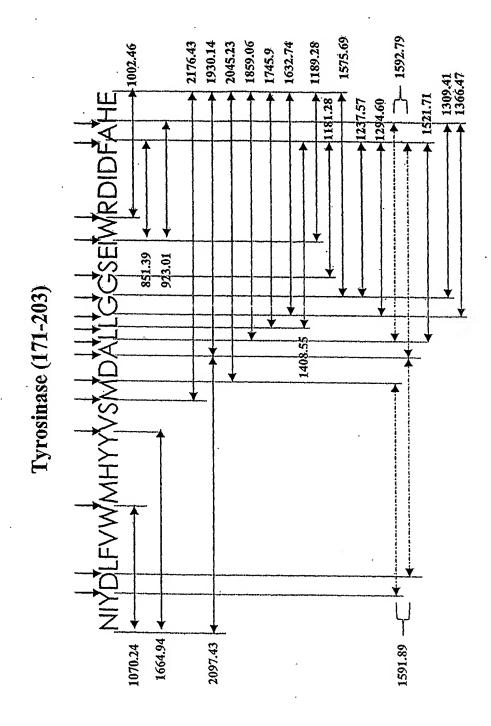


Days after tumor challenge

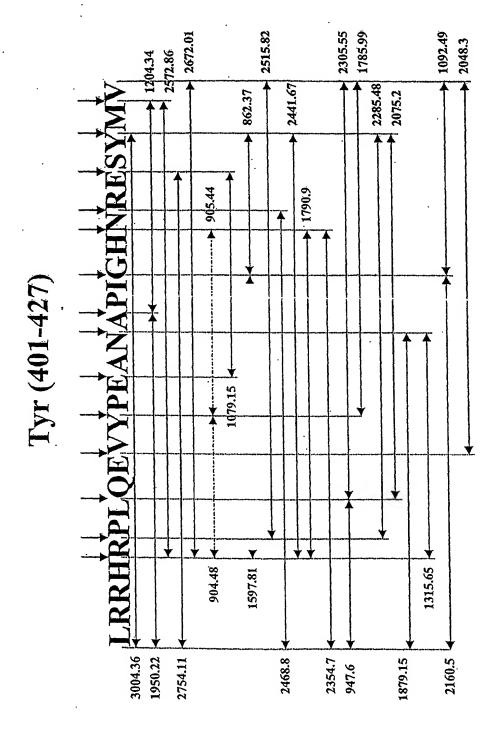
Mean tumor volumes \pm 1SD are shown for mice immunized with pEFGPL33A DNA (solid circles) or control pEGFP-N3 DNA (open triangles). Numbers in brackets indicate number of mice with tumors/total number of mice in group. One of two similar experiments is shown.



PICOGRAMS DNA



Tigure 1



ligare 19

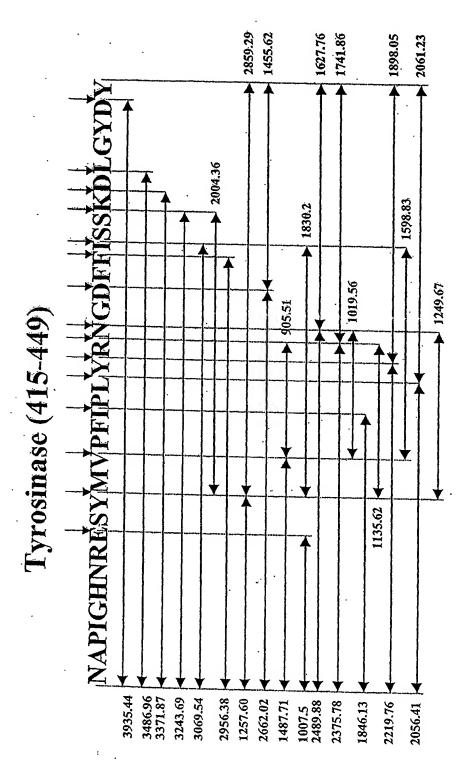


Figure 20

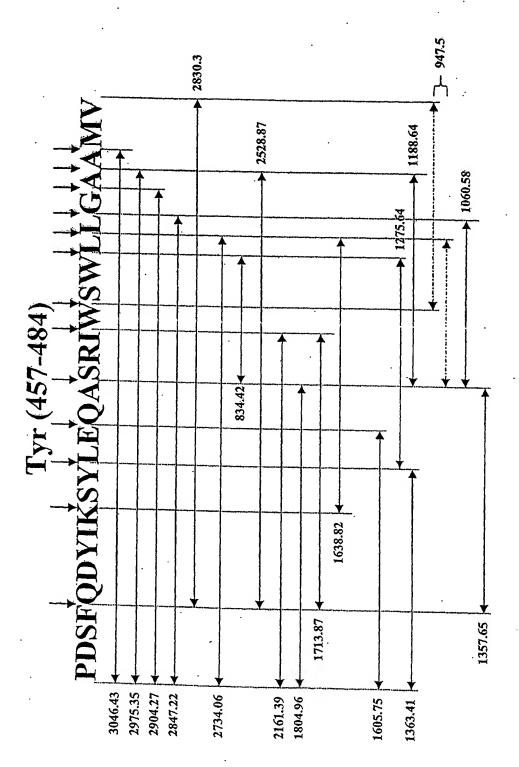
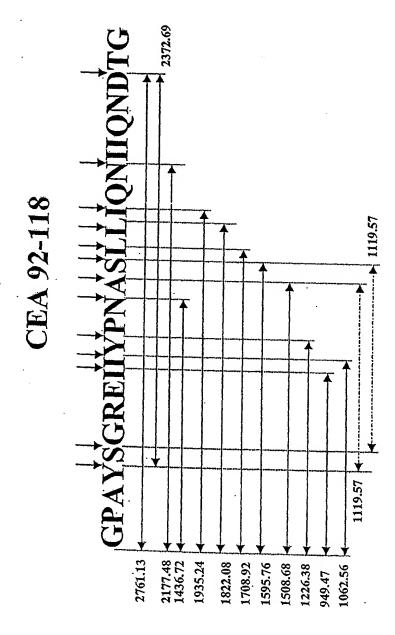


figure 21



rigare 7

▶ 941.47 2176.45 2263.53

Figure 23

CEA 225-251

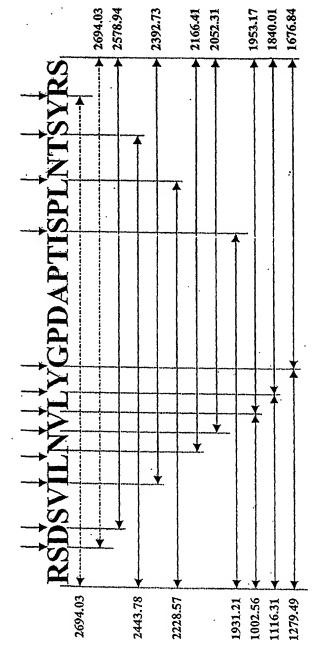
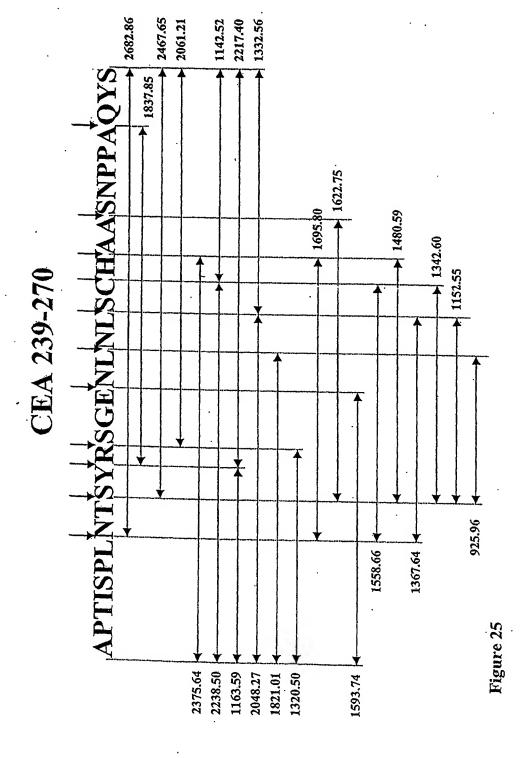


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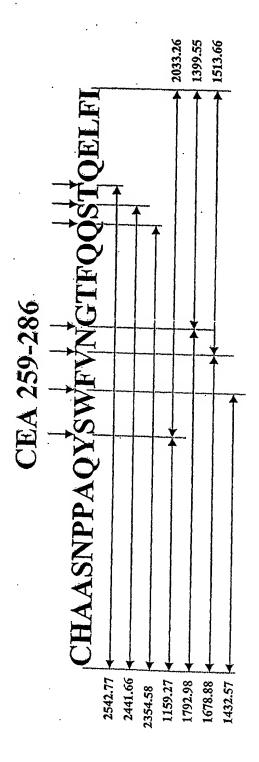


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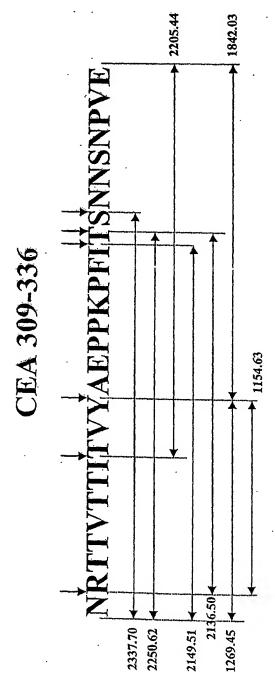


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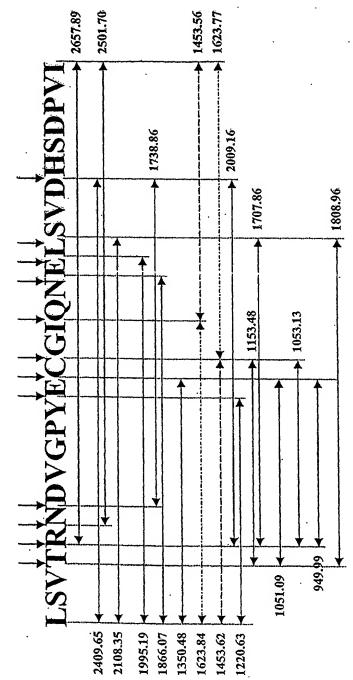


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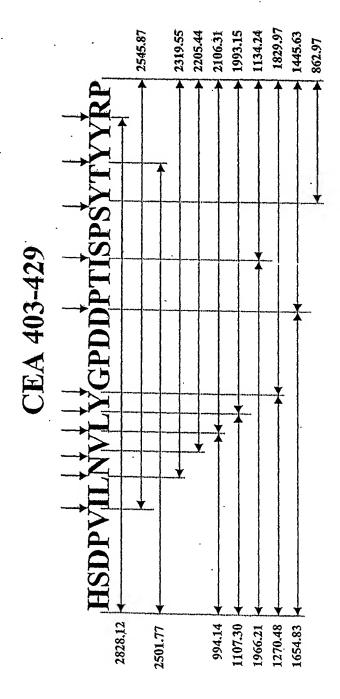


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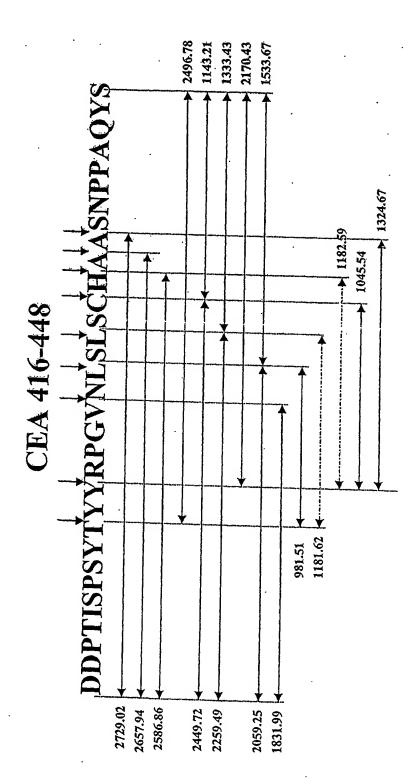


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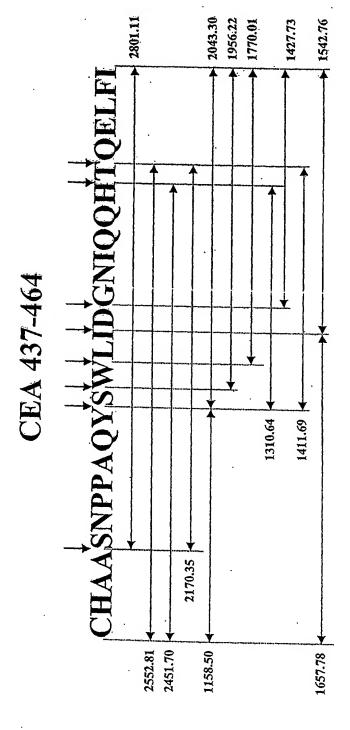


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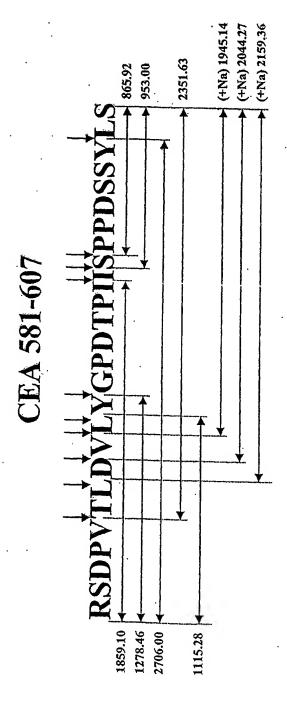


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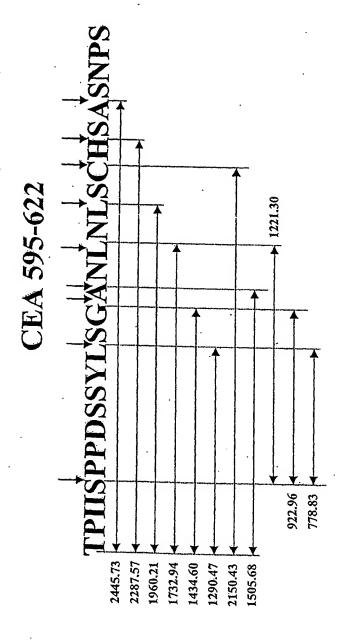


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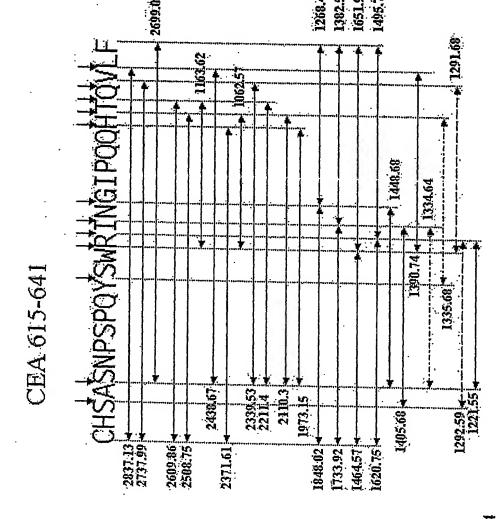


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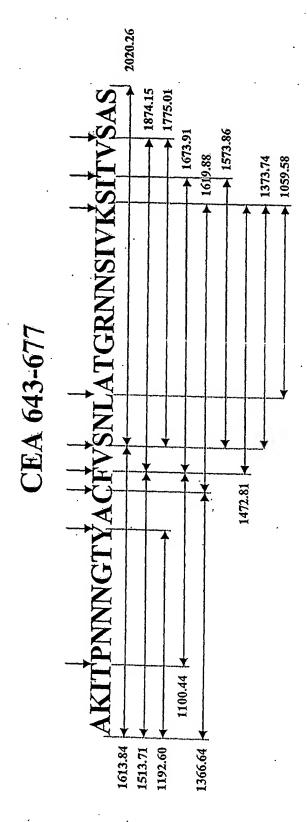
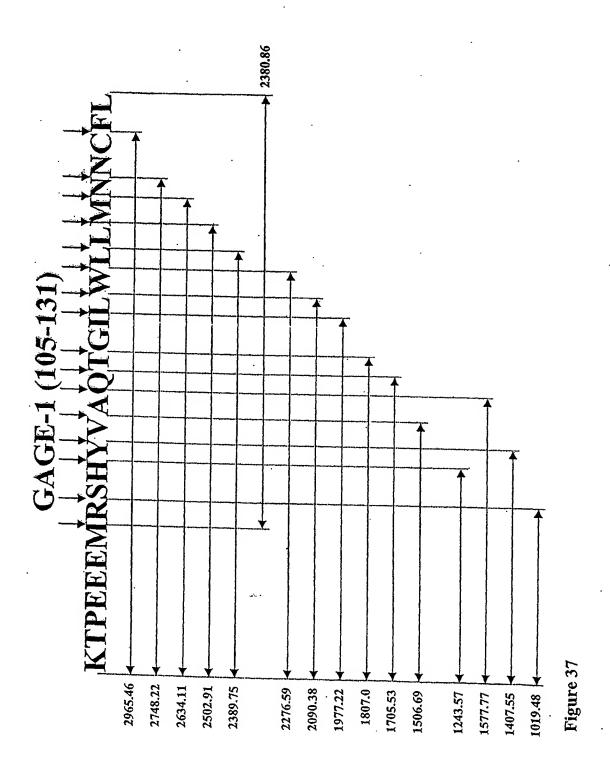


Figure 35

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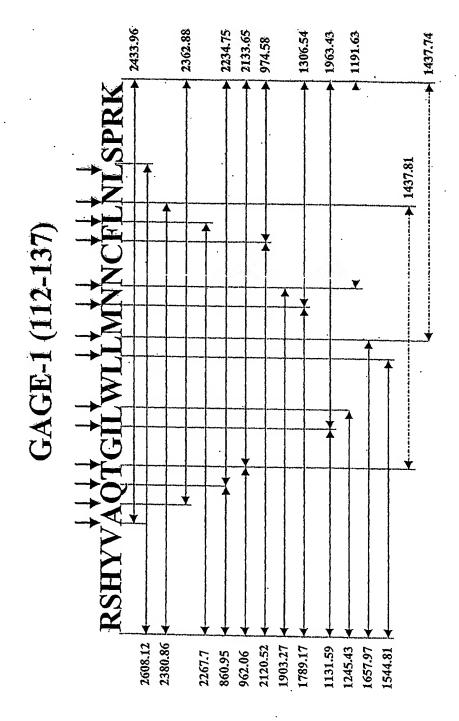


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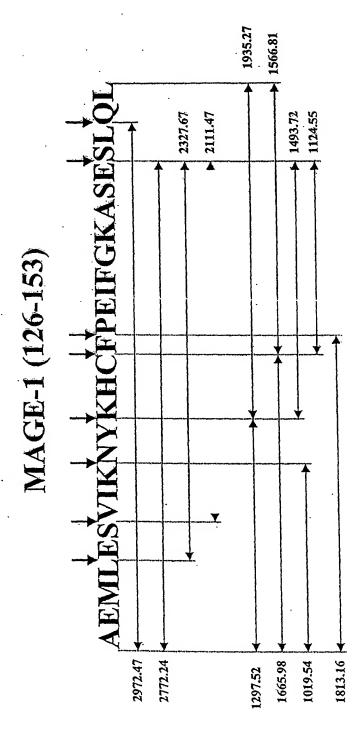
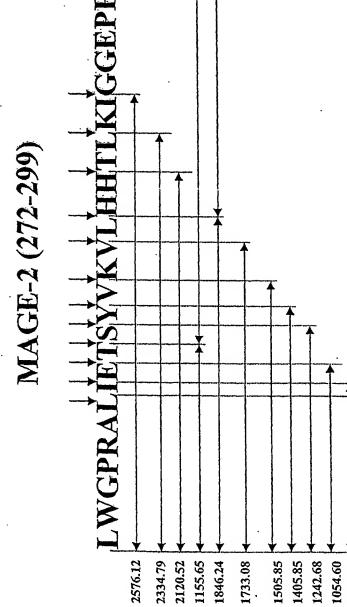


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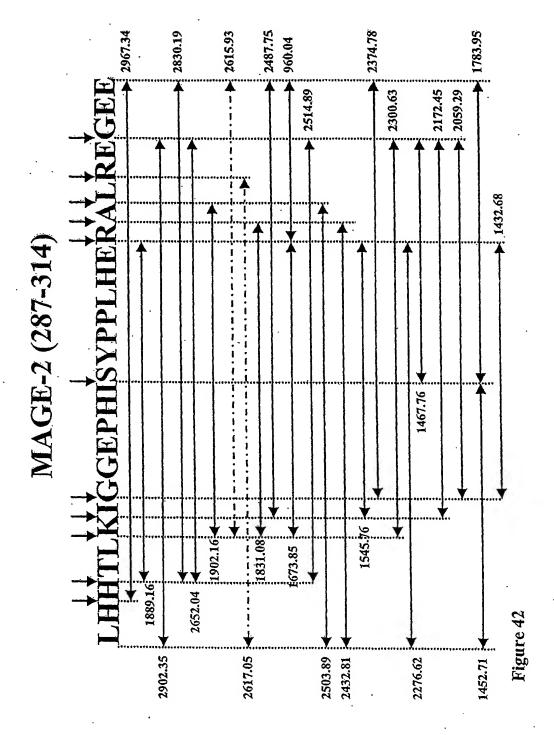
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Figure 41



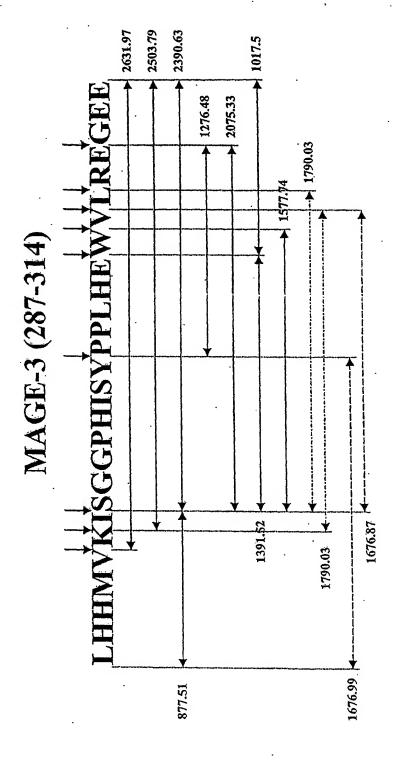


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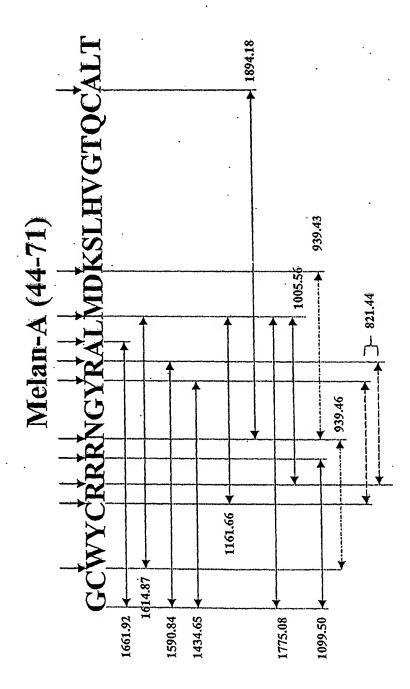


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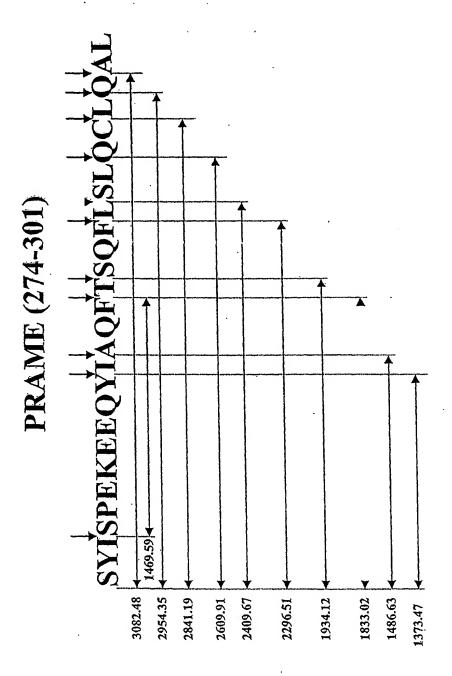
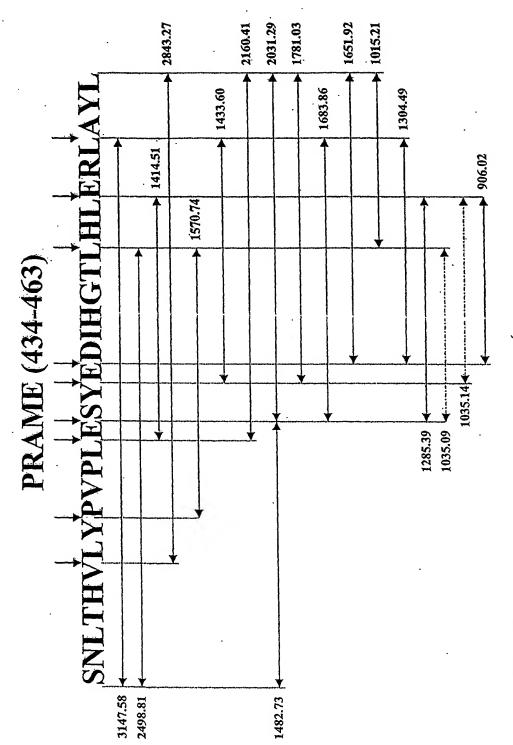


Figure 4



Pipure 46

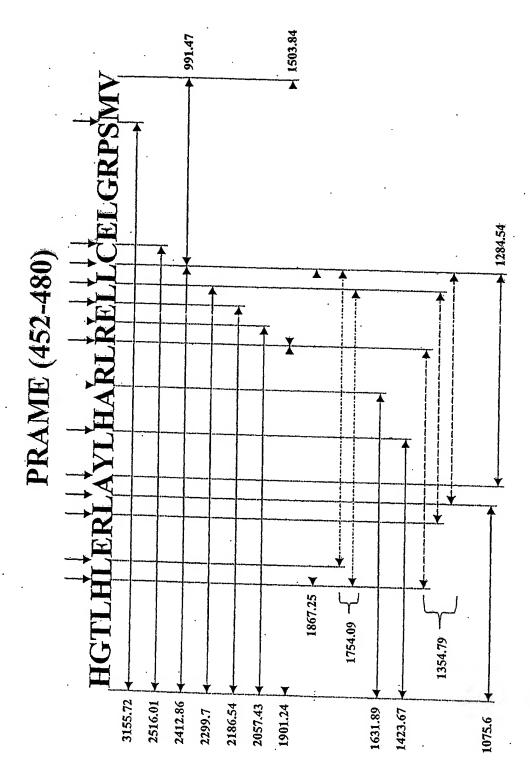
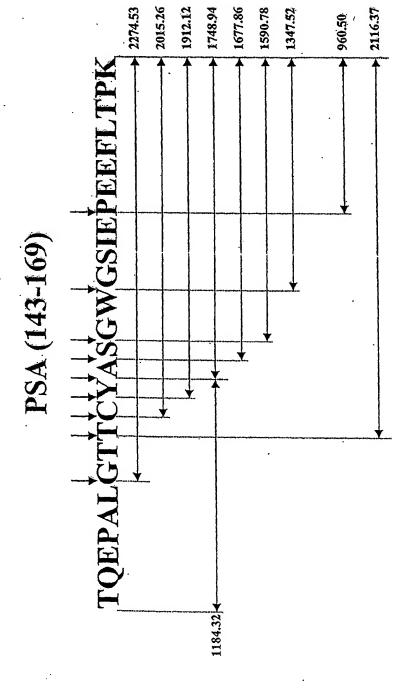


Figure 47



igure 4

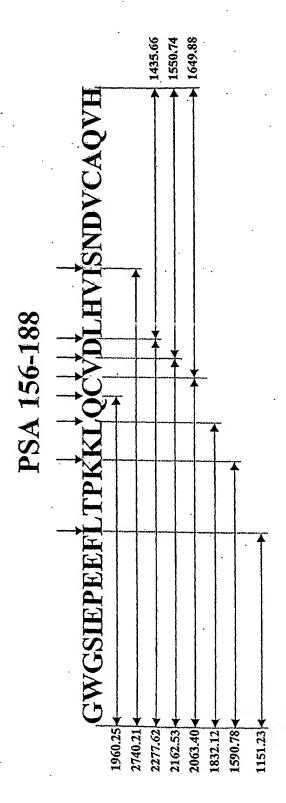


Figure 4

PSCA 67-94

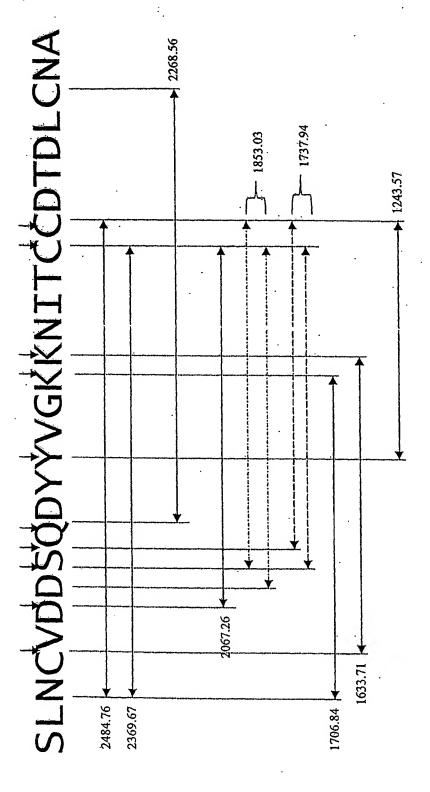


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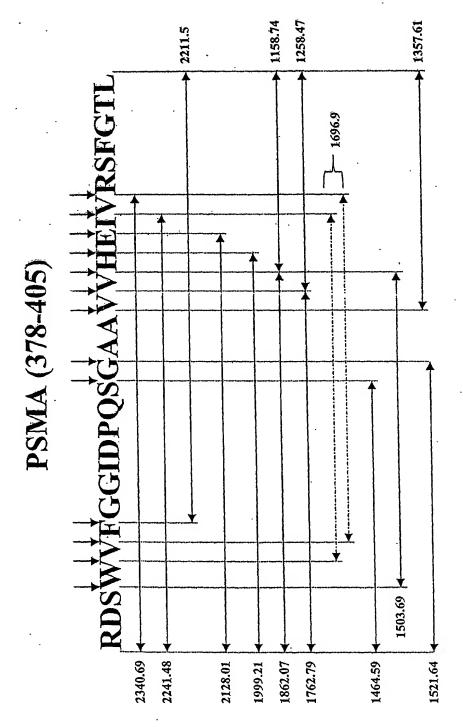


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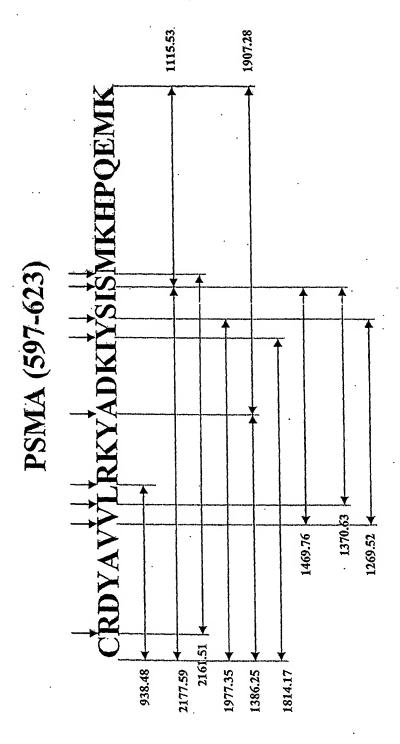


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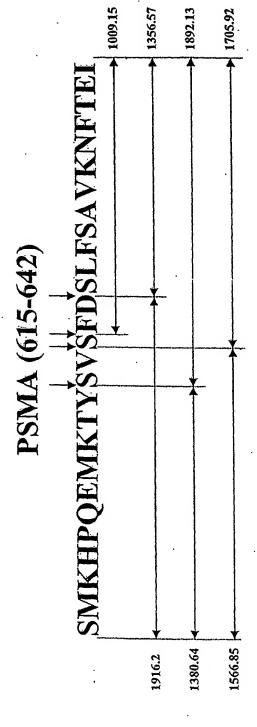


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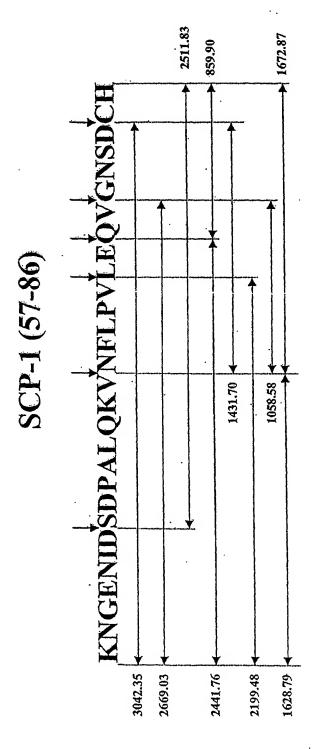


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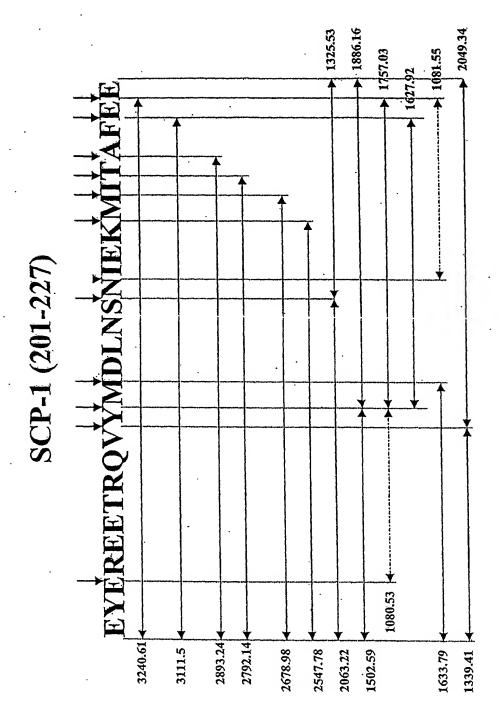


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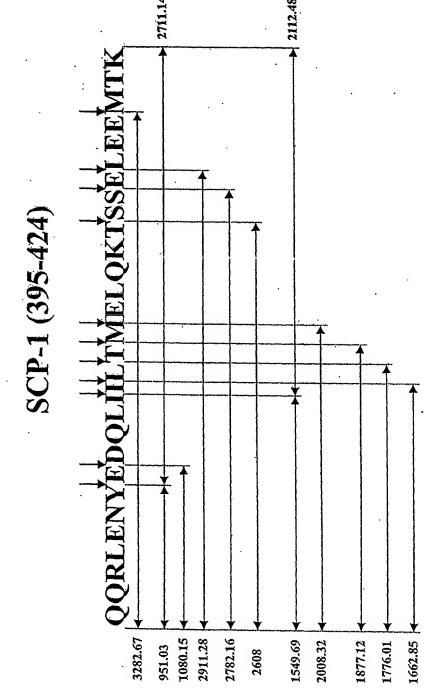


Figure 56

SCP-1 (416-442)

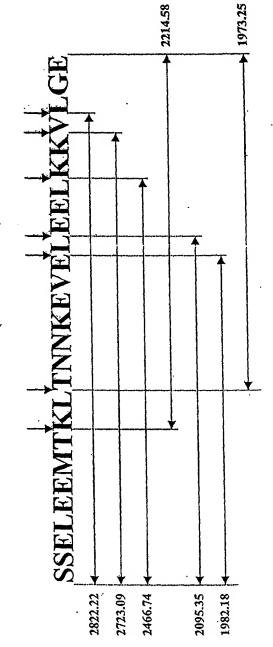


Figure 5

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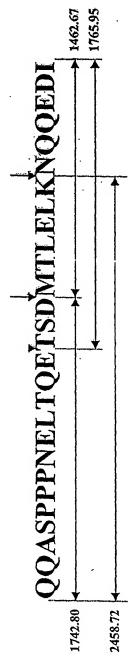


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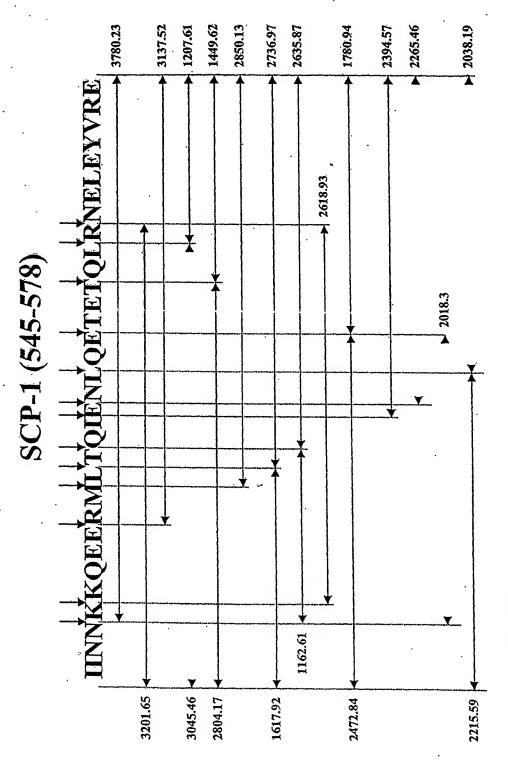


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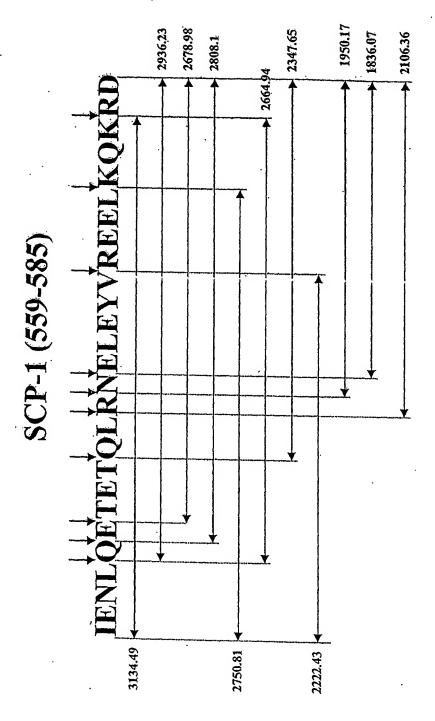


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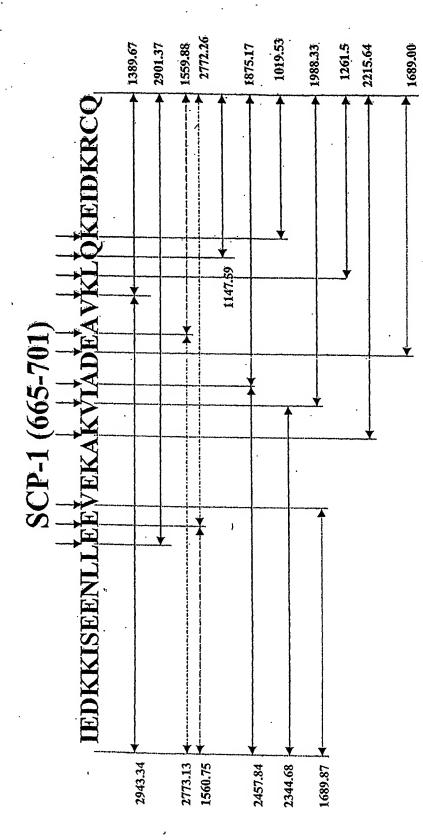


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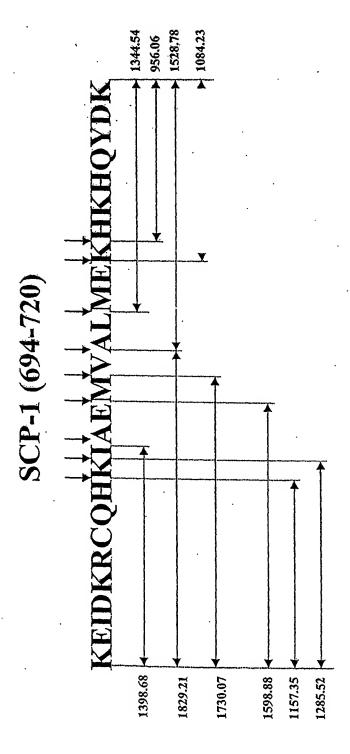
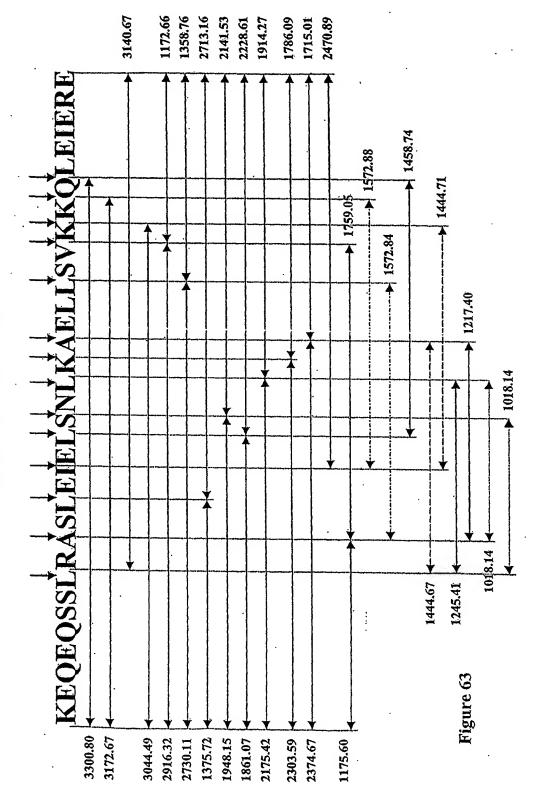


Figure 6

SCP-1 735-769



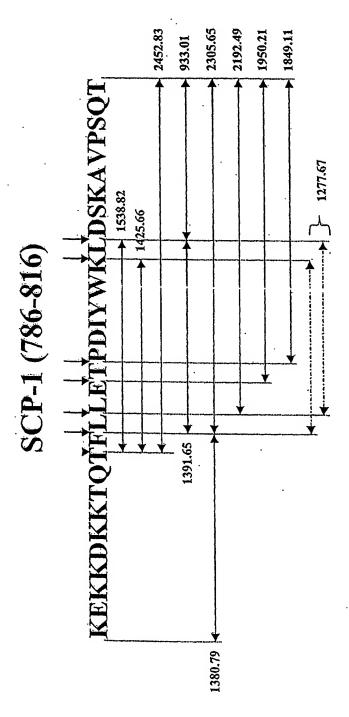


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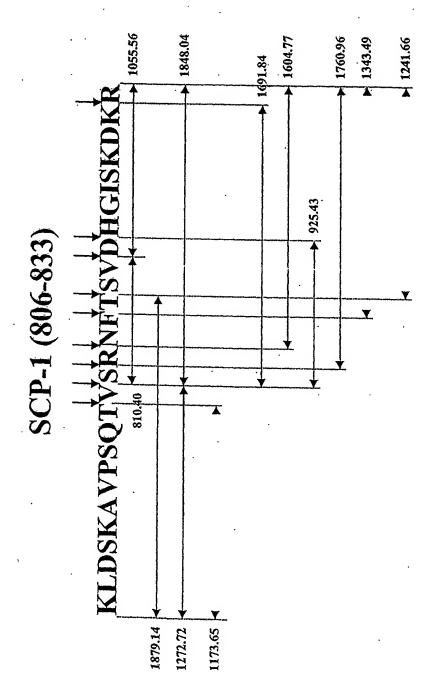


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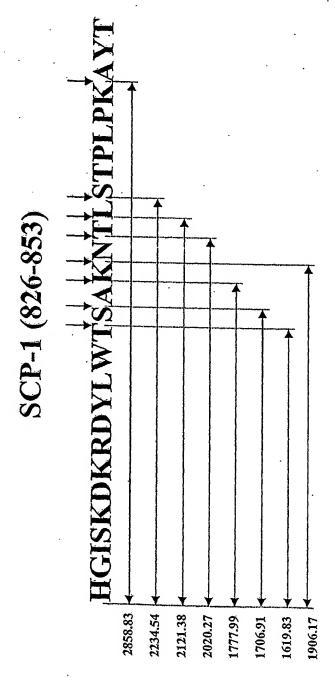


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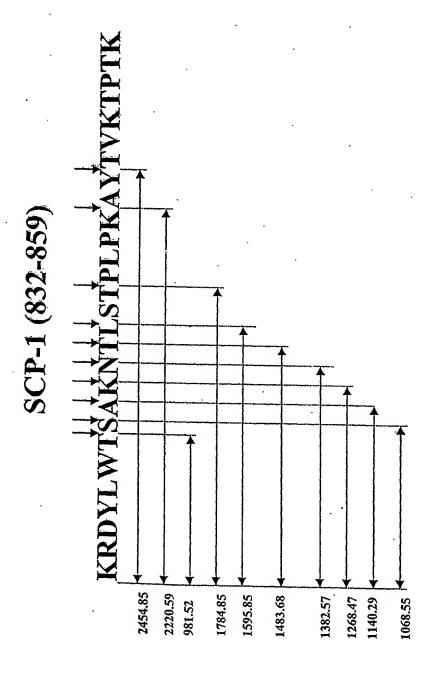


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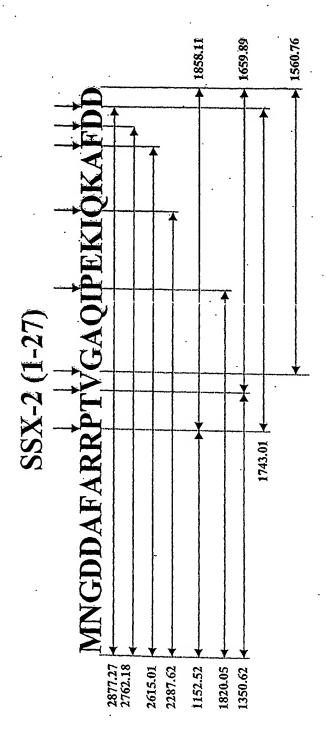


Figure 6

956.55

Survivin (116-142)

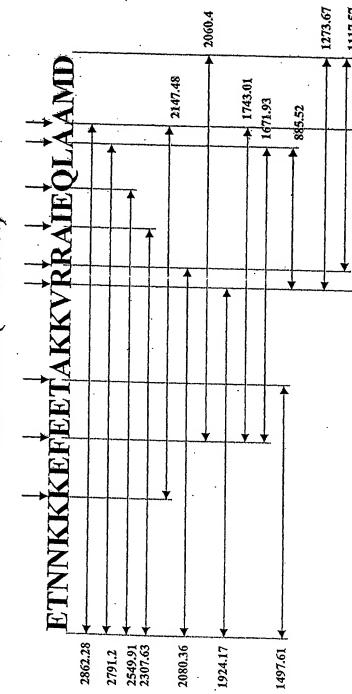


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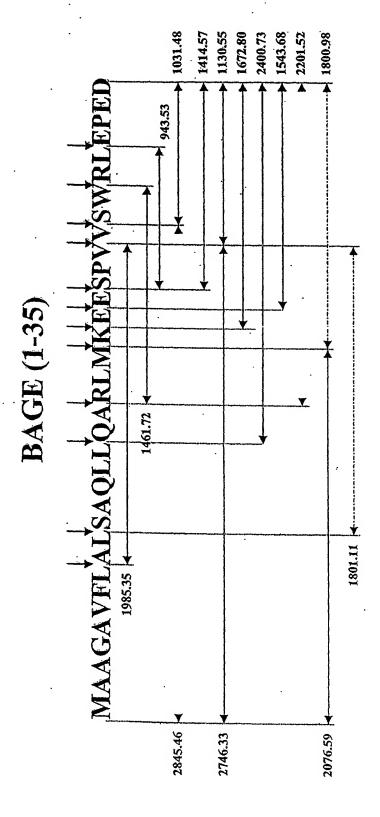


Figure 70

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Tyr Val Asn Tyr Ala Arg Thr Glu Asp Phe
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 Tyr Ala Arg Thr Glu Asp Phe Phe
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Ile Pro Val His Pro Ile Gly Tyr
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Tyr Tyr Asp Ala Gln Lys Leu Leu Glu
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Glu Gly Asn Tyr Thr Leu Arg Val
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Pro Phe Tyr
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Arg Met Met Asn Asp Gln Leu Met Phe Leu
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Tyr Pro Glu Trp Thr Glu Ala Gln Arg Leu Asp Cys Trp Arg Gly Gly
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Gln Val Ser Leu Lys Val Ser Asn Asp Gly Pro Thr Leu Ile Gly Ala
Asn Ala Ser Phe Ser Ile Ala Leu Asn Phe Pro Gly Ser Gln Lys Val
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Leu Pro Asp Gly Gln Val Ile Trp Val Asn Asn Thr Ile Ile Asn Gly
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Ser Gln Val Trp Gly Gly Gln Pro Val Tyr Pro Gln Glu Thr Asp Asp
                           120
Ala Cys Ile Phe Pro Asp Gly Gly Pro Cys Pro Ser Gly Ser Trp Ser
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Gln Lys Arg Ser Phe Val Tyr Val Trp Lys Thr Trp Gly Gln Tyr Trp
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Gln Val Leu Gly Gly Pro Val Ser Gly Leu Ser Ile Gly Thr Gly Arg
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                                 170
Ala Met Leu Gly Thr His Thr Met Glu Val Thr Val Tyr His Arg Arg
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                              185
                                                 190
Gly Ser Arg Ser Tyr Val Pro Leu Ala His Ser Ser Ser Ala Phe Thr
                         200
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Ile Thr Asp Gln Val Pro Phe Ser Val Ser Val Ser Gln Leu Arg Ala
                      215
Leu Asp Gly Gly Asn Lys His Phe Leu Arg Asn Gln Pro Leu Thr Phe
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                                     235
Ala Leu Gln Leu His Asp Pro Ser Gly Tyr Leu Ala Glu Ala Asp Leu
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Ser Tyr Thr Trp Asp Phe Gly Asp Ser Ser Gly Thr Leu Ile Ser Arg
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Ala Pro Val Val Thr His Thr Tyr Leu Glu Pro Gly Pro Val Thr Ala
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Val Pro Thr Thr Glu Val Ile Ser Thr Ala Pro Val Gln Met Pro Thr
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Ala Glu Ser Thr Gly Met Thr Pro Glu Lys Val Pro Val Ser Glu Val
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Met Gly Thr Thr Leu Ala Glu Met Ser Thr Pro Glu Ala Thr Gly Met
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Thr Pro Ala Glu Val Ser Ile Val Val Leu Ser Gly Thr Thr Ala Ala
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Gln Val Thr Thr Glu Trp Val Glu Thr Thr Ala Arg Glu Leu Pro
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Arg Leu Val Lys Arg Gln Val Pro Leu Asp Cys Val Leu Tyr Arg Tyr
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Gly Ser Phe Ser Val Thr Leu Asp Ile Val Gln Gly Ile Glu Ser Ala
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Thr Val Ser Cys Gln Gly Gly Leu Pro Lys Glu Ala Cys Met Glu Ile
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Ser Ser Pro Gly Cys Gln Pro Pro Ala Gln Arg Leu Cys Gln Pro Val
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Leu Pro Ser Pro Ala Cys Gln Leu Val Leu His Gln Ile Leu Lys Gly
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Gly Ser Gly Thr Tyr Cys Leu Asn Val Ser Leu Ala Asp Thr Asn Ser
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Leu Gly Gln Val Pro Leu Ile Val Gly Ile Leu Leu Val Leu Met Ala
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                                605
Val Val Leu Ala Ser Leu Ile Tyr Arg Arg Arg Leu Met Lys Gln Asp
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Phe Ser Val Pro Gln Leu Pro His Ser Ser Ser His Trp Leu Arg Leu
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Ser Gly Gln Gln Val
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Leu Phe Arg Ala Val Ile Thr Lys Lys Val Ala Asp Leu Val Gly Phe
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Leu Leu Leu Lys Tyr Arg Ala Arg Glu Pro Val Thr Lys Ala Glu Met
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Leu Glu Ser Val Ile Lys Asn Tyr Lys His Cys Phe Pro Glu Ile Phe
 130 135
Gly Lys Ala Ser Glu Ser Leu Gln Leu Val Phe Gly Ile Asp Val Lys
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Glu Ala Asp Pro Thr Gly His Ser Tyr Val Leu Val Thr Cys Leu Gly
                              170
            165
Leu Ser Tyr Asp Gly Leu Leu Gly Asp Asn Gln Ile Met Pro Lys Thr
                                             190
                           185
Gly Phe Leu Ile Ile Val Leu Val Met Ile Ala Met Glu Gly Gly His
                                          205
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Ala Pro Glu Glu Glu Ile Trp Glu Glu Leu Ser Val Met Glu Val Tyr
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Asp Gly Arg Glu His Ser Ala Tyr Gly Glu Pro Arg Lys Leu Leu Thr
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Gln Asp Leu Val Gln Glu Lys Tyr Leu Glu Tyr Arg Gln Val Pro Asp
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Ser Asp Pro Ala Arg Tyr Glu Phe Leu Trp Gly Pro Arg Ala Leu Ala
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Glu Thr Ser Tyr Val Lys Val Leu Glu Tyr Val Ile Lys Val Ser Ala
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Asp Phe Phe Pro Val Ile Phe Ser Lys Ala Ser Glu Tyr Leu Gln Leu 150 Val Phe Gly Ile Glu Val Val Glu Val Val Pro Ile Ser His Leu Tyr 175 170 Ile Leu Val Thr Cys Leu Gly Leu Ser Tyr Asp Gly Leu Leu Gly Asp 185 180 Asn Gln Val Met Pro Lys Thr Gly Leu Leu Ile Ile Val Leu Ala Ile 205 200 195 Ile Ala Ile Glu Gly Asp Cys Ala Pro Glu Glu Lys Ile Trp Glu Glu 215 Leu Ser Met Leu Glu Val Phe Glu Gly Arg Glu Asp Ser Val Phe Ala 235 230 His Pro Arg Lys Leu Leu Met Gln Asp Leu Val Gln Glu Asn Tyr Leu 245 250 255 Glu Tyr Arg Gln Val Pro Gly Ser Asp Pro Ala Cys Tyr Glu Phe Leu 270 265 Trp Gly Pro Arg Ala Leu Ile Glu Thr Ser Tyr Val Lys Val Leu His 280 285 275 His Thr Leu Lys Ile Gly Gly Glu Pro His Ile Ser Tyr Pro Pro Leu 295 His Glu Arg Ala Leu Arg Glu Gly Glu Glu

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85

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Phe Lys Ala Val Leu Asp Gly Leu Asp Val Leu Leu Ala Gln Glu Val
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Ser Lys Lys Glu Trp Glu Lys Met Lys Ser Ser Glu Lys Ile Val Tyr
                          40
Val Tyr Met Lys Leu Asn Tyr Glu Val Met Thr Lys Leu Gly Phe Lys
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                                           60
Val Thr Leu Pro Pro Phe Met Arg Ser Lys Arg Ala Ala Asp Phe His
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Gly Asn Asp Phe Gly Asn Asp Arg Asn His Arg Asn Gln Val Glu Arg
                                   90
Pro Gln Met Thr Phe Gly Ser Leu Gln Arg Ile Phe Pro Lys Ile Met
            100
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Pro Lys Lys Pro Ala Glu Glu Asn Gly Leu Lys Glu Val Pro Glu
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                            120
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Ala Ser Gly Pro Gln Asn Asp Gly Lys Gln Leu Cys Pro Pro Gly Asn
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Pro Ser Thr Leu Glu Lys Ile Asn Lys Thr Ser Gly Pro Lys Arg Gly
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Lys His Ala Trp Thr His Arg Leu Arg Glu Arg Lys Gln Leu Val Val
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Tyr Glu Glu Ile Ser Asp Pro Glu Glu Asp Asp Glu
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                            40
Gly Gln Glu Met Asp Pro Pro Asn Pro Glu Glu Val Lys Thr Pro Glu
                        55
                                            60
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<211> 98
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<213> Homo sapiens
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            20
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His Lys Lys His Ser Ser Gly Cys Ala Phe Leu Ser Val Lys Lys Gln
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                            40
                                                45
Phe Glu Glu Leu Thr Leu Gly Glu Phe Leu Lys Leu Asp Arg Glu Arg
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Ala Lys Asn Lys Ile Ala Lys Glu Thr Asn Asn Lys Lys Glu Phe
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Met Asp

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gtgccaccag ccttcctgtg ggccccttag caatgtctta ggaaaggaga tcaacatttt 600
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<213> Homo sapiens
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Glu Gly Phe Asp His Arg Asp Ser Lys Val Ser Leu Gln Glu Lys Asn
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Cys Glu Pro Val Val Pro Asn Ala Pro Pro Ala Tyr Glu Lys Leu Ser
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Ala Glu Gln Ser Pro Pro Pro Tyr Ser Pro
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          20
                               25
Glu Gln His Ser Gln Pro Trp Gln Ala Ala Leu Tyr His Phe Ser Thr
       35
                           40
                                              45
Phe Gln Cys Gly Gly Ile Leu Val His Arg Gln Trp Val Leu Thr Ala
                       55
                                          60
Ala His Cys Ile Ser Asp Asn Tyr Gln Leu Trp Leu Gly Arg His Asn
                   70
                                       75
Leu Phe Asp Asp Glu Asn Thr Ala Gln Phe Val His Val Ser Glu Ser
               85
                                   90
Phe Pro His Pro Gly Phe Asn Met Ser Leu Leu Glu Asn His Thr Arg
           100
                               105
                                                  110
Gln Ala Asp Glu Asp Tyr Ser His Asp Leu Met Leu Leu Arg Leu Thr
                                             125
      115
                        120
Glu Pro Ala Asp Thr Ile Thr Asp Ala Val Lys Val Val Glu Leu Pro
                      135
                                        140
Thr Gln Glu Pro Glu Val Gly Ser Thr Cys Leu Ala Ser Gly Trp Gly
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                                      155
Ser Ile Glu Pro Glu Asn Phe Ser Phe Pro Asp Asp Leu Gln Cys Val
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                                   170
                                                       175
Asp Leu Lys Ile Leu Pro Asn Asp Glu Cys Glu Lys Ala His Val Gln
                               185
                                                  190
Lys Val Thr Asp Phe Met Leu Cys Val Gly His Leu Glu Gly Gly Lys
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Asp Thr Cys Val Gly Asp Ser Gly Gly Pro Leu Met Cys Asp Gly Val
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                                          220
Leu Gln Gly Val Thr Ser Trp Gly Tyr Val Pro Cys Gly Thr Pro Asn
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Lys Pro Ser Val Ala Val Arg Val Leu Ser Tyr Val Lys Trp Ile Glu
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Asp Thr Ile Ala Glu Asn Ser
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Gly Glu Glu Ala Arg Pro Asn Ser Trp Pro Trp Gln Val Ser Leu Gln
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Tyr Ser Ser Asn Gly Lys Trp Tyr His Thr Cys Gly Gly Ser Leu Ile
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                              60
Ala Asn Ser Trp Val Leu Thr Ala Ala His Cys Ile Ser Ser Arg
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                                 75
Thr Tyr Arg Val Gly Leu Gly Arg His Asn Leu Tyr Val Ala Glu Ser
           85
                             90
Gly Ser Leu Ala Val Ser Val Ser Lys Ile Val Val His Lys Asp Trp
         100
                          105
Asn Ser Asn Gln Ile Ser Lys Gly Asn Asp Ile Ala Leu Leu Lys Leu
                       120
                                        125
Ala Asn Pro Val Ser Leu Thr Asp Lys Ile Gln Leu Ala Cys Leu Pro
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Pro Ala Gly Thr Ile Leu Pro Asn Asn Tyr Pro Cys Tyr Val Thr Gly
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Trp Gly Arg Leu Gln Thr Asn Gly Ala Val Pro Asp Val Leu Gln Gln
          165 170 175
Gly Arg Leu Leu Val Val Asp Tyr Ala Thr Cys Ser Ser Ser Ala Trp
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Trp Gly Ser Ser Val Lys Thr Ser Met Ile Cys Ala Gly Gly Asp Gly
      195 200
                                         205
Val Ile Ser Ser Cys Asn Gly Asp Ser Gly Gly Pro Leu Asn Cys Gln
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Ala Ser Asp Gly Arg Trp Gln Val His Gly Ile Val Ser Phe Gly Ser
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Arg Leu Gly Cys Asn Tyr Tyr His Lys Pro Ser Val Phe Thr Arg Val
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<211> 270

<212> PRT

<213> Homo sapiens

<400> 107

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Trp Gly Arg Leu Gln Thr Asn Gly Ala Leu Pro Asp Asp Leu Lys Gln
    165 170 175
Gly Arg Leu Leu Val Val Asp Tyr Ala Thr Cys Ser Ser Ser Gly Trp
              185
                                          190
 . 180
Trp Gly Ser Thr Val Lys Thr Asn Met Ile Cys Ala Gly Gly Asp Gly
   195 200 205
Val Ile Cys Thr Cys Asn Gly Asp Ser Gly Gly Pro Leu Asn Cys Gln 210 215 220
Ala Ser Asp Gly Arg Trp Glu Val His Gly Ile Gly Ser Leu Thr Ser 225 230 235 240
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(57) Abstract: Disclosed herein are polypeptides, including epitopes, clusters, and antigens. Also disclosed are compositions that include said polypeptides and methods for their use.



INTERNATIONAL SEARCH REPORT

International application No.

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A. CLASSIFICATION OF SUBJECT MATTER					
IPC(7) : A61K 39/395, 39/00, 38/03, 48/00; A01N 63/00; C12N 15/12, 15/63 US CL : 424/184.1, 185,1, 93.71; 435/320.1; 514/2, 44; 530/300; 536/23.1					
According to International Patent Classification (IPC) or to both national classification and IPC					
B. FIEL	DS SEARCHED				
Minimum documentation searched (classification system followed by classification symbols) U.S.: 424/184.1, 185,1, 93.71; 435/320.1; 514/2, 44; 530/300; 536/23.1					
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched					
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) Please See Continuation Sheet					
C. DOCUMENTS CONSIDERED TO BE RELEVANT					
Category *	Citation of document, with indication, where a	ppropriate, of the relevant passages	Relevant to claim No.		
Y	US 6,037,135 A (KUBO et al.) 14 March 2000 (14	1.03.2000), see entire document.	1-82		
Y	WO 94/020127 A1 (CYTEL CORPOARTION) 15 entire document.	September 1994 (15.09.1994), see	1-82		
Further documents are listed in the continuation of Box C. See patent family annex.					
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Date of the actual completion of the international search		Date of mailing of the international sear	rch report		
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Name and mailing address of the ISA/US Mail Stop PCT, Attu: ISA/US Commissioner for Patents P.O. Box 1450 Alexandea, Virginia 22313-1450		Ron Schwadron, Ph.D. Z. D. R. for Telephone No. 571-272-1600			
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Continuation of B. FIELDS SEARCHED Item 3: WEST 2.1, MEDICINE/BIOTECH (compendium databases on DIALOG) search peptid?, mhc	terms: inventor names, mage?, cea?, gage?, hla?,
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